

Subcutaneous Adipose Tissue: Implications in Dermatological Diseases and Beyond

Reihane Ziadlou¹, Ganesh N. Pandian², Jürg Hafner³, Cezmi Akdis¹, Georg Stingl⁴, Emanuel Maverakis⁵, and Marie-Charlotte Brüggen¹

¹Universität Zurich Medizinische Fakultät

²Kyoto University

³UniversitätsSpital Zurich Dermatologische Klinik

⁴Medical University of Vienna

⁵University of California Davis Department of Dermatology

March 25, 2024

Abstract

Subcutaneous adipose tissue (SAT) is the deepest component of the three-layered cutaneous integument. While mesenteric adipose tissue-based immune processes have gained recognition in the context of the metabolic syndrome, SAT has been traditionally considered primarily for energy storage, with less attention to its immune functions. SAT harbors a reservoir of immune and stromal cells that significantly impact metabolic and immunologic processes not only in the skin, but even on a systemic level. These processes include wound healing, cutaneous and systemic infections, immunometabolic and autoimmune diseases, inflammatory skin diseases, as well as neoplastic conditions. A better understanding of SAT immune functions in different processes, could open avenues for novel therapeutic interventions. Targeting SAT may not only address SAT-specific diseases but also offer potential treatments for cutaneous or even systemic conditions. This review aims to provide a comprehensive overview on SAT's structure and functions, highlight recent advancements in understanding its role in both homeostatic and pathological conditions within and beyond the skin, and discuss the main questions for future research in the field.

Introduction

Adipose tissue (AT) has traditionally been viewed as an inert "cushioning" layer providing mechanical protection and serving as an energy storage site (1). However, research over the past decades has uncovered its dynamic nature, revealing AT as a highly active organ with metabolic, endocrine, immune, and biomechanical functions (2). AT plays a central role in the pathogenesis of various diseases, including diabetes, cardiovascular disease, osteoarthritis, and cancer (3-5). Situated throughout the body, AT encompasses the deepest layer of the cutaneous integument, known as subcutaneous adipose tissue (SAT), along with the epidermis and dermis (6). SAT's involvement in both immune and metabolic processes has been insufficiently explored.

Given that obesity has become a worldwide pandemic (7), additional attention to SAT physiology is necessitated, especially its contributions to conditions like diabetes and immune-mediated skin diseases such as psoriasis and hidradenitis suppurativa (6, 8-11).

This review aims to provide an overview of the current understanding of SAT structure and functions, emphasizing its association with various diseases. Additionally, we will explore the immunological functions of SAT in the context of both cutaneous and systemic diseases, examining its potential role in immune-mediated skin infections.

AT types: structures and cellular composition

AT subtypes can be organized by their anatomical localization in mammals. SAT, found beneath the skin, contrasts with visceral adipose tissue (VAT), which lines internal organs (12). In addition, structural and functional features can be used to divide AT into three subtypes: white adipose tissue (WAT), brown adipose tissue (BAT), and beige adipose tissue (Fig 1a).

WAT is mainly known for its role in energy storage and immune regulation (13). It is the main constituent of visceral, and ectopic subcutaneous AT (SAT). In rodents, a striated muscle layer, the panniculus carnosus, subdivides SAT into two functionally distinct compartments, namely subcutaneous and dermal SAT (14). Such a barrier is missing in human SAT (14, 15).

In contrast to WAT, BAT is highly specialized for thermogenesis, capable of dissipating stored energy as heat to maintain optimal body temperature (16). BAT is abundant and broadly distributed in newborns. In adults, BAT is limited to cervical, supraclavicular, paravertebral, mesenteric, and pericardial areas. Beige AT emerges through “browning” of WAT, induced by external stimuli, such as low temperature or exercise (17, 18).

In all AT types, approximately one-third of the cellular content consists of adipocytes. The remaining two-thirds constitute the stromal vascular fraction (SVF) (Figure 1b). The stromal component of AT contains adipose stem cells (ASCs), preadipocytes, fibroblasts, endothelial cells, and immune cells. ASCs serve as precursor cells for preadipocytes (19, 20), specialized progenitors committed to becoming adipocytes and residing in a unique perivascular tissue niche (21-23). Fibroblasts in the SVF provide support to preadipocytes and help to maintain the adipose tissue homeostasis (24). In SAT, immune cells include macrophages (Mac), helper, cytotoxic, and regulatory T cells, natural killer cells, and B-lymphocytes (25). In healthy individuals, T cells and macrophages in SAT tend to favor a type 2 and regulatory phenotype (26-28). While T cells in epidermis and dermis generally adopt a T-helper 1 (Th1) phenotype, acting as the primary defense line in homeostatic conditions (26), T cells in SAT may function as counter-regulators (Fig 1c).

Immune processes in the context of cutaneous and systemic diseases in the SAT

Moving into the discussion of immune processes, the skin acts as a physical barrier, orchestrating a complex interplay of structural and cellular elements. Resident and migrating immune cells protect against pathogens. The cutaneous immune system can also trigger pathologic responses, leading to allergies, autoimmunity, and autoinflammatory conditions (29-31). However, the role of SAT in the cutaneous immune system and its impact under homeostatic and pathogenic conditions has been poorly characterized.

Evidence suggests that SAT’s reservoir of immune and stromal cells may direct metabolic and immunologic processes (29-31). SAT-mediated pathologic responses can manifest within SAT, the overlying dermis or epidermis, or extracutaneous sites throughout the body (Fig 2). Examples include SAT’s involvement in (i) cutaneous wound healing (27, 32, 33), (ii) induction of a protective immune responses (34), (iii) modulation of immunologic and metabolic processes, (iv) regulation of cutaneous inflammatory diseases, (v) promotion of neoplastic processes, and (vi) influence on the phenotype of various genodermatoses (35, 36) (Fig 2). However, much of this evidence is derived from animal studies, necessitating further investigations to understand SAT-mediated pathologies in humans and its communication with superficial skin layers.

1. SAT in wound healing

Wound healing consists of several regenerative phases (Fig 3), in which keratinocytes act as the main effectors by supporting fibroblasts, leukocytes, and mesenchymal cells (37). SAT-based processes play an essential role in all phases of wound healing via the secretion of glucocorticoids, adipokines (e.g., interleukins (IL)-1 β , -6, -8, -10; leptin, adiponectin, MCP-1, TNF), and other bioactive molecules (e.g., vascular endothelial growth factor (VEGF), basic fibroblast growth factor (bFGF), transforming growth factor beta (TGF β) (38-41). In a mouse model, dermal adipocytes played a crucial role in initiating inflammation post-injury contributed to wound repair by dedifferentiating into myofibroblasts, for extracellular matrix (ECM) production (32,

42). A particularly important role in wound healing has been attributed to ASCs. ASCs promote cutaneous neovascularization and re-epithelialization through secretion of growth factors and cytokines (43-45). Several pre-clinical studies have shown the potential therapeutic effect of ASCs in wound repair (46, 47). Despite ASCs being considered a relatively safe source of stem cells, their widespread therapeutic application is currently hindered by barriers such as cost and the absence of highly standardized cell preparation methodologies (46). As an alternative to ASC-based cell therapy, the administration of ASC-derived exosomes (48-50) has been explored, demonstrating immunomodulatory effects and the ability to promote angiogenesis and re-epithelization (51, 52) (Fig 3a).

2. The role of SAT in induction of a protective immune response against pathogens

The skin serves as the primary defense against pathogen invasion. It provides both a physical barrier and an intrinsic warning system to trigger innate and adaptive immune responses when the physical barrier is breached. The role of epidermal/dermal leukocytes, keratinocytes, and other non-leukocyte populations in antimicrobial defense has been well investigated. In contrast, the contribution of underlying SAT to this process remains largely unexplored (53).

One avenue through which adipocytes can participate in antimicrobial defense is through the release of soluble mediators. Adipokines released by adipocytes, as shown in a series of mouse studies, have the ability to recruit immune cells to infection sites and modulate their effector functions (54, 55). Leptin, a well-characterized adipokine known for its role in hunger regulation, also exhibits immunomodulatory properties, contributing to antimicrobial immune responses (56-58). Studies on leptin/leptin receptor-deficient mice have revealed increased susceptibility to viral or bacterial infections (59-61). In obese individuals, elevated blood levels of leptin lead to leptin resistance, which in turn induces a reduced type I interferon (IFN) response and increased susceptibility to viral infection (62, 63).

Adipocytes are also a major secretor of cathelicidins, short cationic antimicrobial peptides (30, 64) (Fig 4). Obese animals produce fewer cathelicidins, thereby contributing to compromised infection control (65) (Table 1). Beyond adipocytes, one finding that links AT to the immune system is that WAT harbors a significant number of resident memory T-cells. This population can be rapidly reactivated to provide protection against pathogens (66). Studies in mice and humans indicate that obesity is associated with impaired memory T-cell responses and reduced natural killer cell cytotoxicity (67-76). Furthermore, systemic viral infections have been shown to alter SAT immune-metabolic functions in mice, notably by inducing AT expansion (77-79). Unraveling the specific mechanisms through which SAT contributes to immune defense may open avenues for therapeutic interventions targeting both metabolic and immunologic aspects, with potential implications for preventing and managing infectious diseases.

3. SAT in immuno-metabolic diseases

Obesity is associated with a state of low-grade inflammation in SAT. This poses a heightened risk for the development of various health conditions, including type 2 diabetes (T2D), autoimmune and autoinflammatory diseases, cardiovascular disease, and cancer (5, 80-88). The systemic low-grade inflammation associated with obesity contributes to insulin resistance in skeletal muscle and liver (89, 90). Additionally, AT macrophages and innate lymphoid cells type 1 (ILCs1) promote AT fibrosis by inducing ECM deposition, which contributes to insulin resistance and T2D (91, 92). Inhibition of AT fibrosis may be a mechanism to improve glucose intolerance (93).

The inflammatory state linked to obesity stems from multiple mechanisms. In individuals with obesity, the expansion of adipocytes leads to increased release of adipokines like leptin and resistin, alongside decreased levels of the anti-inflammatory adiponectin (94, 95). This directly promotes a phenotypic shift of adipose tissue-resident immune cells toward a pro-inflammatory state (96-98). Investigations into lymphocyte responses in obesity highlight a skewed polarization of SAT-resident helper T cells in obese individuals towards a pro-inflammatory Th1 phenotype (99-101) (Fig 5). SAT adipocytes of obese patients also express all 10 Toll-like receptors (TLRs), with TLR-4 exhibiting the highest expression (102, 103). TLR4 activation triggers the NF- κ B signaling pathway in adipocytes and monocytes/macrophages, subsequently leading to

the release of monocyte chemoattractant protein-1 (MCP-1) and pro-inflammatory cytokines such as interleukin β (IL1 β), tumor necrosis factor- α (TNF- α), interleukin 6 (IL-6) (104, 105). Elevated MCP-1 levels further prompt the infiltration of monocytes into SAT, where they differentiate into pro-inflammatory (M1) macrophages (Fig 5) (106, 107). Increased levels of TNF- α have significant effects in induction of lipolysis, the breakdown of fat stored in adipose tissue. TNF-induced lipolysis is a complex process involving the activation of inflammatory pathways, lipolytic enzyme activity and release of free fatty acids (FFAs) (108, 109). Elevated levels of FFAs released during lipolysis can impair insulin signaling in peripheral tissues such as muscle and liver, contributing to insulin resistance and metabolic dysfunction (110). Understanding these mechanisms is important for elucidating the role of TNF in metabolic disorders and inflammatory conditions associated with dysregulated lipid metabolism.

Excessive caloric intake in obesity also leads to increased reactive oxygen species (ROS) production in adipocytes, causing mitochondrial dysfunction (111). Abnormal mitochondrial function in adipocytes leads to lipid accumulation, ultimately contributing to metabolic syndrome (112). Therefore, mitigating excessive ROS production and chronic inflammation in SAT of obese individuals present a novel approach to address obesity-related immunometabolic disorders.

4. SAT in autoimmune diseases

SAT is recognized as an active contributor to immune regulation and modulation. Dysregulation in resident macrophages, T-regs and other immune cells within SAT can lead to excessive cytokine production and autoimmune diseases (113-115). This disrupted balance can be originated within the adipocytes in SAT secreting various pro-inflammatory cytokines and chemokines, creating an environment conducive to immune dysregulation. Furthermore, imbalances in adipokine levels may contribute to the dysregulation of immune responses and exacerbate autoimmune conditions (116, 117). A focused investigation into the specific roles of resident T-regs and macrophages along with exploration of the involvement of cytokines and adipokines in this dysregulation is crucial for understanding the pathways leading to autoimmune diseases.

MHC-like cell surface CD1 family proteins have the capacity to present lipid antigens (118, 119). Several studies suggest a possible role of AT-derived CD1-presented lipid antigens in autoimmunity. For example, adipocytes from obese mice express CD1d, contributing to the induction of an autoreactive immune response (120). A better understanding of the interplay between adipocytes, lipid autoantigens, and CD1 presentation will elucidate a new, and potentially targetable, pathway in autoimmunity. In healthy human skin, there appears to be competition between permissive and blocking lipids for presentation by CD1a, the balance of which can modulate T cell responses (121). Specifically, presentation of very long chain FAs, such as 42:2 sphingomyelin lipids, by CD1a has been observed to impede the engagement of CD1a-directed autoreactive T-cells (122). A disruption of this balance may favor the development of autoimmune processes. Therefore, it is intriguing to explore the CD1a-related functions and pathways as potential targets in the prevention and treatment of autoimmune conditions.

5. SAT in inflammatory skin diseases

Inflammatory processes within the SAT of the skin differ from those in the epidermis and dermis. There is limited research on this subject and most evidence comes from studies on psoriasis (123). Psoriasis is associated with an increased risk of cardiovascular and immunometabolic disorders, notably obesity (124, 125). The increased production of pro-inflammatory adipokines and decreased production of anti-inflammatory adiponectin in obesity may predispose individuals to develop psoriasis (126, 127). Animal models also indicate that diets high in saturated fatty acids can promote IL-17-mediated immune responses, leading to increased susceptibility to psoriasis (128, 129).

Dermal sclerosis is another pathogenic process that might be aided by aberrant responses in AT. Recent studies suggest the involvement of ECM produced by WAT-derived myofibroblasts in scleroderma pathogenesis (130, 131). As of yet, other neutrophilic and fibrotic diseases such as hidradenitis suppurativa (HS) have not yet been linked to AT; clinical evidence, namely the high incidence of obesity in HS patients and the distribution of inflammatory infiltrates in the follicular epithelium, strongly suggest a role of SAT (132,

133).

Inflammatory conditions primarily originating and taking place in SAT are grouped under the term “panniculitis”. Panniculitides encompass a range of heterogeneous etiologies, including infection, trauma, connective tissue diseases, vasculitis, and certain types of cancer (Table 2). Their classification considers location, lesion etiology, and histopathology. The latter takes into account whether SAT infiltration is septal or lobular and whether it is accompanied by vasculitis (134-136). Despite diverse etiologies, the cellular and molecular pathomechanisms underlying panniculitis remain poorly characterized. Therapeutic approaches remain widely nonspecific, including non-steroidal anti-inflammatory drugs, oral potassium iodide, dapsone, and hydroxychloroquine (137-141).

Panniculitides can originate either as primary pathologies within AT or as secondary manifestations of systemic diseases. For instance, erythema nodosum (EN), the most common type of panniculitis, may be idiopathic or triggered by infections, sarcoidosis, Crohn’s disease, or other conditions (142). In rare cases, neutrophilic dermatoses or pregnancy can induce an EN eruption (143). The pathogenesis of EN is postulated to involve type III or IV hypersensitivity reactions. There is evidence suggesting a pathogenic role of neutrophils via their production of reactive oxygen intermediates, which induce tissue damage. (144-147). This process ultimately results in increased expression of adhesion molecules, VEGF, and cytokines (i.e., TNF- α , IL-6, and IL-8) both locally and systemically, facilitating immune cell migration to the SAT septae (148) (Fig 6A).

Erythema Induratum of Bazin (EIB) is a lobular panniculitis with lymphocytic vasculitis (149). It is recognized as a multifactorial disease associated with several triggers, including infection with tuberculosis (150). Similar to EN, type III and IV hypersensitivity reactions are hypothesized to play a role in EIB (149) (Fig 6B).

Lupus panniculitis is also a predominantly lobular process with lymphocytic vasculitis and mucin or calcium deposition. The infiltrating cells consist mainly of T-cells, B-cells, and macrophages (151). Partial deficiency in C4, which causes defective opsonization of immune complexes and disease pathogenesis has been linked to some cases of early-onset lupus (152).

6. Neoplastic processes in SAT

Beyond inflammatory processes, SAT can also harbor neoplasms, originating either from SAT-resident cells or secondary infiltration/metastasis. The most common primary SAT neoplasms are benign lipoma (153), while malignant liposarcoma is quite rare (154, 155).

Most primary cutaneous lymphomas, whether of the T cell- (CTCL) or B-cell-lineage (CBCL) (156-158), typically develop in the dermis and may subsequently extend to the SAT. In contrast, certain lymphomas, such as intravascular B-cell lymphoma, can have their primary origin in different target organs, including SAT (159). However, only a few lymphomas have their primary origin in SAT. Subcutaneous panniculitis-like T-cell lymphoma (SPTCL) specifically involves the subcutis, characterized by neoplastic T-cells rimming fat cells (157, 160). Two distinct types of SPTCL have been identified: (i) SPTCL with an α/β T-cell receptor (SPTCL-AB), which is characterized by a CD4-CD8+CD56- phenotype, and (ii) the highly aggressive SPTCL with a $\gamma\delta$ T-cell phenotype (SPTCL-GD), characterized by a CD4-CD8- phenotype with variable CD56 expression (160). An investigation of SPTCL skin samples showed significantly increased expression of the tolerogenic enzyme indoleamine 2,3-dioxygenase (IDO-1) and Th1-specific cytokine, INF γ (146). It is suggested that IDO-1 overexpression creates an immunosuppressive environment conducive to SPTCL development (146). However, the clonal specificity and underlying mechanisms of SPTCL development remain largely unknown.

7. Hereditary SAT diseases

Hereditary SAT disorders such as lipedema, multiple symmetric lipomatosis (MSL), Dercum’s disease, and familial partial lipodystrophy (FPLD) are characterized by a disproportional SAT hypertrophy that can be associated with systemic symptoms (161). Unlike obesity, hereditary SAT disorders are resistant to dietary

changes or physical exercise (161). Among them, lipedema is the most prevalent, marked by the enlargement and deposition of subcutaneous adipocytes (161-165). The occurrence of lipedema during hormonal changes in women, such as puberty, pregnancy, or menopause suggests a potential involvement of estrogen in its pathogenesis. However, the underlying pathomechanisms of lipedema development remain unclear (166). Clinical and histological studies do not show any morphological alterations of the vascular/lymphatic system (167). However, recent evidence suggests an immune-related origin, as observed through macrophage infiltration in lipedema AT (167). Furthermore, lipedema-derived ASCs express proliferative markers (Ki67 and CD34) and show an increased adipogenic differentiation potential in 2D cultures (168-170). The specific roles of these cells and their pathophysiological significance remain to be elucidated.

FPLD is a rarer hereditary lipodystrophy associated with the development of metabolic syndromes and cardiovascular disease in affected patients (171, 172). Investigating the pathomechanisms underlying hereditary lipodystrophies in the context of metabolic syndrome can contribute to a better understanding of obesity related metabolic diseases (table 3).

Conclusion and clinical perspectives:

There is a growing body of evidence highlighting the intricate and crucial immune functions of AT (25-27). Understanding the specific contributions of SAT in both homeostatic and pathological states remains a central challenge. Key questions need to be addressed to unravel immune loops between SAT and the skin or other organ systems.

Primarily, there is a need for a better understanding of the immunological reservoir within SAT in humans under homeostatic conditions. This necessitates a thorough characterization and functional exploration of both cellular (leukocytic and non-leukocytic) and molecular immune components within SAT. Also, characterizing the distinctions in SAT resident immune cells across various topographical locations of the body is crucial for elucidating their impact on skin homeostasis.

A pivotal aspect of this exploration is deciphering antigen presentation in SAT, including the identification of antigen-presenting cells (APC) and the nature of presented antigens. While AT-resident macrophages are the primary APC population in mice (173), obesity models have shown adipocytes expressing major histocompatibility complex II and activating CD4+ T-cells (174-176). The involvement of APCs beyond macrophages in humans remains unclear, necessitating further research to develop novel therapeutic strategies for SAT-based immune diseases.

In addition to comprehending immune dynamics under homeostatic conditions, it is crucial to delve into the pathomechanisms of SAT inflammation, using panniculitis as a representative model. The investigation of “immune loops” connecting SAT with the superficial skin layers or the systemic level, as observed in psoriasis and potentially other inflammatory conditions holds significant importance (177). Moreover, understanding the impact of SAT-based processes on both inflammatory and neoplastic conditions, as illustrated by data from breast cancer and SPTLC, is crucial (178-180). Additionally, the potential contribution of leaky barriers to increased inflammation in adipose tissue (181), along with the migration of proinflammatory cells (DC, Mac, CD1) from the adipose tissue to inflammatory organs, warrants exploration.

To investigate specific antigens and signaling pathways, and cell-cell interactions in various contexts, the development of full thickness skin models, comprising SAT, dermis and epidermis is warranted. A detailed understanding of SAT-based pathomechanisms facilitates the development of small molecule inhibitors targeting immunogenic antigens to mitigate inflammatory-driven complications. Moreover, considering the potential impact of obesity on these conditions, modulating SAT immune responses emerges as a promising avenue for developing targeted therapies against cutaneous / systemic immune-related diseases and obesity (Fig 7).

Table 1: Secreted antimicrobial molecules, adipokines and cytokines in obese adipose tissue

Antimicrobial peptides

—Cathelicidin	Anti-bacterial
Adipokines	Adipokines
—Leptin	Immunomodulatory effects
—Resistin	Immunomodulatory effects
—Adiponectin	Increase insulin sensitivity and glucose tolerance, anti-inflammatory
—Visfatin	Regulate insulin secretion, pro-inflammatory effects
Cytokines	Cytokines
— IL-6	Pro-inflammatory
—TNF- α	Pro-inflammatory
—IL-1 β	Pro-inflammatory
—MCP-1	Pro-inflammatory

IL-6: interleukin 6, TNF- α : tumor necrosis factor- α , IL-1 β : interleukin-1 β , MCP-1: monocyte chemoattractant protein1

Table 2: Classification of panniculitis

Predominantly septal panniculitis without vasculitis

<i>Type Etiology Pathogenesis Reference</i>	<i>Type Etiology Pathogenesis Reference</i>
Erythema nodosum	Idiopathic, Streptococcal infections, viral upper respiratory tract infection, Coccidioidomycosis
Scleroderma	Idiopathic, an overproduction and accumulation of collagen in connective tissue due to autoimmune disease
α 1-antitrypsin deficiency panniculitis	α 1-antitrypsin deficiency

Predominantly septal panniculitis with vasculitis

<i>Type Etiology Pathogenesis Reference</i>	<i>Type Etiology Pathogenesis Reference</i>
Cutaneous polyarteritis nodosa	Ideopathic, Group A β hemolytic Streptococcus infection, hepatitis B infection, influenza
Erythema nodosum leprosum	Mycobacterium leprae Type 2 reaction
Leukocytoclastic vasculitis	Infection, Inflammatory disease, medication or drugs
Superficial thrombophlebitis	Thrombosis in superficial vein, trauma, venos stasis, malignancy, pregnancy,

Lobular and mixed septal-lobular panniculitis without vasculitis

<i>Type Etiology Pathogenesis Reference</i>	<i>Type Etiology Pathogenesis Reference</i>	<i>Type Etiology Pathogenesis Reference</i>	<i>Type Etiology Pathogenesis Reference</i>
Lupus panniculitis (lupus profundus)	Autoimmune connective tissue disease	Infiltration of T-lymphocytes and macrophages, type III hypersensitivity in patients with C4 deficiency, interferon-driven Th1 immune response	(188)
Sclerosing panniculitis (Lipodermatosclerosis)	Venous insufficiency, obesity	Lymphocytic infiltration, lipomembranous changes and thickened membrane	(189, 190)

Sclerema neonatorum	Hypothermia, asphyxia, dehydration	Inflammation sparse to absent, crystallization of fat due to an increased saturated : unsaturated fatty acid ratio	(191)
Neonatal subcutaneous fat necrosis	Hypercalcemia, hypothermia, hypoglycemia	Infiltration of neutrophils, lymphocytes and macrophages	(192)
Pancreatic panniculitis	Pancreatic disorders	Elevated enzyme levels (lipase, amylase and trypsin), infiltration of neutrophils, macrophages and multinucleated giant cells	(193)
Infection-induced panniculitis	Infectious agents such as “bacteria, mycobacteria, coxiella, borrelia, fungi and helminths”, vascular proliferation, hemorrhage, necrosis	Neutrophilic infiltration	(194)
Traumatic panniculitis	External injury such as cold in infants, injections, radiation in deep tissue, self-injection of oily materials on the male genitalia, adipocyte necrosis	Infiltration of lymphocytes, neutrophils, foamy macrophages, plasma cells, eosinophils	(195-197)
Factitious panniculitis	Self-induction of unknown substances	Unknown	(198)
Subcutaneous sarcoidosis	Systemic sarcoidosis	Granulomatous infiltration	(199)
Post-steroid panniculitis	Follows rapid corticosteroid withdrawal	Neutrophilic infiltration	(200)
Panniculitis like T-cell lymphoma	Malignancy-related panniculitis-like infiltrates	Neoplastic T-cells (CD8+ cells) and macrophages infiltration	(201)
Weber-Christian Disease	Idiopathic nodular panniculitis	Unknown	

Lobular and mixed septal-lobular panniculitis with vasculitis

<i>Type</i>	<i>Etiology</i>	<i>Pathogenesis</i>	<i>Reference</i>	<i>Type</i>	<i>Etiology</i>	<i>Pathogenesis</i>	<i>Reference</i>	<i>Type</i>	<i>Etiology</i>	<i>Pathogenesis</i>	<i>Reference</i>
Erythema induratum of Bazin				« Id-reaction » to mycobacterium tuberculosis infection				Hypersensitivity type III			
Neutrophilic lobular panniculitis				Hematologic malignancies, rheumatoid arthritis				Predominant neutrophils			
Erythema nodosum leprosum				lepomatous leprosy, reaction to mycobacterium leprae				Type II reaction, neutrophils			

Table 3: Hereditary SAT disease characteristics

Hereditary SAT	Inheritance pattern	Associated comorbidities
Lipedema	Autosomal dominant, receive penetrance	Painful SAT, depression, joint pain, arthralgia
MSL	Autosomal dominant or recessive	Hyperlipidemia, hyperuricemia, hypothyroidism
Dercum's disease	Autosomal dominant	Gastrointestinal problems, joint pain, vasculopathy
Familial partial lipodystrophy	Autosomal dominant	Metabolic syndrome, T2D, insuline resist
congenital generalized lipodystrophy	Autosomal recessive	Metabolic syndrome, T2D, hepatosplenomegaly

Figures

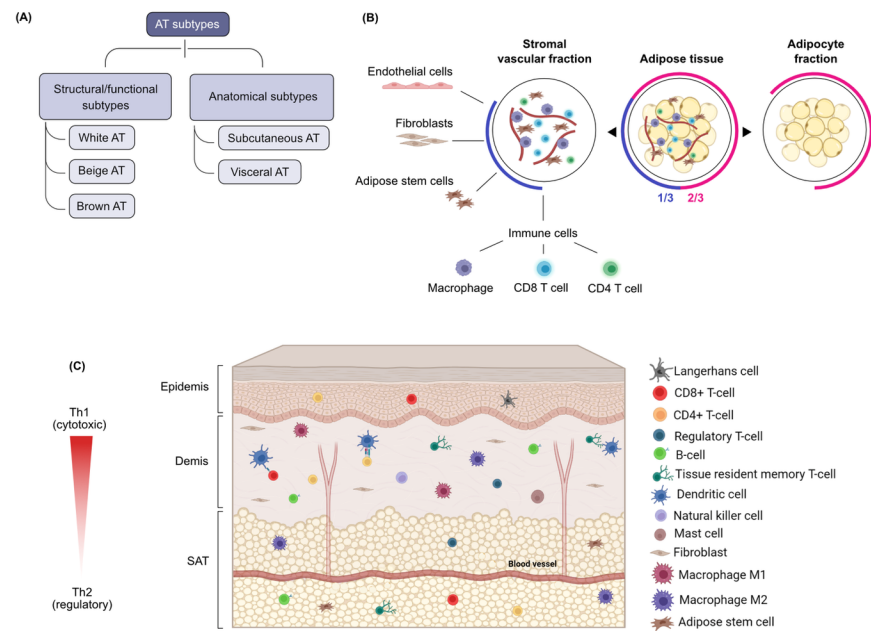


Fig 1: Structural and cellular composition of adipose tissue (AT).

(A) Different structural and anatomical subtypes of AT (B) Cellular components of AT. AT consists of a 2/3 adipocyte fraction and a 1/3 of stromal vascular fraction (C) Immune cells in the three-layered cutaneous integument, which consists of epidermis, dermis and subcutaneous adipose tissue (SAT).

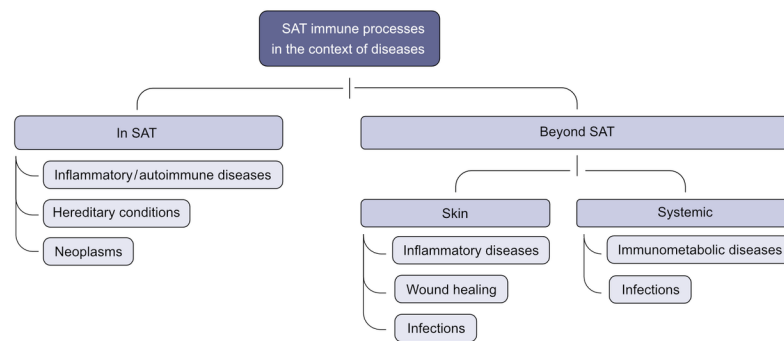


Fig 2: Subcutaneous adipose tissue (SAT)-mediated immune processes in clinical conditions.

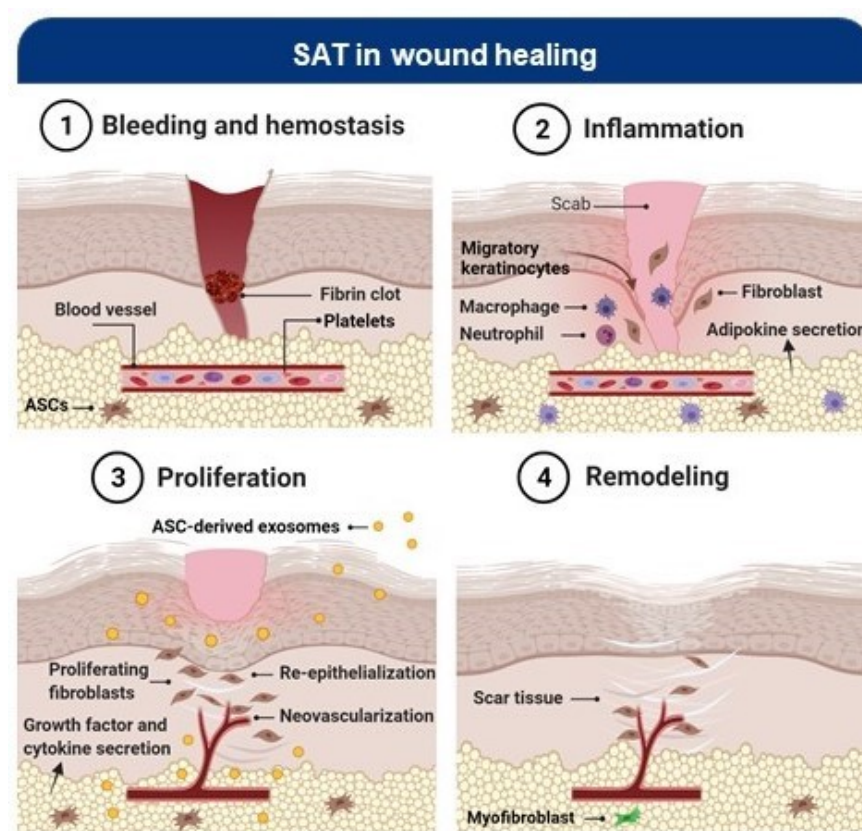


Fig 3: SAT in wound healing. Wound healing consists of several regenerative phases: (1) Bleeding and hemostasis lead to platelet aggregation and coagulation (2) Inflammatory cells, such as neutrophils, macrophages are recruited to the site of injury to clear debris and microbes. Fibroblast and macrophages support the migration of keratinocytes and adipocytes secrete adipokines such as interleukins, leptin and adiponectin. (3) Secretion of growth factors and cytokines from adipocytes promotes fibroblast proliferation, re-epithelialization and neovascularization. Administration of ASC-derived exosomes can promote angiogenesis and re-epithelization (4) Adipocytes de-differentiate to myofibroblasts, which contributed to

wound repair by producing extracellular matrix (ECM) which serve as scaffold.

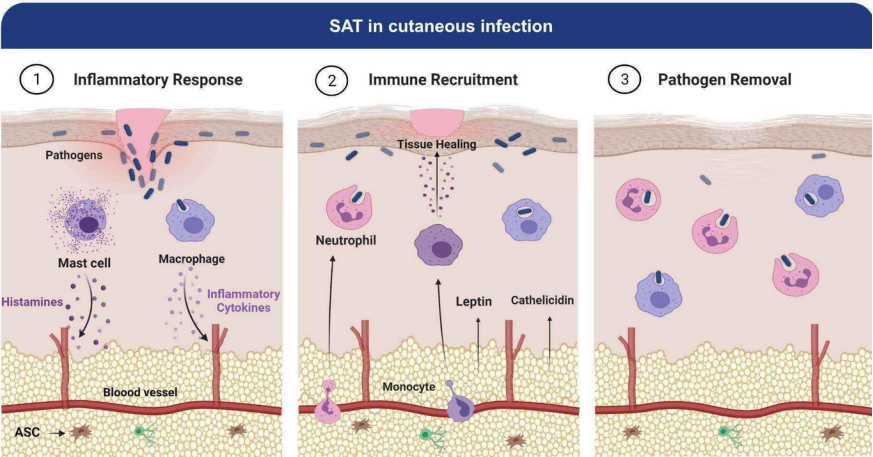


Fig 4: SAT in cutaneous infection. (1) Inflammatory cytokines and histamine are released by macrophages and recruit monocytes and neutrophils to the site of infection. (2) Adipocytes secrete antimicrobial peptide cathelicidin and leptin adipokine (3) Pathogens are removed from the site of infection.

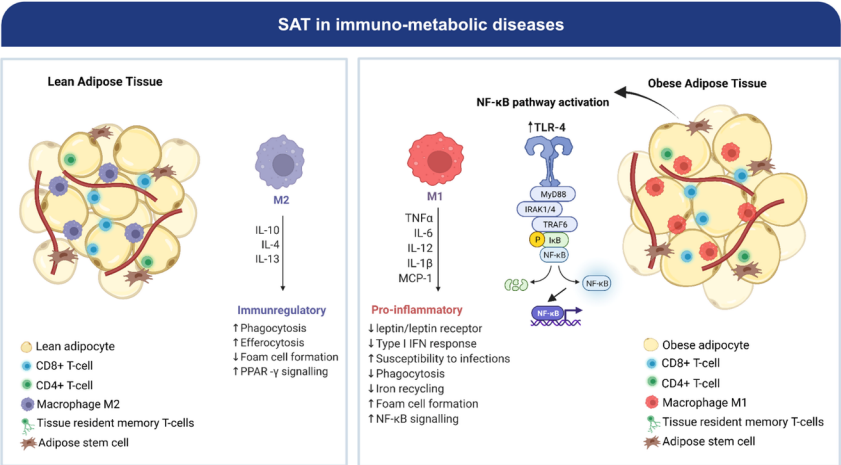
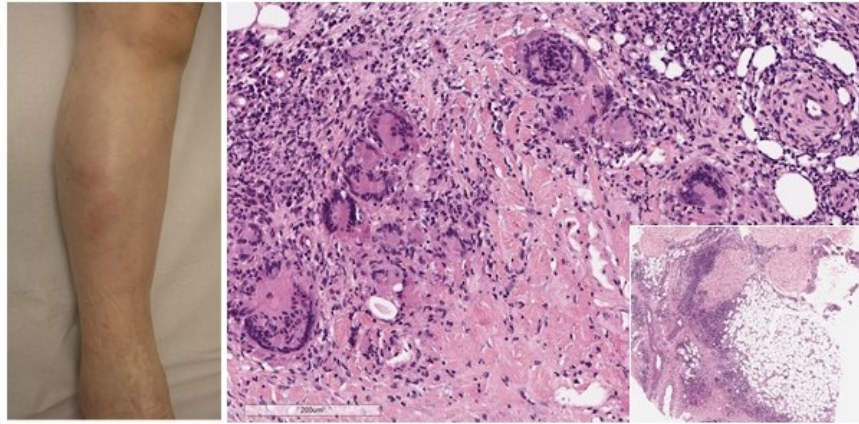


Fig 5: Lean and obese adipose tissue immune function. In obesity macrophages are polarized towards M1 phenotype with pro-inflammatory properties, while M2 macrophages with immunoregulatory properties are predominant in lean AT. In obese AT, NFκ-B signaling pathway will be activated upon overexpression of Toll-like receptor 4 (TLR-4). Upon activation of NFκ-B signaling, monocyte chemoattractant protein-1 (MCP-1) and pro-inflammatory markers such as interleukins β , 6, 12 (IL1 β IL-6, IL-12) and tumor necrosis factor- α (TNF- α) will be expressed.

Clinical and histopathological images of panniculitis

(A) Erythema nodosum



(B) Erythema induratum

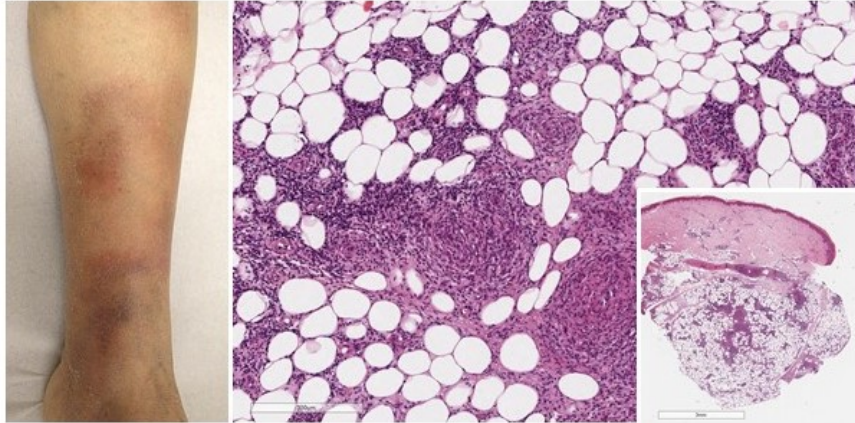


Fig 6: Clinical and histopathological images of panniculitis. **(A)** Septal panniculitis (Erythema nodosum). H&E stain shows the inflammatory infiltrate is predominately confined to the thickened and fibrotic septa of the subcutis. The inflammatory infiltrate is mostly lymphocytic, with admixture of eosinophilic granulocytes, plasma cells and many multinucleate giant cells. The vessels are inconspicuous. **(B)** Lobular panniculitis (Erythema induratum). H&E stain shows nodular vasculitis with granuloma formation and vasculitis.

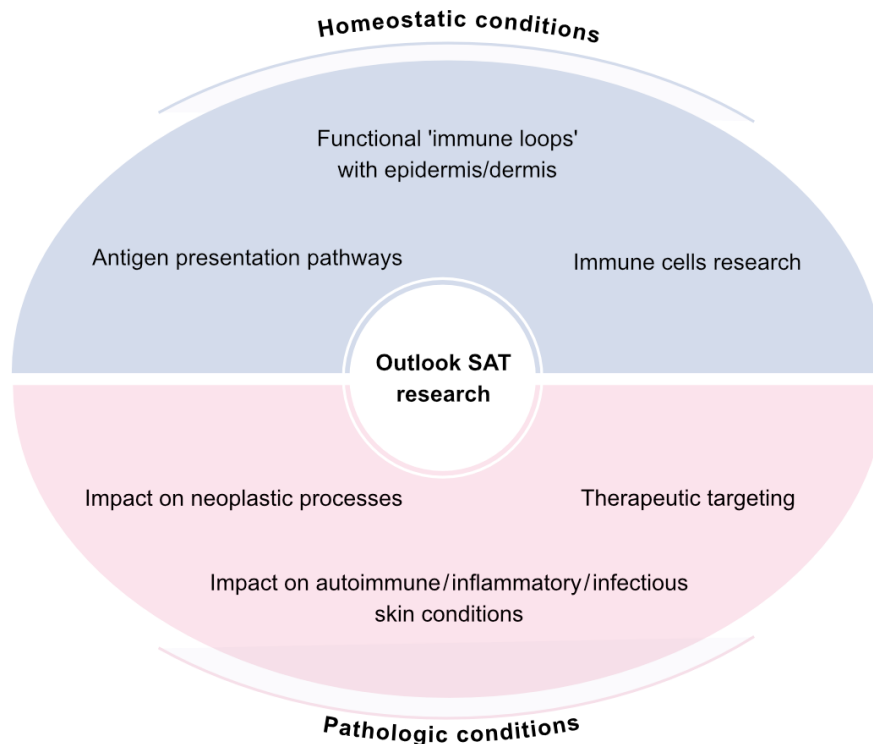


Fig 7: Future perspectives in SAT translational / clinical research.

References

1. Ottaviani E, Malagoli D, Franceschi C. The evolution of the adipose tissue: A neglected enigma. *General and Comparative Endocrinology*. 2011;174(1):1-4.
2. Scheja L, Heeren J. The endocrine function of adipose tissues in health and cardiometabolic disease. *Nature Reviews Endocrinology*. 2019;15(9):507-24.
3. Khan S, Chan YT, Revelo XS, Winer DA. The Immune Landscape of Visceral Adipose Tissue During Obesity and Aging. *Frontiers in endocrinology*. 2020;11:267-.
4. Singh GM, Danaei G, Farzadfar F, Stevens GA, Woodward M, Wormser D, et al. The age-specific quantitative effects of metabolic risk factors on cardiovascular diseases and diabetes: a pooled analysis. *PloS one*. 2013;8(7):e65174.
5. Collins KH, Lenz KL, Pollitt EN, Ferguson D, Hutson I, Springer LE, et al. Adipose tissue is a critical regulator of osteoarthritis. *Proc Natl Acad Sci U S A*. 2021;118(1).
6. Leung DYM, Berdyshev E, Goleva E. Cutaneous barrier dysfunction in allergic diseases. *Journal of Allergy and Clinical Immunology*. 2020;145(6):1485-97.
7. Nyberg ST, Batty GD, Pentti J, Virtanen M, Alfredsson L, Fransson EI, et al. Obesity and loss of disease-free years owing to major non-communicable diseases: a multicohort study. *The Lancet Public Health*. 2018;3(10):e490-e7.
8. Campanella G, Gunter MJ, Polidoro S, Krogh V, Palli D, Panico S, et al. Epigenome-wide association study of adiposity and future risk of obesity-related diseases. *International Journal of Obesity*. 2018;42(12):2022-35.
9. Jensen P, Skov L. Psoriasis and Obesity. *Dermatology*. 2016;232(6):633-9.

10. Coimbra S, Oliveira H, Reis F, Belo L, Rocha S, Quintanilha A, et al. Circulating levels of adiponectin, oxidized LDL and C-reactive protein in Portuguese patients with psoriasis vulgaris, according to body mass index, severity and duration of the disease. *J Dermatol Sci*. 2009;55(3):202-4.
11. Kromann CB, Ibler KS, Kristiansen VB, Jemec GB. The influence of body weight on the prevalence and severity of hidradenitis suppurativa. *Acta Derm Venereol*. 2014;94(5):553-7.
12. Ibrahim MM. Subcutaneous and visceral adipose tissue: structural and functional differences. *Obes Rev*. 2010;11(1):11-8.
13. Chen SX, Zhang L-J, Gallo RL. Dermal White Adipose Tissue: A Newly Recognized Layer of Skin Innate Defense. *Journal of Investigative Dermatology*. 2019;139(5):1002-9.
14. Zhang Z, Shao M, Hepler C, Zi Z, Zhao S, An YA, et al. Dermal adipose tissue has high plasticity and undergoes reversible dedifferentiation in mice. *The Journal of Clinical Investigation*. 2019;129(12):5327-42.
15. Driskell RR, Jahoda CAB, Chuong C-M, Watt FM, Horsley V. Defining dermal adipose tissue. *Experimental dermatology*. 2014;23(9):629-31.
16. Jung SM, Sanchez-Gurmaches J, Guertin DA. Brown Adipose Tissue Development and Metabolism. *Handb Exp Pharmacol*. 2019;251:3-36.
17. Whitehead A, Krause FN, Moran A, MacCannell ADV, Scragg JL, McNally BD, et al. Brown and beige adipose tissue regulate systemic metabolism through a metabolite interorgan signaling axis. *Nature Communications*. 2021;12(1):1905.
18. Sidossis L, Kajimura S. Brown and beige fat in humans: thermogenic adipocytes that control energy and glucose homeostasis. *J Clin Invest*. 2015;125(2):478-86.
19. Cai X, Lin Y, Hauschka PV, Grottkau BE. Adipose stem cells originate from perivascular cells. *Biol Cell*. 2011;103(9):435-47.
20. Si Z, Wang X, Sun C, Kang Y, Xu J, Wang X, Hui Y. Adipose-derived stem cells: Sources, potency, and implications for regenerative therapies. *Biomedicine & Pharmacotherapy*. 2019;114:108765.
21. Andersen E, Ingerslev LR, Fabre O, Donkin I, Altıntaş A, Versteyhe S, et al. Preadipocytes from obese humans with type 2 diabetes are epigenetically reprogrammed at genes controlling adipose tissue function. *International Journal of Obesity*. 2019;43(2):306-18.
22. Sengenès C, Lohmède K, Zakaroff-Girard A, Busse R, Bouloumié A. Preadipocytes in the human subcutaneous adipose tissue display distinct features from the adult mesenchymal and hematopoietic stem cells. *Journal of Cellular Physiology*. 2005;205(1):114-22.
23. Rodeheffer MS, Birsoy K, Friedman JM. Identification of white adipocyte progenitor cells in vivo. *Cell*. 2008;135(2):240-9.
24. Zhang R, Gao Y, Zhao X, Gao M, Wu Y, Han Y, et al. FSP1-positive fibroblasts are adipogenic niche and regulate adipose homeostasis. *PLoS Biol*. 2018;16(8):e2001493.
25. Anderson EK, Gutierrez DA, Hasty AH. Adipose tissue recruitment of leukocytes. *Curr Opin Lipidol*. 2010;21(3):172-7.
26. Brüggén MC, Strobl J, Koszik F, Naito R, Vierhapper M, Li N, et al. Subcutaneous White Adipose Tissue of Healthy Young Individuals Harbors a Leukocyte Compartment Distinct from Skin and Blood. *J Invest Dermatol*. 2019;139(9):2052-5.e7.
27. Li Y, Yun K, Mu R. A review on the biology and properties of adipose tissue macrophages involved in adipose tissue physiological and pathophysiological processes. *Lipids in Health and Disease*. 2020;19(1):164.

28. Bolus WR, Hasty AH. Contributions of innate type 2 inflammation to adipose function. *Journal of Lipid Research*. 2019;60(10):1698-709.
29. Brestoff JR, Artis D. Immune regulation of metabolic homeostasis in health and disease. *Cell*. 2015;161(1):146-60.
30. Zhang LJ, Guerrero-Juarez CF, Hata T, Bapat SP, Ramos R, Plikus MV, Gallo RL. Innate immunity. Dermal adipocytes protect against invasive *Staphylococcus aureus* skin infection. *Science (New York, NY)*. 2015;347(6217):67-71.
31. Nguyen AV, Soulika AM. The Dynamics of the Skin's Immune System. *International journal of molecular sciences*. 2019;20(8):1811.
32. Shook BA, Wasko RR, Mano O, Rutenberg-Schoenberg M, Rudolph MC, Zirak B, et al. Dermal Adipocyte Lipolysis and Myofibroblast Conversion Are Required for Efficient Skin Repair. *Cell Stem Cell*. 2020;26(6):880-95.e6.
33. Shook B, Xiao E, Kumamoto Y, Iwasaki A, Horsley V. CD301b+ Macrophages Are Essential for Effective Skin Wound Healing. *J Invest Dermatol*. 2016;136(9):1885-91.
34. Zhang L-j, Guerrero-Juarez CF, Hata T, Bapat SP, Ramos R, Plikus MV, Gallo RL. Innate immunity. Dermal adipocytes protect against invasive *Staphylococcus aureus* skin infection. *Science (New York, NY)*. 2015;347(6217):67-71.
35. Wong Y, Nakamizo S, Tan KJ, Kabashima K. An Update on the Role of Adipose Tissues in Psoriasis. *Frontiers in Immunology*. 2019;10:1507.
36. Raud B, McGuire PJ, Jones RG, Sparwasser T, Berod L. Fatty acid metabolism in CD8+ T cell memory: Challenging current concepts. *Immunological reviews*. 2018;283(1):213-31.
37. Rodrigues M, Kosaric N, Bonham CA, Gurtner GC. Wound Healing: A Cellular Perspective. *Physiological Reviews*. 2018;99(1):665-706.
38. Salgado AJ, Reis RL, Sousa NJ, Gimble JM. Adipose tissue derived stem cells secretome: soluble factors and their roles in regenerative medicine. *Curr Stem Cell Res Ther*. 2010;5(2):103-10.
39. López JF, Sarkanen JR, Huttala O, Kaartinen IS, Kuokkanen HO, Ylikomi T. Adipose tissue extract shows potential for wound healing: in vitro proliferation and migration of cell types contributing to wound healing in the presence of adipose tissue preparation and platelet rich plasma. *Cytotechnology*. 2018;70(4):1193-204.
40. Cui L, Yin S, Liu W, Li N, Zhang W, Cao Y. Expanded adipose-derived stem cells suppress mixed lymphocyte reaction by secretion of prostaglandin E2. *Tissue Eng*. 2007;13(6):1185-95.
41. Wang M, Crisostomo PR, Herring C, Meldrum KK, Meldrum DR. Human progenitor cells from bone marrow or adipose tissue produce VEGF, HGF, and IGF-I in response to TNF by a p38 MAPK-dependent mechanism. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*. 2006;291(4):R880-R4.
42. Merrick D, Seale P. Skinny Fat Cells Stimulate Wound Healing. *Cell Stem Cell*. 2020;26(6):801-3.
43. Raghuram AC, Yu RP, Lo AY, Sung CJ, Bircan M, Thompson HJ, Wong AK. Role of stem cell therapies in treating chronic wounds: A systematic review. *World J Stem Cells*. 2020;12(7):659-75.
44. Fujiwara O, Prasai A, Perez-Bello D, El Ayadi A, Petrov IY, Esenaliev RO, et al. Adipose-derived stem cells improve grafted burn wound healing by promoting wound bed blood flow. *Burns & Trauma*. 2020;8.
45. Kuo Y-R, Wang C-T, Cheng J-T, Kao G-S, Chiang Y-C, Wang C-J. Adipose-Derived Stem Cells Accelerate Diabetic Wound Healing through the Induction of Autocrine and Paracrine Effects. *Cell Transplantation*. 2016;25(1):71-81.

46. Kokai LE, Marra K, Rubin JP. Adipose stem cells: biology and clinical applications for tissue repair and regeneration. *Translational Research*. 2014;163(4):399-408.
47. Gimble JM, Guilak F, Bunnell BA. Clinical and preclinical translation of cell-based therapies using adipose tissue-derived cells. *Stem Cell Res Ther*. 2010;1(2):19.
48. An Y, Lin S, Tan X, Zhu S, Nie F, Zhen Y, et al. Exosomes from adipose-derived stem cells and application to skin wound healing. *Cell Prolif*. 2021;54(3):e12993.
49. Golchin A, Hosseinzadeh S, Ardeshtyrlajimi A. The exosomes released from different cell types and their effects in wound healing. *Journal of Cellular Biochemistry*. 2018;119(7):5043-52.
50. Baglio SR, Pegtel DM, Baldini N. Mesenchymal stem cell secreted vesicles provide novel opportunities in (stem) cell-free therapy. *Frontiers in physiology*. 2012;3:359.
51. Hu L, Wang J, Zhou X, Xiong Z, Zhao J, Yu R, et al. Exosomes derived from human adipose mesenchymal stem cells accelerates cutaneous wound healing via optimizing the characteristics of fibroblasts. *Scientific Reports*. 2016;6(1):32993.
52. Wang L, Hu L, Zhou X, Xiong Z, Zhang C, Shehada HMA, et al. Exosomes secreted by human adipose mesenchymal stem cells promote scarless cutaneous repair by regulating extracellular matrix remodelling. *Scientific Reports*. 2017;7(1):13321.
53. Mahlaköiv T, Flamar AL, Johnston LK, Moriyama S, Putzel GG, Bryce PJ, Artis D. Stromal cells maintain immune cell homeostasis in adipose tissue via production of interleukin-33. *Sci Immunol*. 2019;4(35).
54. Ronti T, Lupattelli G, Mannarino E. The endocrine function of adipose tissue: an update. *Clinical Endocrinology*. 2006;64(4):355-65.
55. Rajesh Y, Sarkar D. Association of Adipose Tissue and Adipokines with Development of Obesity-Induced Liver Cancer. *International Journal of Molecular Sciences*. 2021;22(4).
56. Bates SH, Stearns WH, Dundon TA, Schubert M, Tso AW, Wang Y, et al. STAT3 signalling is required for leptin regulation of energy balance but not reproduction. *Nature*. 2003;421(6925):856-9.
57. Procaccini C, Jirillo E, Matarese G. Leptin as an immunomodulator. *Molecular Aspects of Medicine*. 2012;33(1):35-45.
58. Klein J, Perwitz N, Kraus D, Fasshauer M. Adipose tissue as source and target for novel therapies. *Trends in Endocrinology & Metabolism*. 2006;17(1):26-32.
59. Mancuso P, Gottschalk A, Phare SM, Peters-Golden M, Lukacs NW, Huffnagle GB. Leptin-deficient mice exhibit impaired host defense in Gram-negative pneumonia. *J Immunol*. 2002;168(8):4018-24.
60. Maurya R, Bhattacharya P, Dey R, Nakhasi HL. Leptin Functions in Infectious Diseases. *Frontiers in Immunology*. 2018;9(2741).
61. Milner JJ, Beck MA. The impact of obesity on the immune response to infection. *Proceedings of the Nutrition Society*. 2012;71(2):298-306.
62. Terán-Cabanillas E, Hernández J. Role of Leptin and SOCS3 in Inhibiting the Type I Interferon Response During Obesity. *Inflammation*. 2017;40(1):58-67.
63. Schaab M, Kratzsch J. The soluble leptin receptor. *Best Pract Res Clin Endocrinol Metab*. 2015;29(5):661-70.
64. Singanayagam A, Glanville N, Cuthbertson L, Bartlett NW, Finney LJ, Turek E, et al. Inhaled corticosteroid suppression of cathelicidin drives dysbiosis and bacterial infection in chronic obstructive pulmonary disease. *Science Translational Medicine*. 2019;11(507):eaav3879.
65. Boman HG. Antibacterial peptides: basic facts and emerging concepts. *J Intern Med*. 2003;254(3):197-215.

66. Han SJ, Glatman Zaretsky A, Andrade-Oliveira V, Collins N, Dzutsev A, Shaik J, et al. White Adipose Tissue Is a Reservoir for Memory T Cells and Promotes Protective Memory Responses to Infection. *Immunity*. 2017;47(6):1154-68.e6.
67. Reina-Campos M, Scharping NE, Goldrath AW. CD8+ T cell metabolism in infection and cancer. *Nature Reviews Immunology*. 2021.
68. O'Shea D, Corrigan M, Dunne MR, Jackson R, Woods C, Gaoatswe G, et al. Changes in human dendritic cell number and function in severe obesity may contribute to increased susceptibility to viral infection. *International Journal of Obesity*. 2013;37(11):1510-3.
69. Smith AG, Sheridan PA, Tseng RJ, Sheridan JF, Beck MA. Selective impairment in dendritic cell function and altered antigen-specific CD8+ T-cell responses in diet-induced obese mice infected with influenza virus. *Immunology*. 2009;126(2):268-79.
70. Lynch L, Nowak M, Varghese B, Clark J, Hogan AE, Toxavidis V, et al. Adipose tissue invariant NKT cells protect against diet-induced obesity and metabolic disorder through regulatory cytokine production. *Immunity*. 2012;37(3):574-87.
71. Viel S, Besson L, Charrier E, Marçais A, Disse E, Bienvenu J, et al. Alteration of Natural Killer cell phenotype and function in obese individuals. *Clin Immunol*. 2017;177:12-7.
72. Michelet X, Dyck L, Hogan A, Loftus RM, Duquette D, Wei K, et al. Metabolic reprogramming of natural killer cells in obesity limits antitumor responses. *Nat Immunol*. 2018;19(12):1330-40.
73. O'Shea D, Hogan AE. Dysregulation of Natural Killer Cells in Obesity. *Cancers (Basel)*. 2019;11(4).
74. Smith AG, Sheridan PA, Tseng RJ, Sheridan JF, Beck MA. Selective impairment in dendritic cell function and altered antigen-specific CD8+ T-cell responses in diet-induced obese mice infected with influenza virus. *Immunology*. 2009;126(2):268-79.
75. Kanneganti TD, Dixit VD. Immunological complications of obesity. *Nat Immunol*. 2012;13(8):707-12.
76. Misumi I, Starmer J, Uchimura T, Beck MA, Magnuson T, Whitmire JK. Obesity Expands a Distinct Population of T Cells in Adipose Tissue and Increases Vulnerability to Infection. *Cell Reports*. 2019;27(2):514-24.e5.
77. Dhurandhar NV. Infections and body weight: an emerging relationship? *International Journal of Obesity*. 2002;26(6):745-6.
78. Pasarica M, Shin AC, Yu M, Ou Yang HM, Rathod M, Jen KL, et al. Human adenovirus 36 induces adiposity, increases insulin sensitivity, and alters hypothalamic monoamines in rats. *Obesity (Silver Spring)*. 2006;14(11):1905-13.
79. Dhurandhar NV, Whigham LD, Abbott DH, Schultz-Darken NJ, Israel BA, Bradley SM, et al. Human adenovirus Ad-36 promotes weight gain in male rhesus and marmoset monkeys. *J Nutr*. 2002;132(10):3155-60.
80. Rudrapatna S, Bhatt M, Wang KW, Bierbrier R, Wang PW, Banfield L, et al. Obesity and muscle-macrophage crosstalk in humans and mice: A systematic review. *Obes Rev*. 2019;20(11):1572-96.
81. Bray GA, Kim KK, Wilding JPH, on behalf of the World Obesity F. Obesity: a chronic relapsing progressive disease process. A position statement of the World Obesity Federation. *Obesity Reviews*. 2017;18(7):715-23.
82. Kahn SE, Hull RL, Utzschneider KM. Mechanisms linking obesity to insulin resistance and type 2 diabetes. *Nature*. 2006;444(7121):840-6.
83. Klingberg E, Bilberg A, Björkman S, Hedberg M, Jacobsson L, Forsblad-d'Elia H, et al. Weight loss improves disease activity in patients with psoriatic arthritis and obesity: an interventional study. *Arthritis Research & Therapy*. 2019;21(1):17.

84. Vlietstra L, Stebbings S, Meredith-Jones K, Abbott JH, Treharne GJ, Waters DL. Sarcopenia in osteoarthritis and rheumatoid arthritis: The association with self-reported fatigue, physical function and obesity. *PLOS ONE*. 2019;14(6):e0217462.
85. Vekic J, Zeljkovic A, Stefanovic A, Jelic-Ivanovic Z, Spasojevic-Kalimanovska V. Obesity and dyslipidemia. *Metabolism*. 2019;92:71-81.
86. Koliaki C, Liatis S, Kokkinos A. Obesity and cardiovascular disease: revisiting an old relationship. *Metabolism*. 2019;92:98-107.
87. Avgerinos KI, Spyrou N, Mantzoros CS, Dalamaga M. Obesity and cancer risk: Emerging biological mechanisms and perspectives. *Metabolism*. 2019;92:121-35.
88. Stone TW, McPherson M, Gail Darlington L. Obesity and Cancer: Existing and New Hypotheses for a Causal Connection. *EBioMedicine*. 2018;30:14-28.
89. Stinkens R, Goossens GH, Jocken JW, Blaak EE. Targeting fatty acid metabolism to improve glucose metabolism. *Obes Rev*. 2015;16(9):715-57.
90. van der Kolk BW, Kalafati M, Adriaens M, van Greevenbroek MMJ, Vogelzangs N, Saris WHM, et al. Subcutaneous Adipose Tissue and Systemic Inflammation Are Associated With Peripheral but Not Hepatic Insulin Resistance in Humans. *Diabetes*. 2019;68(12):2247-58.
91. Keophiphath M, Achard V, Henegar C, Rouault C, Clément K, Lacasa D. Macrophage-secreted factors promote a profibrotic phenotype in human preadipocytes. *Mol Endocrinol*. 2009;23(1):11-24.
92. Tanaka M, Ikeda K, Suganami T, Komiya C, Ochi K, Shirakawa I, et al. Macrophage-inducible C-type lectin underlies obesity-induced adipose tissue fibrosis. *Nature Communications*. 2014;5(1):4982.
93. Wang H, Shen L, Sun X, Liu F, Feng W, Jiang C, et al. Adipose group 1 innate lymphoid cells promote adipose tissue fibrosis and diabetes in obesity. *Nature Communications*. 2019;10(1):3254.
94. Voss K, Hong HS, Bader JE, Sugiura A, Lyssiotis CA, Rathmell JC. A guide to interrogating immunometabolism. *Nature Reviews Immunology*. 2021.
95. Makowski L, Chaib M, Rathmell JC. Immunometabolism: From basic mechanisms to translation. *Immunological Reviews*. 2020;295(1):5-14.
96. Unamuno X, Gómez-Ambrosi J, Rodríguez A, Becerril S, Frühbeck G, Catalán V. Adipokine dysregulation and adipose tissue inflammation in human obesity. *European Journal of Clinical Investigation*. 2018;48(9):e12997.
97. Jung UJ, Choi MS. Obesity and its metabolic complications: the role of adipokines and the relationship between obesity, inflammation, insulin resistance, dyslipidemia and nonalcoholic fatty liver disease. *Int J Mol Sci*. 2014;15(4):6184-223.
98. Procaccini C, Jirillo E, Matarese G. Leptin as an immunomodulator. *Mol Aspects Med*. 2012;33(1):35-45.
99. Travers RL, Motta AC, Betts JA, Bouloumié A, Thompson D. The impact of adiposity on adipose tissue-resident lymphocyte activation in humans. *International Journal of Obesity*. 2015;39(5):762-9.
100. Strissel KJ, DeFuria J, Shaul ME, Bennett G, Greenberg AS, Obin MS. T-Cell Recruitment and Th1 Polarization in Adipose Tissue During Diet-Induced Obesity in C57BL/6 Mice. *Obesity*. 2010;18(10):1918-25.
101. Wang Q, Wu H. T Cells in Adipose Tissue: Critical Players in Immunometabolism. *Frontiers in immunology*. 2018;9:2509-.
102. Vitseva OI, Tanriverdi K, Tchkonina TT, Kirkland JL, McDonnell ME, Apovian CM, et al. Inducible Toll-like receptor and NF-kappaB regulatory pathway expression in human adipose tissue. *Obesity (Silver Spring)*. 2008;16(5):932-7.

103. Schäffler A, Schölmerich J. Innate immunity and adipose tissue biology. *Trends in Immunology*. 2010;31(6):228-35.
104. Kopp A, Gross P, Falk W, Bala M, Weigert J, Buechler C, et al. Fatty acids as metabolic mediators in innate immunity. *Eur J Clin Invest*. 2009;39(10):924-33.
105. Schaeffler A, Gross P, Buettner R, Bollheimer C, Buechler C, Neumeier M, et al. Fatty acid-induced induction of Toll-like receptor-4/nuclear factor-kappaB pathway in adipocytes links nutritional signalling with innate immunity. *Immunology*. 2009;126(2):233-45.
106. Cullberg KB, Larsen JØ, Pedersen SB, Richelsen B. Effects of LPS and dietary free fatty acids on MCP-1 in 3T3-L1 adipocytes and macrophages in vitro. *Nutr Diabetes*. 2014;4(3):e113-e.
107. Russo L, Lumeng CN. Properties and functions of adipose tissue macrophages in obesity. *Immunology*. 2018;155(4):407-17.
108. Sharma VM, Puri V. Mechanism of TNF- α -induced lipolysis in human adipocytes uncovered. *Obesity*. 2016;24(5):990-.
109. Sethi JK, Hotamisligil GS. Metabolic Messengers: tumour necrosis factor. *Nature Metabolism*. 2021;3(10):1302-12.
110. Du X, Liu M, Tai W, Yu H, Hao X, Looor JJ, et al. Tumor necrosis factor- α promotes lipolysis and reduces insulin sensitivity by activating nuclear factor kappa B and c-Jun N-terminal kinase in primary bovine adipocytes. *Journal of Dairy Science*. 2022;105(10):8426-38.
111. Xia W, Veeragandham P, Cao Y, Xu Y, Rhyne TE, Qian J, et al. Obesity causes mitochondrial fragmentation and dysfunction in white adipocytes due to RalA activation. *Nature Metabolism*. 2024.
112. Heinonen S, Buzkova J, Muniandy M, Kaksonen R, Ollikainen M, Ismail K, et al. Impaired Mitochondrial Biogenesis in Adipose Tissue in Acquired Obesity. *Diabetes*. 2015;64(9):3135-45.
113. Chavakis T, Alexaki VI, Ferrante AW. Macrophage function in adipose tissue homeostasis and metabolic inflammation. *Nature Immunology*. 2023;24(5):757-66.
114. Becker M, Dirschl SM, Scherm MG, Serr I, Daniel C. Niche-specific control of tissue function by regulatory T cells—Current challenges and perspectives for targeting metabolic disease. *Cell Metabolism*. 2024.
115. Becker M, Levings MK, Daniel C. Adipose-tissue regulatory T cells: Critical players in adipose-immune crosstalk. *European Journal of Immunology*. 2017;47(11):1867-74.
116. Clemente-Suárez VJ, Redondo-Flórez L, Beltrán-Velasco AI, Martín-Rodríguez A, Martínez-Guardado I, Navarro-Jiménez E, et al. The Role of Adipokines in Health and Disease. *Biomedicines*. 2023;11(5):1290.
117. Zorena K, Jachimowicz-Duda O, Ślezak D, Robakowska M, Mrugacz M. Adipokines and Obesity. Potential Link to Metabolic Disorders and Chronic Complications. *Int J Mol Sci*. 2020;21(10).
118. Barral DC, Brenner MB. CD1 antigen presentation: how it works. *Nat Rev Immunol*. 2007;7(12):929-41.
119. de Jong A, Peña-Cruz V, Cheng T-Y, Clark RA, Van Rhijn I, Moody DB. CD1a-autoreactive T cells are a normal component of the human $\alpha\beta$ T cell repertoire. *Nature immunology*. 2010;11(12):1102-9.
120. Frasca D, Diaz A, Romero M, Garcia D, Jayram D, Thaller S, et al. Identification and Characterization of Adipose Tissue-Derived Human Antibodies With "Anti-self" Specificity. *Frontiers in immunology*. 2020;11:392-.
121. Cotton RN, Wegrecki M, Cheng T-Y, Chen Y-L, Veerapen N, Le Nours J, et al. CD1a selectively captures endogenous cellular lipids that broadly block T cell response. *Journal of Experimental Medicine*.

2021;218(7).

122. Gapin L. CD1a autoreactivity: When size does matter. *Journal of Experimental Medicine*. 2021;218(7).
123. Guo Z, Yang Y, Liao Y, Shi Y, Zhang LJ. Emerging Roles of Adipose Tissue in the Pathogenesis of Psoriasis and Atopic Dermatitis in Obesity. *JID Innov*. 2022;2(1):100064.
124. Hossler E, Wood G, Still C, Mowad C, Maroon M. The effect of weight loss surgery on the severity of psoriasis. *The British journal of dermatology*. 2013;168(3):660.
125. Gisondi P, Del Giglio M, Di Francesco V, Zamboni M, Girolomoni G. Weight loss improves the response of obese patients with moderate-to-severe chronic plaque psoriasis to low-dose cyclosporine therapy: a randomized, controlled, investigator-blinded clinical trial. *The American journal of clinical nutrition*. 2008;88(5):1242-7.
126. Bokarewa M, Nagaev I, Dahlberg L, Smith U, Tarkowski A. Resistin, an adipokine with potent proinflammatory properties. *The Journal of Immunology*. 2005;174(9):5789-95.
127. Shibata S, Saeki H, Tada Y, Karakawa M, Komine M, Tamaki K. Serum high molecular weight adiponectin levels are decreased in psoriasis patients. *Journal of dermatological science*. 2009;55(1):62-3.
128. Nakamizo S, Honda T, Adachi A, Nagatake T, Kunisawa J, Kitoh A, et al. High fat diet exacerbates murine psoriatic dermatitis by increasing the number of IL-17-producing $\gamma\delta$ T cells. *Scientific reports*. 2017;7(1):1-13.
129. Zhang Y, Li Q, Rao E, Sun Y, Grossmann ME, Morris RJ, et al. Epidermal fatty acid binding protein promotes skin inflammation induced by high-fat diet. *Immunity*. 2015;42(5):953-64.
130. Varga J, Marangoni RG. Dermal white adipose tissue implicated in SSc pathogenesis. *Nature Reviews Rheumatology*. 2017;13(2):71-2.
131. Varga J, Marangoni RG. Systemic sclerosis in 2016: Dermal white adipose tissue implicated in SSc pathogenesis. *Nat Rev Rheumatol*. 2017;13(2):71-2.
132. Vossen ARJV, van der Zee HH, Prens EP. Hidradenitis Suppurativa: A Systematic Review Integrating Inflammatory Pathways Into a Cohesive Pathogenic Model. *Frontiers in Immunology*. 2018;9(2965).
133. Sabat R, Jemec GBE, Matusiak L, Kimball AB, Prens E, Wolk K. Hidradenitis suppurativa. *Nature Reviews Disease Primers*. 2020;6(1):18.
134. Segura S, Requena L. Anatomy and Histology of Normal Subcutaneous Fat, Necrosis of Adipocytes, and Classification of the Panniculitides. *Dermatologic Clinics*. 2008;26(4):419-24.
135. Requena L, Sánchez Yus E. Panniculitis. Part II. Mostly lobular panniculitis. *J Am Acad Dermatol*. 2001;45(3):325-61; quiz 62-4.
136. Diaz Cascajo C, Borghi S, Weyers W. Panniculitis: definition of terms and diagnostic strategy. *Am J Dermatopathol*. 2000;22(6):530-49.
137. Blake T, Manahan M, Rodins K. Erythema nodosum - a review of an uncommon panniculitis. *Dermatol Online J*. 2014;20(4):22376.
138. Mokhtari F, Abtahi-Naeini B, Pourazizi M. Erythema nodosum migrans successfully treated with indomethacin: A rare entity. *Adv Biomed Res*. 2014;3:264.
139. Anzengruber F, Mergenthaler C, Murer C, Dummer R. Potassium Iodide for Cutaneous Inflammatory Disorders: A Monocentric, Retrospective Study. *Dermatology*. 2019;235(2):137-43.
140. Lehman CW. Control of chronic erythema nodosum with naproxen. *Cutis*. 1980;26(1):66-7.

141. Hayashi S, Ishikawa S, Ishii E, Koike M, Kaminaga T, Hamasaki Y, et al. Anti-Inflammatory Effects of Potassium Iodide on SDS-Induced Murine Skin Inflammation. *J Invest Dermatol*. 2020;140(10):2001-8.
142. Szczech J, Matławska M, Rutka M, Reich A. Clinical presentation of erythema nodosum. *Post N Med*. 2018;31(1A):25-8.
143. Pérez-Garza DM, Chavez-Alvarez S, Ocampo-Candiani J, Gomez-Flores M. Erythema Nodosum: A Practical Approach and Diagnostic Algorithm. *American Journal of Clinical Dermatology*. 2021;22(3):367-78.
144. Requena L, Sánchez Yus E. Erythema nodosum. *Semin Cutan Med Surg*. 2007;26(2):114-25.
145. Jones JV, Cumming RH, Asplin CM. Evidence for circulating immune complexes in erythema nodosum and early sarcoidosis. *Ann N Y Acad Sci*. 1976;278:212-9.
146. Maliniemi P, Hahtola S, Ovaska K, Jeskanen L, Väkevä L, Jäntti K, et al. Molecular characterization of subcutaneous panniculitis-like T-cell lymphoma reveals upregulation of immunosuppression- and autoimmunity-associated genes. *Orphanet J Rare Dis*. 2014;9:160-.
147. Kunz M, Beutel S, Bröcker E. Leucocyte activation in erythema nodosum. *Clin Exp Dermatol*. 1999;24(5):396-401.
148. De Simone C, Caldarola G, Scaldaferrì F, Petito V, Perino F, Arena V, et al. Clinical, histopathological, and immunological evaluation of a series of patients with erythema nodosum. *Int J Dermatol*. 2016;55(5):e289-94.
149. Schneider JW, Jordaan HF. The Histopathologic Spectrum of Erythema Induratum of Bazin. *The American Journal of Dermatopathology*. 1997;19(4).
150. Mascaró JM, Baselga E. Erythema Induratum of Bazin. *Dermatologic Clinics*. 2008;26(4):439-45.
151. Wu X, Subtil A, Craiglow B, Watsky K, Marks A, Ko C. The coexistence of lupus erythematosus panniculitis and subcutaneous panniculitis-like T-cell lymphoma in the same patient. *JAAD Case Rep*. 2018;4(2):179-84.
152. Burrows NP, Walport MJ, Hammond AH, Davey N, Jones RR. Lupus erythematosus profundus with partial C4 deficiency responding to thalidomide. *Br J Dermatol*. 1991;125(1):62-7.
153. Bolognia JL. *Dermatology*. 3rd ed. edited by Jean L. Bolognia ... [et al.] ed. Edinburgh: Elsevier Saunders; 2012.
154. Lee ATJ, Thway K, Huang PH, Jones RL. Clinical and Molecular Spectrum of Liposarcoma. *J Clin Oncol*. 2018;36(2):151-9.
155. Mentzel T. Cutaneous lipomatous neoplasms. *Semin Diagn Pathol*. 2001;18(4):250-7.
156. Willemze R, Cerroni L, Kempf W, Berti E, Facchetti F, Swerdlow SH, Jaffe ES. The 2018 update of the WHO-EORTC classification for primary cutaneous lymphomas. *Blood*. 2019;133(16):1703-14.
157. Parveen Z, Thompson K. Subcutaneous panniculitis-like T-cell lymphoma: redefinition of diagnostic criteria in the recent World Health Organization-European Organization for Research and Treatment of Cancer classification for cutaneous lymphomas. *Arch Pathol Lab Med*. 2009;133(2):303-8.
158. Fink-Puches R, Zenahlik P, Bäck B, Smolle J, Kerl H, Cerroni L. Primary cutaneous lymphomas: applicability of current classification schemes (European Organization for Research and Treatment of Cancer, World Health Organization) based on clinicopathologic features observed in a large group of patients. *Blood*. 2002;99(3):800-5.

159. Bauer WM, Aichelburg MC, Griss J, Skrabs C, Simonitsch-Klupp I, Schiefer AI, et al. Molecular classification of tumour cells in a patient with intravascular large B-cell lymphoma. *Br J Dermatol.* 2018;178(1):215-21.
160. Gonzalez CL, Medeiros LJ, Brazier RM, Jaffe ES. T-cell lymphoma involving subcutaneous tissue. A clinicopathologic entity commonly associated with hemophagocytic syndrome. *Am J Surg Pathol.* 1991;15(1):17-27.
161. Herbst KL. Rare adipose disorders (RADs) masquerading as obesity. *Acta Pharmacol Sin.* 2012;33(2):155-72.
162. Szolnoky G, Ifeoluwa A, Tucza M, Varga E, Varga M, Dosa-Racz E, Kemeny L. Measurement of capillary fragility: a useful tool to differentiate lipedema from obesity? *Lymphology.* 2017;50(4):203-9.
163. Al-Ghadban S, L. Herbst K, A. Bunnell B. Lipedema: A Painful Adipose Tissue Disorder. *Adipose Tissue - An Update*2019.
164. Torre YS-DI, Wadea R, Rosas V, Herbst KL. Lipedema: friend and foe. *Horm Mol Biol Clin Investig.* 2018;33(1):/j/hmbci.2018.33.issue-1/hmbci-7-0076/hmbci-2017-0076.xml.
165. Kruppa P, Georgiou I, Biermann N, Prantl L, Klein-Weigel P, Ghods M. Lipedema-Pathogenesis, Diagnosis, and Treatment Options. *Dtsch Arztebl Int.* 2020;117(22-23):396-403.
166. Forner-Cordero I, Szolnoky G, Forner-Cordero A, Kemény L. Lipedema: an overview of its clinical manifestations, diagnosis and treatment of the disproportional fatty deposition syndrome - systematic review. *Clin Obes.* 2012;2(3-4):86-95.
167. Felmerer G, Stylianaki A, Hollmén M, Ströbel P, Stepniewski A, Wang A, et al. Increased levels of VEGF-C and macrophage infiltration in lipedema patients without changes in lymphatic vascular morphology. *Scientific Reports.* 2020;10(1):10947.
168. Suga H, Araki J, Aoi N, Kato H, Higashino T, Yoshimura K. Adipose tissue remodeling in lipedema: adipocyte death and concurrent regeneration. *J Cutan Pathol.* 2009;36(12):1293-8.
169. Al-Ghadban S, Diaz ZT, Singer HJ, Mert KB, Bunnell BA. Increase in Leptin and PPAR- γ Gene Expression in Lipedema Adipocytes Differentiated in vitro from Adipose-Derived Stem Cells. *Cells.* 2020;9(2).
170. Al-Ghadban S, Pursell IA, Diaz ZT, Herbst KL, Bunnell BA. 3D Spheroids Derived from Human Lipedema ASCs Demonstrated Similar Adipogenic Differentiation Potential and ECM Remodeling to Non-Lipedema ASCs In Vitro. *International journal of molecular sciences.* 2020;21(21):8350.
171. Mann JP, Savage DB. What lipodystrophies teach us about the metabolic syndrome. *The Journal of Clinical Investigation.* 2019;129(10):4009-21.
172. Melvin A, Stears A, Savage DB. Recent developments in lipodystrophy. *Curr Opin Lipidol.* 2019;30(4):284-90.
173. Blaszczyk AM, Wright VP, Anandani K, Liu J, Jalilvand A, Bergin S, et al. Loss of Antigen Presentation in Adipose Tissue Macrophages or in Adipocytes, but Not Both, Improves Glucose Metabolism. *The Journal of Immunology.* 2019;202(8):2451.
174. Weyer C, Foley J, Bogardus C, Tataranni P, Pratley R. Enlarged subcutaneous abdominal adipocyte size, but not obesity itself, predicts type II diabetes independent of insulin resistance. *Diabetologia.* 2000;43(12):1498-506.
175. Xiao L, Yang X, Lin Y, Li S, Jiang J, Qian S, et al. Large adipocytes function as antigen-presenting cells to activate CD4⁺ T cells via upregulating MHCII in obesity. *International Journal of Obesity.* 2016;40(1):112-20.

176. Deng T, Lyon CJ, Minze LJ, Lin J, Zou J, Liu JZ, et al. Class II major histocompatibility complex plays an essential role in obesity-induced adipose inflammation. *Cell metabolism*. 2013;17(3):411-22.
177. Wong Y, Nakamizo S, Tan KJ, Kabashima K. An Update on the Role of Adipose Tissues in Psoriasis. *Frontiers in Immunology*. 2019;10(1507).
178. Lengyel E, Makowski L, DiGiovanni J, Kolonin MG. Cancer as a Matter of Fat: The Crosstalk between Adipose Tissue and Tumors. *Trends Cancer*. 2018;4(5):374-84.
179. Cozzo AJ, Fuller AM, Makowski L. Contribution of Adipose Tissue to Development of Cancer. *Compr Physiol*. 2017;8(1):237-82.
180. López-Lerma I, Peñate Y, Gallardo F, Martí RM, Mitxelena J, Bielsa I, et al. Subcutaneous panniculitis-like T-cell lymphoma: Clinical features, therapeutic approach, and outcome in a case series of 16 patients. *Journal of the American Academy of Dermatology*. 2018;79(5):892-8.
181. Akdis CA. Does the epithelial barrier hypothesis explain the increase in allergy, autoimmunity and other chronic conditions? *Nat Rev Immunol*. 2021;21(11):739-51.
182. Ter Poorten MC, Thiers BH. Panniculitis. *Dermatol Clin*. 2002;20(3):421-33, vi.
183. Johnson EF, Tolkachjov SN, Gibson LE. Alpha-1 antitrypsin deficiency panniculitis: clinical and pathologic characteristics of 10 cases. *Int J Dermatol*. 2018;57(8):952-8.
184. Morgan AJ, Schwartz RA. Cutaneous polyarteritis nodosa: a comprehensive review. *Int J Dermatol*. 2010;49(7):750-6.
185. Bhat RM, Vaidya TP. What is New in the Pathogenesis and Management of Erythema Nodosum Leprosum. *Indian Dermatol Online J*. 2020;11(4):482-92.
186. Crowson AN, Mihm MC, Jr., Magro CM. Cutaneous vasculitis: a review. *J Cutan Pathol*. 2003;30(3):161-73.
187. Lee JT, Kalani MA. Treating superficial venous thrombophlebitis. *J Natl Compr Canc Netw*. 2008;6(8):760-5.
188. LeBlanc RE, Tavallae M, Kim YH, Kim J. Useful Parameters for Distinguishing Subcutaneous Panniculitis-like T-Cell Lymphoma From Lupus Erythematosus Panniculitis. *Am J Surg Pathol*. 2016;40(6):745-54.
189. Requena C, Sanmartín O, Requena L. Sclerosing panniculitis. *Dermatol Clin*. 2008;26(4):501-4, vii.
190. Greenberg AS, Hasan A, Montalvo BM, Falabella A, Falanga V. Acute lipodermatosclerosis is associated with venous insufficiency. *J Am Acad Dermatol*. 1996;35(4):566-8.
191. Polcari IC, Stein SL. Panniculitis in childhood. *Dermatol Ther*. 2010;23(4):356-67.
192. Burden AD, Krafchik BR. Subcutaneous fat necrosis of the newborn: a review of 11 cases. *Pediatr Dermatol*. 1999;16(5):384-7.
193. Dahl PR, Su WP, Cullimore KC, Dicken CH. Pancreatic panniculitis. *J Am Acad Dermatol*. 1995;33(3):413-7.
194. Delgado-Jimenez Y, Fraga J, García-Díez A. Infective panniculitis. *Dermatol Clin*. 2008;26(4):471-80, vi.
195. Moreno A, Marcoval J, Peyri J. Traumatic panniculitis. *Dermatol Clin*. 2008;26(4):481-3, vii.
196. Quesada-Cortés A, Campos-Muñoz L, Díaz-Díaz RM, Casado-Jiménez M. Cold panniculitis. *Dermatol Clin*. 2008;26(4):485-9, vii.

197. Pielasinski Ú, Machan S, Camacho D, Juárez Á, Cedeño M, Ruiz Maciá JA, Requena L. Postirradiation Pseudosclerodermatous Panniculitis: Three New Cases With Additional Histopathologic Features Supporting the Radiotherapy Etiology. *The American Journal of Dermatopathology*. 2013;35(1):129-34.
198. Yanes AF, Owen JL, Colavincenzo ML. Factitial panniculitis as a manifestation of self-imposed factitious disorder. *Dermatol Online J*. 2019;25(5).
199. Marcoval J, Moreno A, Mañá J, Peyri J. Subcutaneous sarcoidosis. *Dermatol Clin*. 2008;26(4):553-6, ix.
200. Kwon EJ, Emanuel PO, Gribetz CH, Mudgil AV, Phelps RG. Poststeroid panniculitis. *J Cutan Pathol*. 2007;34 Suppl 1:64-7.
201. López-Lerma I, Peñate Y, Gallardo F, Martí RM, Mitxelena J, Bielsa I, et al. Subcutaneous panniculitis-like T-cell lymphoma: Clinical features, therapeutic approach, and outcome in a case series of 16 patients. *Journal of the American Academy of Dermatology*. 2018;79(5):892-8.
202. Mascaró JM, Jr., Baselga E. Erythema induratum of bazin. *Dermatol Clin*. 2008;26(4):439-45, v.
203. Tran TA, DuPree M, Carlson JA. Neutrophilic lobular (pustular) panniculitis associated with rheumatoid arthritis: a case report and review of the literature. *Am J Dermatopathol*. 1999;21(3):247-52.
204. Sutra-Loubet C, Carlotti A, Guillemette J, Wallach D. Neutrophilic panniculitis. *Journal of the American Academy of Dermatology*. 2004;50(2):280-5.