

1 Extreme value distributions describe interannual variability in the North 2 Atlantic spring bloom

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9 **Scientific Significance Statement:** The North Atlantic spring bloom helps fuel the marine food
10 web, impacts fisheries recruitment, contributes to carbon export, and is predicted to change with
11 climate warming. Understanding interannual bloom variability is thus of central oceanographic
12 importance. This study provides a framework for quantifying interannual bloom variability via
13 statistical extreme value theory. We characterize the spatial distribution of extreme value
14 parameters using satellite chlorophyll observations and test whether the distribution of
15 chlorophyll extremes has changed over time in relation to trends in background chlorophyll
16 levels and sea surface temperature.

17 18 **Abstract**

19
20 The North Atlantic spring bloom depends on a confluence of environmental factors that drive
21 transient periods of exponential phytoplankton growth and interannual variability in bloom
22 magnitude. We analyze interannual bloom variability in the North Atlantic via extreme value
23 theory where the Generalized Extreme Value Distribution (GEVD) is fitted spatially to annual
24 maxima of satellite-measured surface chlorophyll. We find excellent agreement between the
25 observed distribution of interannual bloom maxima and those predicted from the GEVD. The
26 spatial distribution of fitted GEVD parameters closely follows basin bathymetry where the
27 largest extremes and heaviest distribution tails are found on the continental shelves and slopes.
28 Trend analyses suggest weak evidence for changes in GEVD parameters, despite regional trends
29 in mean chlorophyll levels and sea surface temperature. These results provide a framework to
30 quantify interannual bloom variability and call for further work examining how extreme blooms
31 propagate through food webs and contribute to carbon export.

32 33 **Introduction**

34
35 Phytoplankton form the base of the marine food web (Falkowski et al. 2003) and play a major
36 role in the global carbon cycle (Falkowski et al. 1998; Ito and Follows 2005). At mid and high-
37 latitudes, the seasonality of phytoplankton is characterized by a late-winter or spring bloom
38 where phytoplankton biomass concentrations reach an annual maximum (Behrenfeld and Boss
39 2014). The spring bloom is important for sustaining higher trophic levels, including species
40 whose annual migrations are timed to coincide with the bloom (Visser et al. 2011). Blooms also
41 contribute to carbon export and are often associated with large export pulses to depth (Briggs et
42 al. 2011).

43
44 Several biophysical mechanisms are thought to control bloom development (Behrenfeld and
45 Boss 2014, 2018). For example, the critical depth hypothesis predicts that bloom initiation begins
46 with mixed layer shoaling and associated increases in mixed layer averaged light availability

47 (Behrenfeld and Boss 2014, 2018). The critical turbulence hypothesis (a variant of the critical
48 depth hypothesis) focuses on light availability in the active mixing layer (Taylor and Ferrari
49 2011; Ferrari et al. 2015), while the dilution-recovery hypothesis predicts that blooms develop
50 due to changes in grazing via dilution from deepening mixed layers (Evans et al. 1985;
51 Behrenfeld 2010). Across proposed mechanisms, the population-level phenomena of blooms
52 arise due to transient positive imbalances between growth and loss rates such that the exponential
53 growth rate is positive, i.e.

54

$$55 \quad \frac{dc}{dt} = (g(t) - l(t))c,$$

56

57 with $g(t) - l(t) > 0$ prior to the bloom. Days to weeks post-bloom, losses increase to match
58 and exceed growth, most often because of increased grazer abundance supported by the elevated
59 phytoplankton biomass (Behrenfeld and Boss 2014, 2018). The magnitude and duration of the
60 transient exponential growth period determines the magnitude of the spring bloom biomass
61 maximum.

62

63 The North Atlantic basin exhibits one of the largest and well-studied seasonal blooms across the
64 global ocean. Deep winter mixing and rapid springtime re-stratification causes ideal conditions
65 for exponential phytoplankton growth, with bloom timing following a northward progression as
66 re-stratification occurs earliest at low latitude (Dutkiewicz et al. 2001; Siegel et al. 2002).
67 Meteorological variability then modulates the precise timing and magnitude of the bloom
68 according to the impacts on local mixing dynamics (Dutkiewicz et al. 2001; Follows and
69 Dutkiewicz 2002). Importantly, variability in bloom timing and magnitude has been linked to the
70 magnitude of carbon export (Briggs et al. 2011) and to fisheries productivity via the ‘match-
71 mismatch hypothesis’ (Platt et al. 2003). Climate change is expected to alter North Atlantic
72 bloom dynamics via a range of factors, including changes in seasonal mixing depths, nutrient
73 fluxes, and the metabolic impacts of warmer temperatures (Sommer and Lengfellner 2008).
74 Quantifying the dynamics of the North Atlantic spring bloom is thus of central importance for
75 understanding the relevant oceanographic and ecological processes and will aid in tracking the
76 associated impacts of climate change.

77

78 Viewing interannual blooms as extreme values in observed phytoplankton time series brings to
79 bear the statistical theory of extreme values. Under the Fisher-Tippet-Gnedenko theorem, the
80 maximum of a sequences of random variables converges in distribution to the Generalized
81 Extreme Value Distribution (GEVD), itself a generalization of the Gumbel, Frechet, and Weibull
82 distributions (Coles 2001). The theorem yields a three-parameter probability density function
83 describing the limiting distribution of maximum values generated from samples of a stochastic
84 process, analogous to the central limit theorem for a the mean of a distribution (Coles 2001;
85 described below). While the GEVD is increasingly applied to geophysical and climate studies
86 (Easterling et al. 2000; Katz 2010; Aghakouchak et al. 2020). there have been fewer applications
87 to biological time series (Batt et al. 2017). Consistent with the exponential nature of biological
88 growth, Batt et al. (2017) found that biological time series exhibited consistently heavier tails in
89 their extreme value distributions relative to chemical and geophysical time series. These findings
90 suggest the North Atlantic bloom as an ideal target of extreme value analysis, due to its annually
91 repeating cycle of transient and variable exponential growth.

92

93 Here we analyzed the North Atlantic satellite chlorophyll record to quantify seasonal
94 phytoplankton bloom variability via extreme value analysis. We estimated GEVD parameters at
95 a $\frac{1}{4}^\circ$ latitude-longitude scale. We mapped the fitted parameters spatially and evaluated the
96 GEVD goodness-of-fit to the chlorophyll time series. We correlated the fitted parameters to
97 bathymetric properties of the North Atlantic basin. We further evaluated evidence for non-
98 stationarity (i.e., time-variability) in GEVD parameters in the context of satellite-observed
99 chlorophyll and temperature trends across the basin. Results of this study will provide a
100 statistical framework to describe interannual bloom variability and allow us to test an important
101 hypothesis with respect to basin-scale environmental change.

102

103 **Methods**

104

105 *Observations*

106

107 We analyzed two sets of basin-scale satellite chlorophyll observations. First, we used chlorophyll
108 estimates from the Moderate-resolution Imaging Spectroradiometer-Aqua (MODIS-Aqua)
109 sensor, spanning years 2002-2021, accessed from the Oregon State University Ocean
110 Productivity database (<https://sites.science.oregonstate.edu/ocean.productivity/>). Associated
111 inherent optical properties were estimated using the Garver-Siegel-Maritorena (GSM) algorithm
112 (Maritorena and Siegel 2005). MODIS-Aqua based chlorophyll estimates were gap-filled for
113 missing observations due to clouds according to the algorithm described at
114 http://orca.science.oregonstate.edu/gap_fill.php. While gap-filling can alter the underlying
115 chlorophyll distribution, it is not expected to affect the annually measured maximum value.
116 Secondly, we used the Ocean Colour Climate Change Initiative (OC-CCI) chlorophyll product
117 (Sathyendranath et al. 2019) covering the same time period as the MODIS-Aqua dataset. OC-
118 CCI chlorophyll is a synthetic product generated by combining information from multiple
119 sensors, including SeaWiFS (Sea-viewing Wide-Field-of-view Sensor), MODIS-Aqua, MERIS
120 (Medium spectral Resolution Imaging Spectrometer) and VIIRS (Visible and Infrared Imaging
121 Radiometer Suite). OC-CCI was not gap-filled and therefore contained missing values due to
122 clouds. We grided both sets of chlorophyll observations to $\frac{1}{4}^\circ$ latitude-longitude rectilinear
123 resolution. Wintertime satellite chlorophyll observations were not available for higher latitudes
124 due to light limitation and were replaced with a value of zero concentration which did not affect
125 the extreme value analysis. Sea surface temperature data from the MODIS-Aqua sensor were
126 taken from the Oregon State University Ocean Productivity Database site cited above.

127

128 *Extreme Value Analysis*

129

130 Given a time series of chlorophyll observations at an individual location, we estimated the
131 parameters of the GEVD via the block maxima approach (Gilleland and Katz 2016), taking time
132 series blocks as individual years. We define $x = \max(y_1, y_2, \dots, y_n)$ as the maximum chlorophyll
133 measurement in a single year of n measurements. Over m years we have m yearly maxima, thus
134 defining the observed annual maxima time series x_1, x_2, \dots, x_m . For an annual maximum x , the
135 GEVD has a probability density function given by

136

137

$$p(x) = \frac{1}{\sigma} t(x)^{\xi+1} e^{-t(x)}$$

138 with

139

$$t(x) = \begin{cases} \left(1 + \xi \left(\frac{x - \mu}{\sigma}\right)\right)^{-\frac{1}{\xi}}, & \xi \neq 0, \\ e^{-\frac{x - \mu}{\sigma}}, & \xi = 0, \end{cases}$$

140

141 where μ , σ , and ξ are the location, scale, and shape parameters, respectively. The location
142 parameter shifts the GEVD along the x axis, the scale parameter controls the spread, and the
143 shape parameter controls the peaked-ness of the mode and heaviness of the distribution tail.

144 Examples of how μ , σ , and ξ modulate the GEVD are given in **Figure 1**. Formulas for the
145 expected value, variance, and mode of the GEVD are given in **Appendix A**.

146

147 In addition to the tree-parameter GEVD described above, we also apply a nonstationary
148 extension where the parameters are described as simple linear functions of time (Gilleland and
149 Katz 2016) of the form

150

$$\theta(t) = \theta_0 + \theta_1 t,$$

151

152

153 where θ is one of the GEVD parameters, θ_0 is the intercept of the linear relationship, and θ_1 is
154 the slope, i.e. the rate of change with respect to time.

155

156 The log-likelihood for the GEVD parameters (Gilleland and Katz 2016) is given by

157

$$158 \quad l(\mu, \sigma, \xi | x_1, x_2, \dots, x_m) = -m \ln \sigma - \left(1 + \frac{1}{\xi}\right) \sum_{i=1}^m \ln \left(1 + \xi \left(\frac{x_i - \mu}{\sigma}\right)\right) - \sum_{i=1}^m \left(1 + \xi \left(\frac{x_i - \mu}{\sigma}\right)\right)^{1/\xi}.$$

159

160 We maximized the log-likelihood function with respect to the parameters to obtain empirical
161 estimates. We maximized with respect to μ , σ , and ξ in the case of stationary GEVDs, and with
162 respect to the intercept and slope in the case of nonstationary GEVDs. We restricted the analysis
163 to estimating one nonstationary parameter at a time due to data restrictions and weak
164 identifiability when multiple parameters are allowed to vary with time. We only considered
165 linear functions of time and suggest nonlinear functions for future work. We used numerical
166 optimization routines implemented in the extRemes library within the R programming language
167 (Gilleland and Katz 2016). We compared stationary and nonstationary fits according to the
168 Bayesian Information Criterion (BIC), given by $\text{BIC} = -2l(\hat{\mu}, \hat{\sigma}, \hat{\xi}) - k \log n$, where $l(\hat{\mu}, \hat{\sigma}, \hat{\xi})$ is
169 the maximized likelihood at empirical estimates $\hat{\mu}$, $\hat{\sigma}$, and $\hat{\xi}$. k is the number of parameters in the
170 GEVD (three for stationary GEVDs, four for nonstationary GEVDs), and n is the number of
171 yearly maxima used in the fit. A GEVD was fit to chlorophyll time series in each $\frac{1}{4}^\circ$ pixel. The
172 parameters were mapped spatially and correlated with basin bathymetry. Parameter uncertainty
173 was derived by taking the square-root of the inverse Hessian matrix evaluated at the maximum
174 likelihood estimates.

175

176 **Results**

177

178 The distributions of annual chlorophyll showed excellent agreement with those predicted from
179 the GEVD (**Figure 2**). Across the Atlantic basin, observed distribution quantiles correlated with
180 those from the fitted GEVDs at $r = 0.97$ on the arithmetic scale (**Figure 2a**) and $r = 0.98$ on the
181 log scale (**Figure 2b**). The spatial distribution of the quantile-quantile correlation was also
182 consistent across the basin, with no apparent relationship with latitude or distance from the coast
183 (**Figure 2c**). The region of largest disagreement occurred off the southwest coast of Europe, yet
184 correlations were still above $r = 0.7$ and remained so across the basin.

185

186 When fitted GEVD parameters were mapped spatially we found that parameter magnitude
187 closely followed basin bathymetry (**Figures 3-4**). Location and shape parameters were
188 consistently elevated on the shelf (<700m depth; **Figure 3c,d; Figure 4a,c**) with magnitudes 2-3
189 fold higher than in waters deeper than 700m. This result demonstrated that interannual bloom
190 magnitude and the ‘heaviness’ of the underlying distribution tail were both elevated on the shelf.
191 This occurred on the eastern and western sides of the basin. Deeper slope waters off the coast of
192 Greenland also showed elevated location and shape parameters. The largest parameter
193 magnitudes were found in the shallow Baltic Sea (**Figure 3c,d; Figure 4a,c**). The scale
194 parameter showed a different pattern with bathymetry where parameter magnitude was only
195 slightly elevated in shelf waters but showed a step-change decrease in the deepest waters
196 (>4000m; **Figure 3c; Figure 4b**); however, we note the aerial distribution of shelf vs. deep water
197 is markedly different, with deep waters limited to the southern half of the basin around the Mid-
198 Atlantic ridge while shelf seas are widely distributed (**Figure 3a**). Parameter uncertainty weakly
199 correlated with parameter magnitude for location and shape parameters with no clear spatial
200 pattern (**Supplementary Figure 1**). Scale parameter uncertainty was elevated in deeper water
201 (**Supplementary Figure 1**). The area-weighted average of the parameter coefficients of variation
202 (the standard deviation of the parameter uncertainty divided by the fitted mean parameter) was
203 0.11, 0.38, 0.24 for location, scale, and shape parameters, meaning the 1σ uncertainty was 11%,
204 38%, and 24% of the mean, respectively (**Supplementary Figure 1**).

205

206 We quantified the correlation between fitted GEVDs parameters using linear relationships
207 (**Figure 4d-f**). Bivariate relationships between parameters were well described by a linear
208 intercept and positive slope. The strongest relationship was found between fitted location and
209 shape parameters, again reflecting that GEVD distributions increase in magnitude and heavy
210 tailed-ness with decreasing bathymetric depth. We visually characterized how the GEVD
211 changes from deep to shelf waters using the fitted linear relationships (**Figure g-h**), noting that
212 this characterization represents the basin-averaged relationships so may not necessarily be
213 representative of individual regions.

214

215 Using a nonstationary GEVD analysis, we found weak evidence for temporal trends in GEVD
216 parameters, despite significant trends in chlorophyll levels and sea surface temperature across the
217 North Atlantic (**Figure 5**). Nonstationary parameters were favored in 34.3%, 29.9%, and 36.3%
218 of basin area for location, scale, and shape parameters, respectively (**Figures 5a-c**). Importantly,
219 the strength of evidence for nonstationary parameters did not clearly correlate with trends in
220 chlorophyll (**Figure 5d**; except possibly on the east Greenland slope), nor sea surface
221 temperature (**Figure 5e**). The weakest evidence for nonstationary parameters was found in the
222 area with the strongest sea surface temperature trends, specifically the warming-cooling dipole

223 pattern on the western side of the basin caused in-part by a slowdown of the Atlantic overturning
224 circulation and associated northward heat transport (i.e. the North Atlantic ‘warming hole’; Keil
225 et al. 2020).

226
227 We repeated the GEVD parameter estimation using the OC-CCI chlorophyll product and found
228 consistent results to those based on MODIS observations presented above. Quantile-quantile
229 correlations showed a similarly good fit between observed OC-CCI quantiles and those predicted
230 from the fitted GEVDs with a correlation of $r = 0.98$ in arithmetic space and $r = 0.98$ in log
231 space (**Supplementary Figure 2**). Spatial patterns in fitted GEVD parameters were consistent
232 across parameter sets estimated using the two datasets, despite some evidence for slightly
233 reduced scale parameter magnitudes using OC-CCI chlorophyll (**Supplementary Figure 3**),
234 which may be due to reduced variance in the synthetic multi-sensor OC-CCI dataset.

235

236 **Discussion**

237

238 Our analysis demonstrates that annual chlorophyll maxima are well-described by the GEVD
239 based on the statistical theory of extreme values. We achieved a high goodness-of-fit and
240 interpretable spatial patterns across the North Atlantic basin. A clear pattern emerged in the
241 correlation between GEVD parameters and bathymetric depth, with the magnitude and tailed-
242 ness of chlorophyll extremes increasing in shelf and slope environments. While the mechanism
243 for this pattern will require further research, we hypothesize that nutrient and stratification
244 dynamics may play a role. For example, elevated blooms are seen in other shelf and slope
245 ecosystems related to nutrient delivery and water column stability controlled by the shelf-break
246 circulation (Garcia et al. 2008). The heavy distribution tail on the shelf may also be related to
247 variability in bloom timing which has been shown to impact interannual variability in shelf
248 bloom magnitude (Friedland et al. 2015). We suggest further work examining the potential
249 mechanisms that could explain different extreme value distributions under contrasting
250 oceanographic environments. Classical models of phytoplankton blooms (e.g. Behrenfeld and
251 Boss 2014) may be extended to include stochastic forcing and generate statistical distributions of
252 bloom interannual magnitude. Targeted sensitivity of annual maxima distributions to different
253 underlying forcing may uncover mechanisms for changes in distribution parameters across the
254 shelf and open ocean.

255

256 Results of the nonstationary analysis suggested surprisingly weak evidence for changes in GEVD
257 parameters over time. The lack of correlation with observed trends in background chlorophyll
258 and sea surface temperature suggests that climate-related drivers are playing a limited role in
259 modulating interannual bloom magnitude, despite the North Atlantic showing significant climate
260 change (Keil et al. 2020). We note, however, that the current nonstationary analysis is limited by
261 time series length. Continued studies will be required to monitor changes in bloom magnitude
262 over time. An extended satellite record will provide greater statistical power to detect climate-
263 driven trends. An extended record will also constrain more complicated functions describing the
264 variation of parameters with time and their statistical association with environmental factors.

265

266 Ecologically, interesting questions arise about how extreme blooms propagate through food
267 webs, contribute to carbon export, and impact ecological processes more broadly. For example,
268 annual fisheries recruitment is often characterized by heavy-tailed distributions where individual

269 years exhibit extremely large cohorts, often fueling the fishery for years (Saetre et al. 2002).
270 Extreme bloom years may increase the probability of strong cohorts via the match-mismatch
271 mechanism (Platt et al. 2003), perhaps with temporal lags between blooms and recruitment
272 modulated by trophic transfer. With respect to carbon export, we expect extreme blooms to
273 contribute disproportionately to interannual carbon fluxes due to the commonly-observed
274 positive effect of phytoplankton productivity on carbon export efficiency (Britten and Primeau
275 2016) and observed carbon fluxes associated with blooms in the North Atlantic (Briggs et al.
276 2011). Databases of carbon flux observations may be used to test this hypothesis at the basin
277 scale (Mouw et al. 2016). Beyond fisheries and carbon export, we envision extreme event
278 distributions to be broadly useful in characterizing the response of ecological processes to
279 environmental extremes. The increase in applications of extreme value theory to environmental
280 processes (Aghakouchak et al. 2020) naturally leads to questions of how environmental extremes
281 impact ecology. The GEVD is one extreme value analysis tool that utilizes the distribution of
282 block maxima (which was particularly appropriate here to describe the maxima of a repeating
283 annual cycle) however other statistical descriptions of ecological heavy tailed-ness can also be
284 useful in this context (e.g. Anderson et al. 2017).

285
286 In summary, the GEVD provided a useful statistical description of bloom variability and a
287 general framework to quantify the spatiotemporal statistics of interannual bloom maxima. We
288 hope this study spurs further analysis of marine ecosystem variability using extreme value
289 theory to better understand how environmental conditions give rise to ecological extreme events,
290 how extreme blooms contribute to fisheries productivity and carbon export, and how these
291 processes may change with climate.

292 **Acknowledgements**

293
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295 Computational Biogeochemical Modeling of Marine Ecosystems and the Simons Foundation
296 Postdoctoral Fellowship in Marine Microbial Ecology.

297
298 **Data and Code Availability:** All data used in this study are publicly available via the links
299 given in the text. Code used to perform the GEVD analysis and visualize the results is publicly
300 available at: https://github.com/gregbritten/chl_extremes_public.

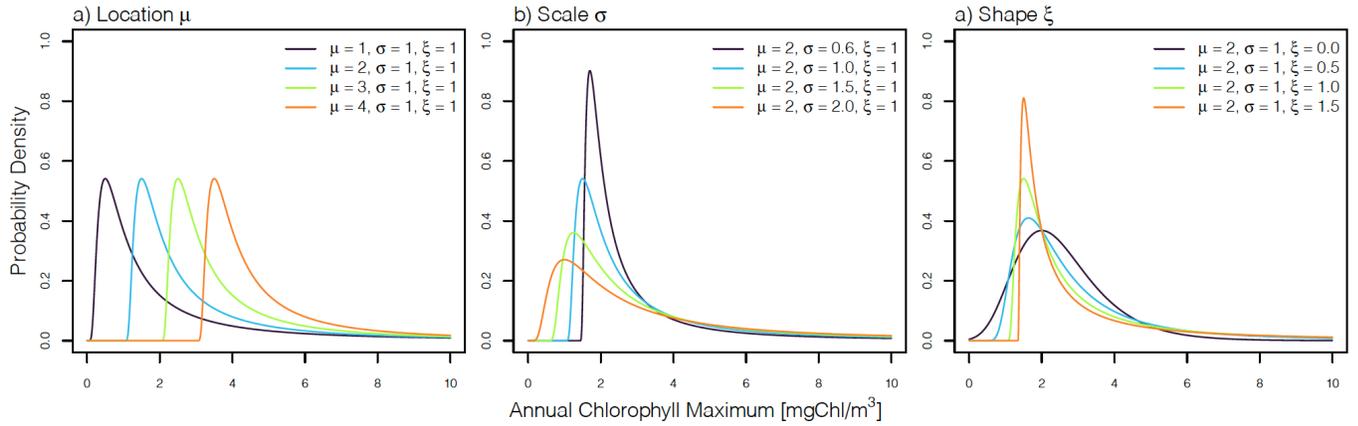
301 **References**

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304
305 Aghakouchak, A., F. Chiang, L. S. Huning, and others. 2020. Climate extremes and compound
306 hazards in a warming world. *Annu. Rev. Earth Planet. Sci.* **48**: 519–548.
307 Anderson, S. C., T. A. Branch, A. B. Cooper, and N. K. Dulvy. 2017. Black-swan events in
308 animal populations. *Proc. Natl. Acad. Sci.* **114**: 3252–3257.
309 Batt, R. D., S. R. Carpenter, and A. R. Ives. 2017. Extreme events in lake ecosystem time series.
310 *Limnol. Oceanogr. Lett.* **2**: 63–69.
311 Behrenfeld, M. J. 2010. Abandoning sverdrup’s critical depth hypothesis on phytoplankton
312 blooms. *Ecology* **91**: 977–989.
313 Behrenfeld, M. J., and E. S. Boss. 2014. Resurrecting the ecological underpinnings of ocean
314 plankton blooms. *Ann. Rev. Mar. Sci.* **6**: 167–194.

- 315 Behrenfeld, M. J., and E. S. Boss. 2018. Student's tutorial on bloom hypotheses in the context of
316 phytoplankton annual cycles. *Glob. Chang. Biol.* **24**: 55–77.
- 317 Briggs, N., M. J. Perry, I. Cetinić, C. Lee, E. D'Asaro, A. M. Gray, and E. Rehm. 2011. High-
318 resolution observations of aggregate flux during a sub-polar North Atlantic spring bloom.
319 *Deep. Res. Part I Oceanogr. Res. Pap.* **58**: 1031–1039.
- 320 Britten, G., and F. Primeau. 2016. Biome-specific scaling of ocean productivity, temperature,
321 and carbon export efficiency. *Geophys. Res. Lett.* **43**: 5210–5216.
- 322 Coles, S. 2001. An introduction to statistical modeling of extreme values, Springer-Verlag.
- 323 Dutkiewicz, S., M. Follows, J. Marshall, and W. W. Gregg. 2001. Interannual variability of
324 phytoplankton abundances in the North Atlantic. *Deep. Res. Part II Top. Stud. Oceanogr.*
325 **48**: 2323–2344.
- 326 Easterling, D. R., G. A. Meehl, C. Parmesan, S. A. Changnon, T. R. Karl, and L. O. Mearns.
327 2000. Climate extremes: observations, modeling, and impacts. *Science* **289**: 2068–2075.
- 328 Evans, G. T., J. S. Parslow, G. T. Evans, and J. S. Parslow. 1985. A model of annual plankton
329 cycles. *Deep Sea Res. Part B. Oceanogr. Lit. Rev.* **32**: 759.
- 330 Falkowski, P., R. Barber, and V. Smetacek. 1998. Biogeochemical controls and feedbacks on
331 ocean primary production. *Science* **281**: 200–206.
- 332 Falkowski, P., E. Laws, R. Barber, and J. Murray. 2003. Phytoplankton and their role in primary,
333 new and export production, p. 99–121. *In* M.J.R. Fasham [ed.], *Ocean Biogeochemistry*.
334 Springer.
- 335 Ferrari, R., S. T. Merrifield, and J. R. Taylor. 2015. Shutdown of convection triggers increase of
336 surface chlorophyll. *J. Mar. Syst.* **147**: 116–122.
- 337 Follows, M., and S. Dutkiewicz. 2002. Meteorological modulation of the North Atlantic spring
338 bloom. *Deep Sea Res. Part II Top. Stud. Oceanogr.* **49**: 321–344.
- 339 Friedland, K. D., R. T. Leaf, J. Kane, and others. 2015. Spring bloom dynamics and zooplankton
340 biomass response on the US Northeast Continental Shelf. *Cont. Shelf Res.* **102**: 47–61.
- 341 Garcia, V. M. T., C. A. E. Garcia, M. M. Mata, R. C. Pollery, A. R. Piola, S. R. Signorini, C. R.
342 McClain, and M. D. Iglesias-Rodriguez. 2008. Environmental factors controlling the
343 phytoplankton blooms at the Patagonia shelf-break in spring. *Deep. Res. Part I Oceanogr.*
344 *Res. Pap.* **55**: 1150–1166.
- 345 Gilleland, E., and R. W. Katz. 2016. extRemes 2.0: An extreme value analysis package in R. *J.*
346 *Stat. Softw.* **72**: 1–39.
- 347 Ito, T., and M. J. Follows. 2005. Preformed phosphate, soft tissue pump and atmospheric CO₂. *J.*
348 *Mar. Res.* **63**: 813–839.
- 349 Katz, R. W. 2010. Statistics of extremes in climate change. *Clim. Change* **100**: 71–76.
- 350 Keil, P., T. Mauritsen, J. Jungclaus, C. Hedemann, D. Olonscheck, and R. Ghosh. 2020. Multiple
351 drivers of the North Atlantic warming hole. *Nat. Clim. Chang.* **10**: 667–671.
- 352 Maritorena, S., and D. a. Siegel. 2005. Consistent merging of satellite ocean color data sets using
353 a bio-optical model. *Remote Sens. Environ.* **94**: 429–440.
- 354 Mouw, C. B., A. Barnett, G. A. Mckinley, L. Gloege, and D. Pilcher. 2016. Global ocean
355 particulate organic carbon flux merged with satellite parameters. *Earth Syst. Sci. Data* **8**:
356 531–541.
- 357 Platt, T., C. Fuentes-Yaco, and K. T. Frank. 2003. Spring algal bloom and larval fish survival.
358 *Nature* **423**: 398–399.
- 359 Saetre, R., R. Toresen, and T. Anker-Nilssen. 2002. Factors affecting the recruitment variability
360 of the Norwegian spring-spawning herring (*Clupea harengus* L.). *ICES J. Mar. Sci.* **59**:

- 361 725–736.
- 362 Sathyendranath, S., R. J. W. Brewin, C. Brockmann, and others. 2019. An ocean-colour time
363 series for use in climate studies: The experience of the ocean-colour climate change
364 initiative (OC-CCI). *Sensors* **19**: 1–31.
- 365 Siegel, D. A., S. C. Doney, and J. A. Yoder. 2002. The North Atlantic spring phytoplankton
366 bloom and Sverdrup’s critical depth hypothesis. *Science* **296**: 730–733.
- 367 Sommer, U., and K. Lengfellner. 2008. Climate change and the timing, magnitude, and
368 composition of the phytoplankton spring bloom. *Glob. Chang. Biol.* **14**: 1199–1208.
- 369 Taylor, J. R., and R. Ferrari. 2011. Ocean fronts trigger high latitude phytoplankton blooms.
370 *Geophys. Res. Lett.* **38**: 1–5.
- 371 Visser, F., K. L. Hartman, G. J. Pierce, V. D. Valavanis, and J. Huisman. 2011. Timing of
372 migratory baleen whales at the azores in relation to the north atlantic spring bloom. *Mar.*
373 *Ecol. Prog. Ser.* **440**: 267–279.
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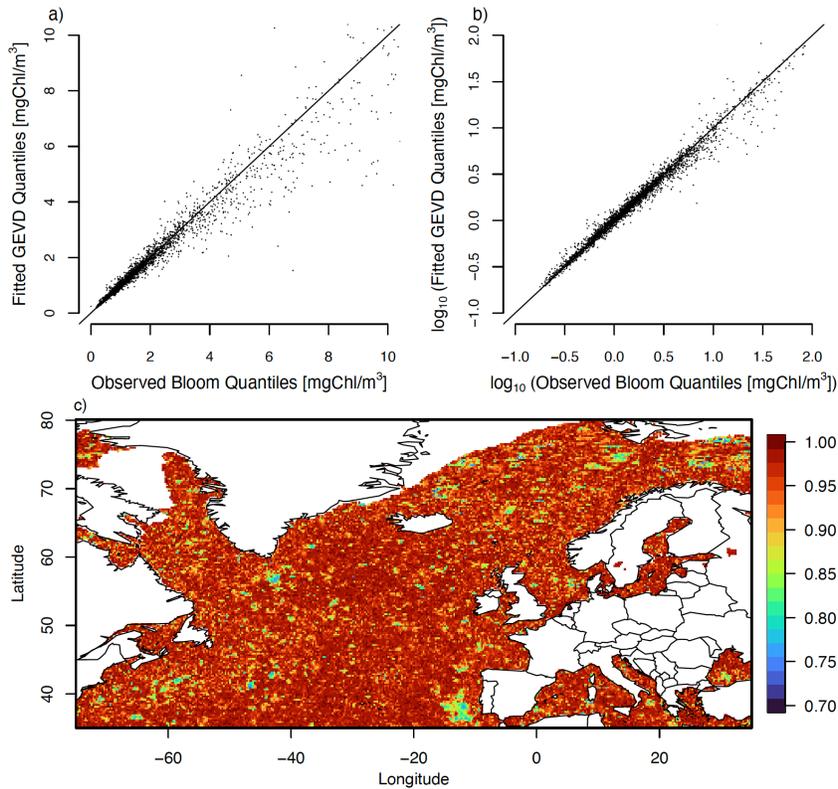
375 **Figures**
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Figure 1. Examples of the GEVD distribution for different values of the location **(a)**, scale **(b)**, and shape **(c)** parameters.

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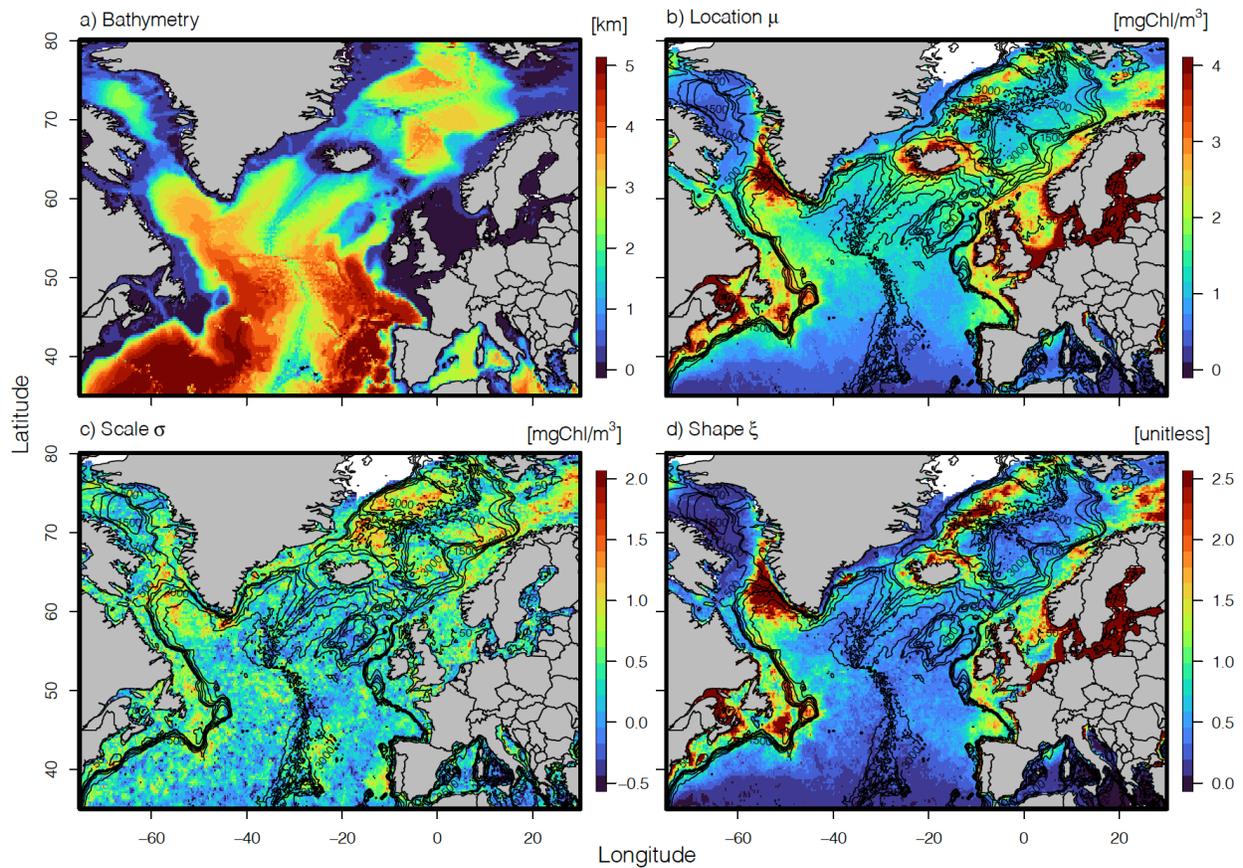


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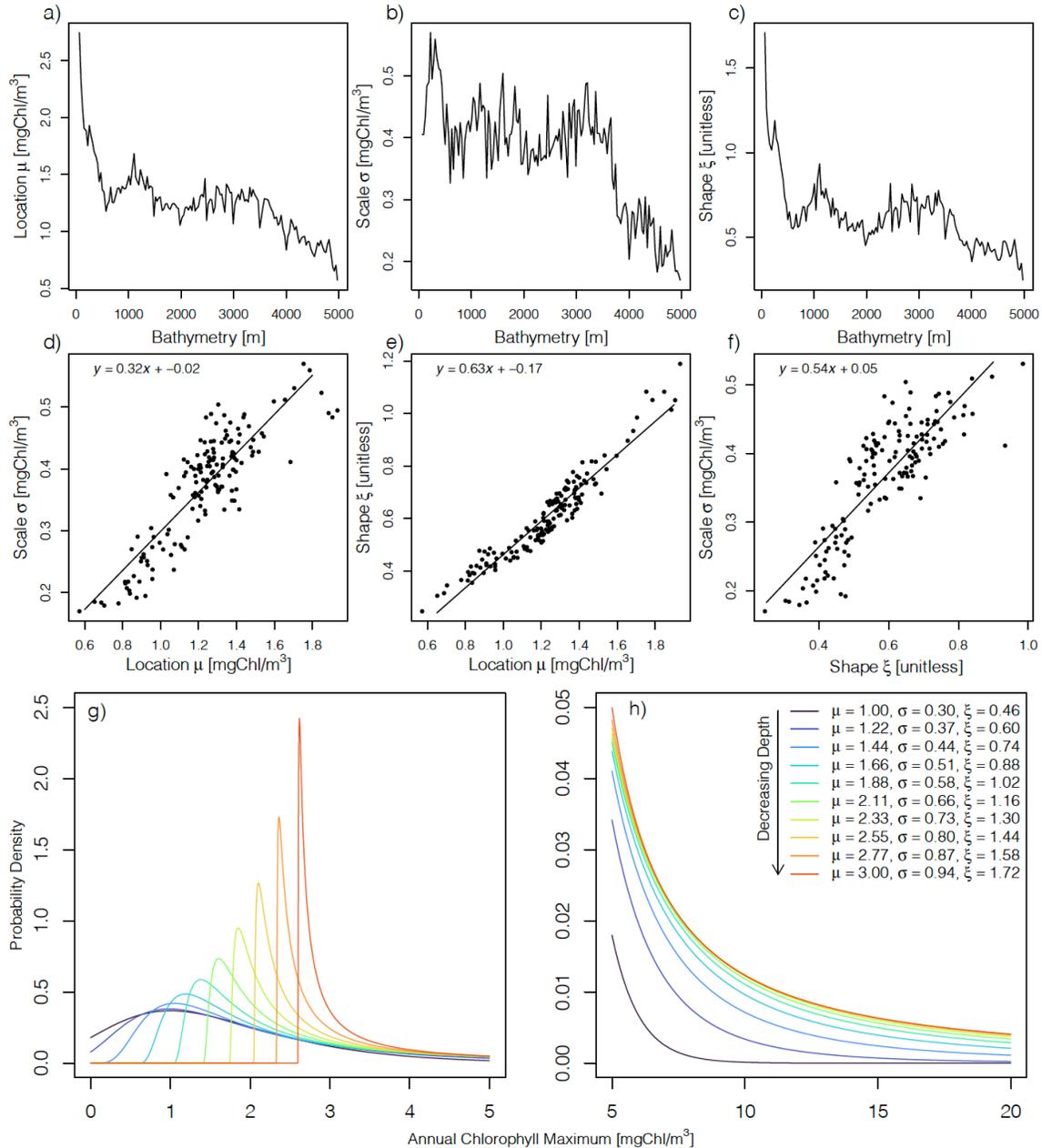
384 Figure 2. Observed quantiles of North Atlantic annual bloom extremes vs. those predicted from
 385 the fitted GEVDs. Panel **a** gives the relationship in arithmetic space; panel **b** gives the
 386 relationship in log (base ten) space. A random sample of 5000 quantiles were drawn in order to
 387 visualize the relationship in **a** and **b**. The correlations in **a** and **b** are 0.97 and 0.98, respectively.
 388 Panel **c** gives the spatial distribution of quantile-quantile correlations. Note the correlation color
 389 bar extends from 0.7 to 1.0.

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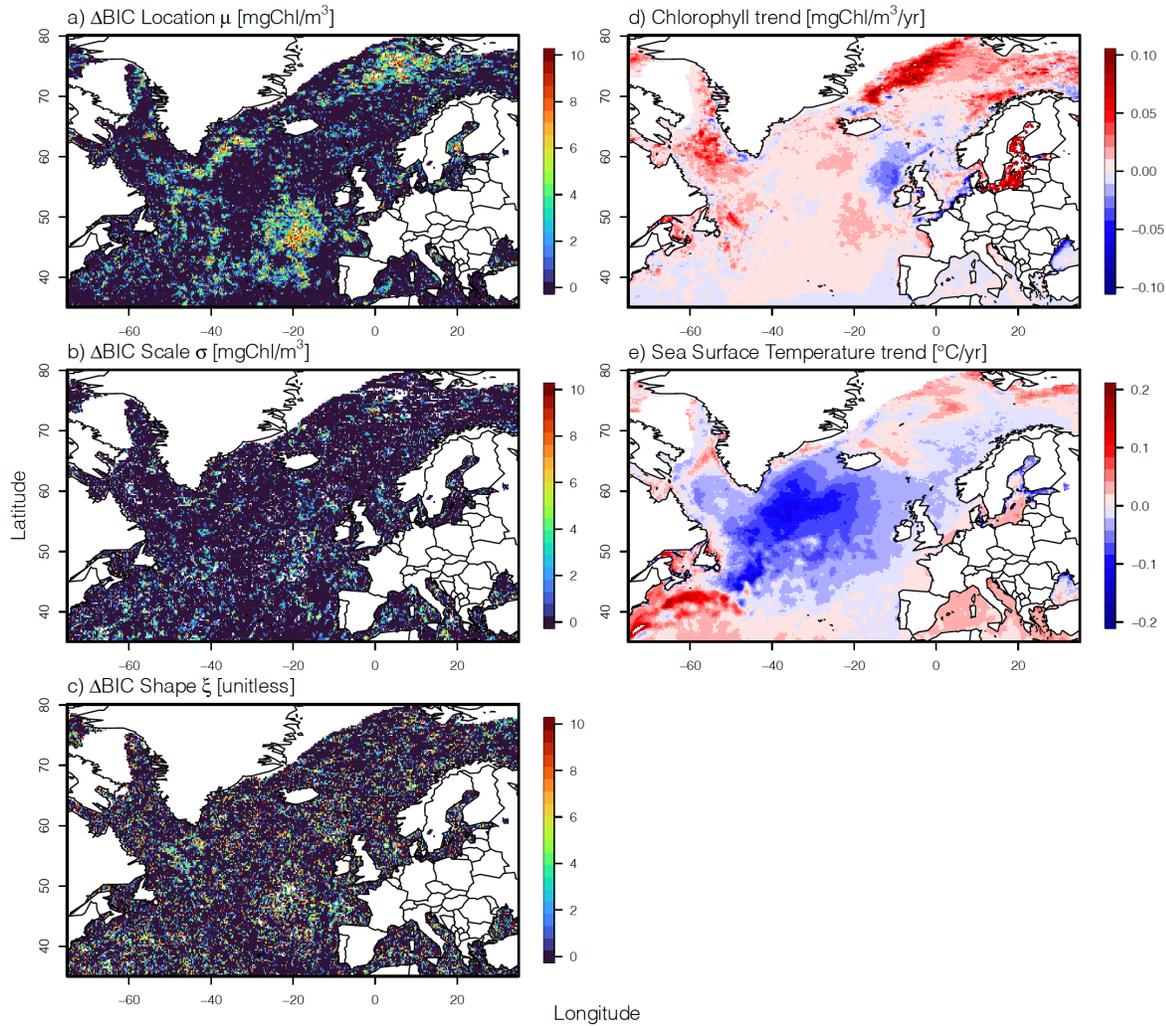
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Figure 3. Spatial distribution of fitted GEVD parameters and basin bathymetry. Panel **a** gives the bathymetric depth in the North Atlantic basin. Panels **b**, **c**, and **d** give the estimated location, scale, and shape parameters, respectively. Bathymetry contours are overlaid in **b-d** with a contour interval of 500 meters.



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399 Figure 4. Relationships between fitted GEVD parameter and bathymetric depth. Panels a-c give
400 the basin-averaged relationship between the fitted parameters and bathymetric depth. Panels d-f
401 give the inter-relationships among fitted parameters with the least squares regression line and
402 equation. Panels g and h give the basin-averaged change of the GEVD with bathymetric depth.
403 Panel h zooms in on the distribution tail in g (note horizontal axis limits).



404
 405 Figure 5. Evidence for nonstationary GEVD parameters in the context of basin-scale trends in
 406 chlorophyll and sea surface temperature. Model selection results comparing stationary and
 407 nonstationary GEVDs using the Bayesian Information (BIC) are given in panel **a** (nonstationary
 408 location parameter), **b** (nonstationary scale), and **c** (nonstationary shape). The difference in BIC
 409 (Δ BIC) is given relative to the stationary model. Non-zero values indicate the nonstationary
 410 model is favoured and the magnitude indicates the relative strength of evidence. Panels **d** and **e**
 411 give the linear trend in surface chlorophyll and sea surface temperature, respectively, estimated
 412 from MODIS observations since 2002.

413 **Appendix A**

414

415 The expected value of the GEVD is given by

416

417
$$\mathbf{E}[x] = \begin{cases} \mu + (g(1) - 1) \frac{\sigma}{\xi} & \text{if } \xi \neq 0, \xi < 1 \\ \mu + \sigma\gamma & \text{if } \xi = 0 \\ \infty & \text{if } \xi > 1 \end{cases}$$

418

419 where $g(k) = \Gamma(1 - k\xi)$, $\Gamma(r)$ is the gamma function, and γ is Euler's constant ($\gamma \cong 0.5772$).

420 The variance of the GEVD is

421

422
$$\mathbf{Var}[x] = (g(2) - g(1)^2) \frac{\sigma^2}{\xi^2}.$$

423

424 The mode of the GEVD is

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426
$$\mathbf{Mode}[x] = \mu + \frac{\sigma}{\xi} ((1 + \xi)^{-\xi} - 1).$$

427

Supplemental Information:

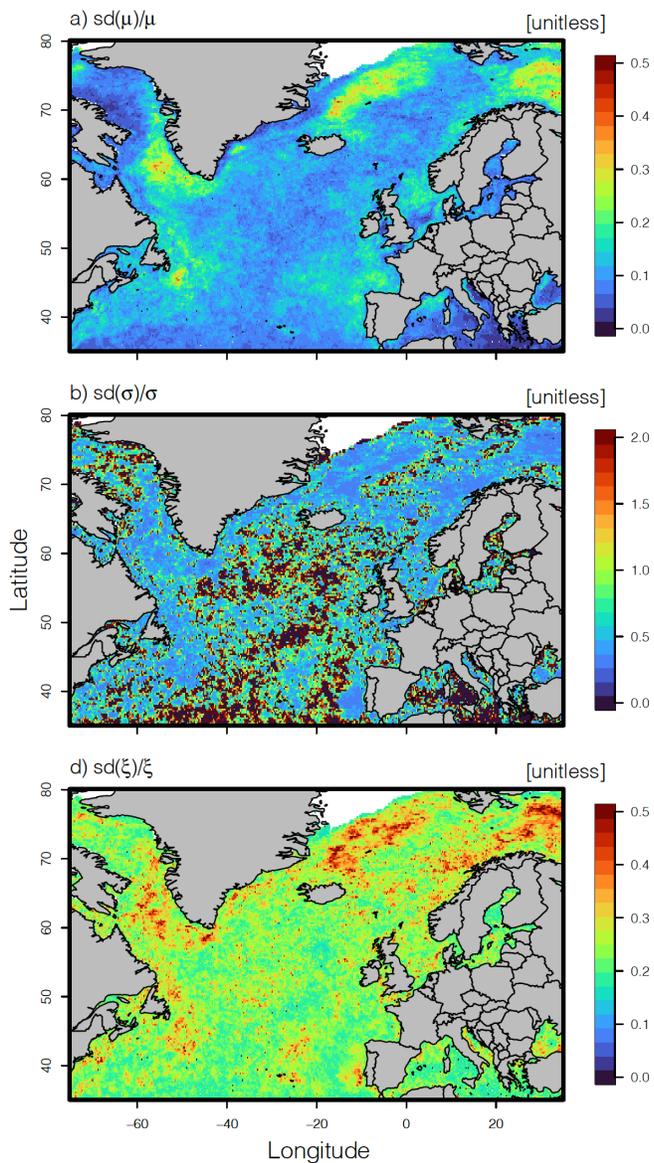
Extreme value distributions describe interannual variability in the North Atlantic spring bloom

Gregory L. Britten

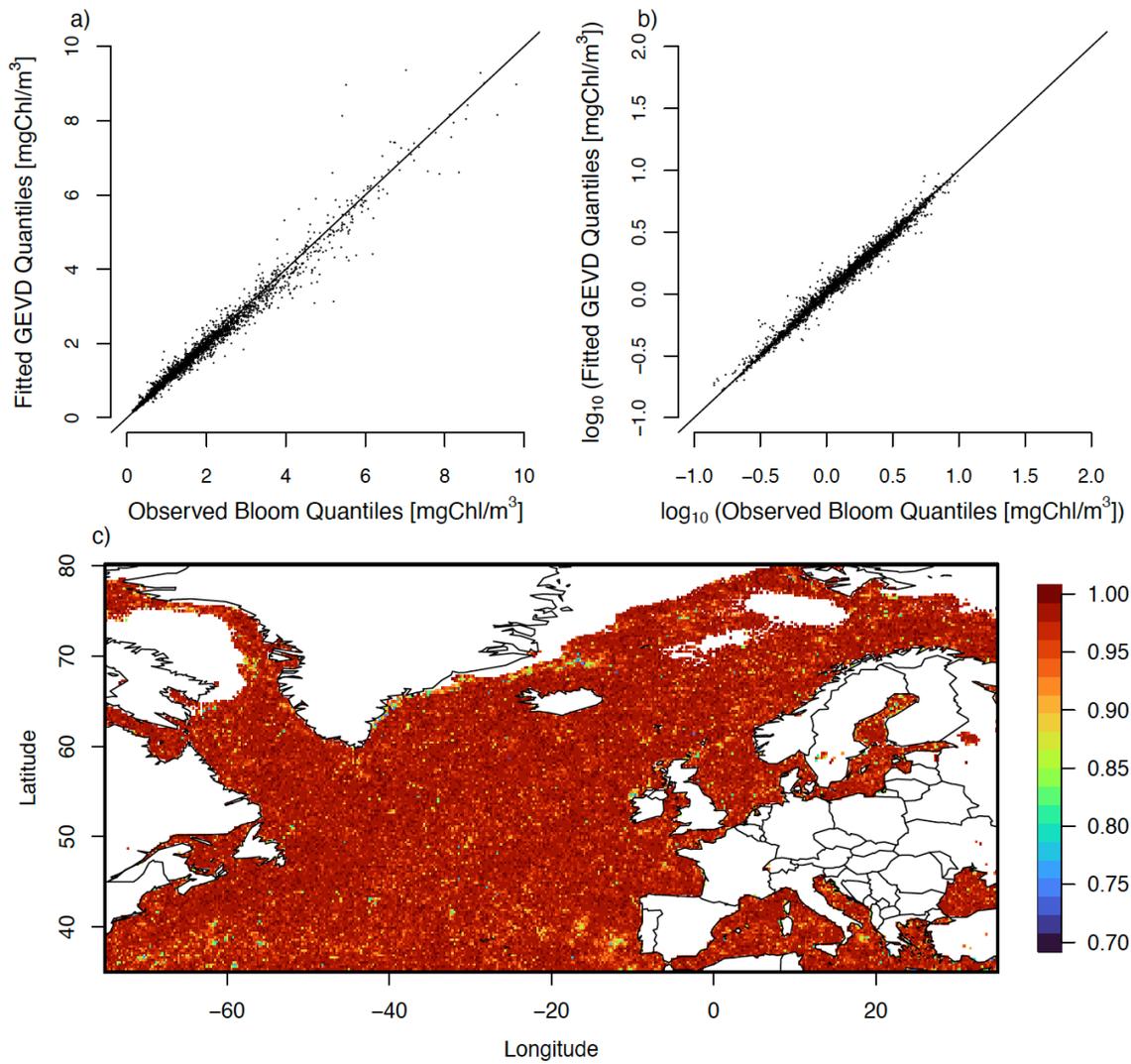
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Massachusetts Institute of Technology, Cambridge, MA, USA

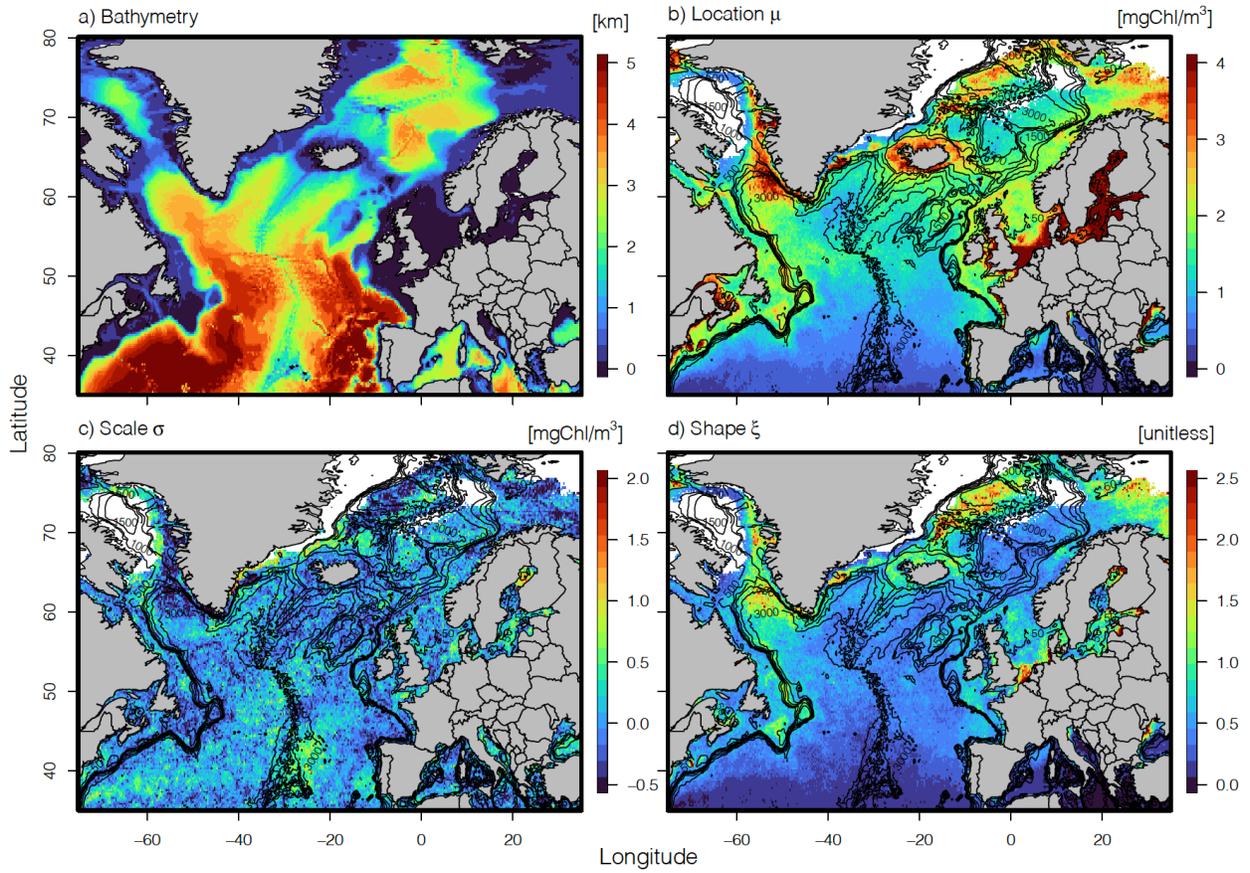
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Supplementary Figure 1. Uncertainty in fitted GEVD parameters expressed as the coefficient of variation (CV). The CV is calculated as the ratio of the uncertainty standard deviation divided by the mean of the fitted parameter.



Supplementary Figure 2. As in Figure 2 of the main text but using OC-CCI chlorophyll estimates



Supplementary Figure 3. As in Figure 3 of the main text but using OC-CCI chlorophyll estimates.