

# Body mass index and adipokines/cytokines dysregulation in Systemic Sclerosis

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## Abstract

Body fat has regulatory functions through producing cytokines and adipokines whose role in the pathogenesis of Systemic Sclerosis (SSc) is currently emerging. Changes in body mass, either overweight or underweight status, entail a dysregulation of the cytokines/adipokines network that may impact on SSc disease activity. We evaluated serum levels of adipokines and cytokines in SSc patients and correlated them to clinical features and body mass index (BMI) categories. The study included 89 SSc patients and 26 healthy donors (HD). Serum levels of adiponectin, leptin, resistin, visfatin, TNF $\alpha$ , IFN $\gamma$ , IL-2, IL-10, and IL-17A were measured by Multiplex Immunoassay, and correlated to BMI, waist to hip ratio, and disease specific features. Mann-Whitney U-test or t-Student for unpaired data, Kruskal-Wallis test or ANOVA, were used for comparisons between groups. Spearman's or Pearson's test were used for correlation analysis. Serum levels of TNF $\alpha$ , IL-2, leptin, and resistin, were significantly higher in SSc than in HD. The highest levels of IL-17A, IL-2, IL-10, leptin and visfatin were detected in obese SSc patients ( $p < 0.01$ ). Conversely, underweight SSc patients showed the highest TNF $\alpha$  levels ( $p < 0.05$ ), which were negatively correlated with BMI ( $p = 0.05$ ). No correlation between adipokines/cytokines and clinical characteristics was found. Adipokines, IL-2, IL-10 and IL-17A were found to be increased in obese SSc patients, but whether they play a role in the pathogenesis of the disease remains to be investigated. Intriguingly, underweight patients had higher TNF $\alpha$  levels, suggesting a potential role of TNF $\alpha$  in inducing the cachexia observed in long-lasting disease.

**Title: Body mass index and adipokines/cytokines dysregulation in Systemic Sclerosis**

**Running Title: Systemic sclerosis, adipokines and BMI**

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## ABSTRACT

Body fat has regulatory functions through producing cytokines and adipokines whose role in the pathogenesis of Systemic Sclerosis (SSc) is currently emerging. Changes in body mass, either overweight or underweight status, entail a dysregulation of the cytokines/adipokines network that may impact on SSc disease activity. We evaluated serum levels of adipokines and cytokines in SSc patients and correlated them to clinical features and body mass index (BMI) categories.

The study included 89 SSc patients and 26 healthy donors (HD). Serum levels of adiponectin, leptin, resistin, visfatin, TNF $\alpha$ , IFN $\gamma$ , IL-2, IL-10, and IL-17A were measured by Multiplex Immunoassay, and correlated to BMI, waist to hip ratio, and disease specific features. Mann-Whitney U-test or t-Student for unpaired data, Kruskal-Wallis test or ANOVA, were used for comparisons between groups. Spearman's or Pearson's test were used for correlation analysis.

Serum levels of TNF $\alpha$ , IL-2, leptin, and resistin, were significantly higher in SSc than in HD. The highest levels of IL-17A, IL-2, IL-10, leptin and visfatin were detected in obese SSc patients ( $p < 0.01$ ). Conversely, underweight SSc patients showed the highest TNF $\alpha$  levels ( $p < 0.05$ ), which were negatively correlated with BMI ( $p = 0.05$ ). No correlation between adipokines/cytokines and clinical characteristics was found.

Adipokines, IL-2, IL-10 and IL-17A were found to be increased in obese SSc patients, but whether they play a role in the pathogenesis of the disease remains to be investigated. Intriguingly, underweight patients had higher TNF $\alpha$  levels, suggesting a potential role of TNF $\alpha$  in inducing the cachexia observed in long-lasting disease.

## Introduction

Systemic sclerosis (SSc) is a connective tissue disease characterized by small vessels vasculopathy associated with fibrosis of multiple organs. The pathogenesis is unclear, but an autoimmune dysregulation and an

abnormal inflammatory response seem to be involved in the early stage of the disease. Emerging evidence suggests that white adipose tissue (WAT), besides having the role of energy storage, is now acknowledged as a pleiotropic organ with endocrine functions and regulating immune and inflammatory responses, being a source of cytokines and adipokines (leptin, adiponectin, visfatin, resistin) (1). Indeed, obesity may have an impact on disease activity as well as on clinical response of patients with rheumatoid arthritis and psoriatic arthritis (2,3). In obese subjects, leptin induces the expression of adhesion molecules on endothelial cells and activates macrophages, while hindering adiponectin production by adipocytes, contributing to the “low-grade inflammatory state” associated with obesity (1).

Recently, the role of cytokines/adipokines in the pathophysiology of SSc has become a matter of investigation but studies correlating them with the clinical subsets or particular organ involvement were not always consistent. Serum leptin levels were found to be positively correlated with body mass index (BMI), negatively with disease activity in SSc patients, but not increased in comparison with healthy controls (4,5). Conversely, serum levels of leptin, resistin and TNF $\alpha$  were higher in a small cohort of 16 SSc patients than in control subjects but no correlation with skin involvement, disease duration and disease activity was seen (6). A newly discovered adipokine, adipisin, was significantly higher in limited cutaneous SSc (lcSSc) than in diffuse cutaneous SSc (dcSSc) and was strongly associated with pulmonary arterial hypertension (7). Interestingly, adiponectin seems to play a protective role in SSc, as the levels were found to be low in dcSSc patients and inversely correlated with the extension of skin fibrosis, C-reactive protein (CRP) and erythrocyte sedimentation rate (ESR) (5,8-10). However, the interweave among body mass, cytokines/adipokines and SSc clinical phenotypes has been poorly investigated.

In this study, we aimed at evaluating serum levels of adipokines (leptin, resistin, visfatin, adiponectin) and cytokines (TNF $\alpha$ , Interferon-gamma (IFN $\gamma$ ), IL-2, IL-10, IL-17A) in SSc patients according to BMI categories and disease specific characteristics.

## Materials & Methods

### Ethical approval

The study obtained the approval of the local ethics committee (Azienda Policlinico Bari, n.5351 14/09/2017). The study was conducted in compliance with the Helsinki’s declaration, and all patients gave their written informed consent to participate and for use of their data for publication, with explicit protection of their identity

### Patient group

We evaluated 89 Caucasian patients with Systemic Sclerosis (SSc) fulfilling the 2013 ACR/ EULAR classification criteria (11) and 26 healthy blood donors (HD) as control group, including subjects not suffering from any disease and not taking medications at enrolment.

### Cytokine assay

Blood samples obtained from all participants were centrifuged and serum was separated and stored in aliquots at  $-80^{\circ}\text{C}$  until use. Serum levels of adipokines (adiponectin, leptin, resistin, visfatin) and cytokines (TNF $\alpha$ , IFN $\gamma$ , IL-2, IL-10, IL-17A) were measured, in duplicate, using kits manufactured by Bio-Rad (Bioplex ProTM Cytokine / Chemokine and Growth Factor Assay and Bioplex Pro Diabetes Assay) for Multiplex Immunoassay (Bioplex 200 System by Bio-Rad Laboratories, Hercules (CA, USA)). The analyses were carried out according to the manufacturer’s instructions.

### Clinical data

In all patients we collected clinical, anthropometric and laboratory features, including disease duration, onset of Raynaud’s phenomenon, skin involvement according to the modified Rodnan Skin Score (mRSS) (12) and cutaneous subset according to LeRoy criteria (13). Moreover, the presence/absence of interstitial lung disease (ILD) defined by chest HR-CT scan, pulmonary function test (PFT) with FVC, FEV1/FVC, DLCO and RV estimation, the 6-minute walking distance test (6mWDT), the presence/absence of pulmonary

arterial hypertension (PAH) diagnosed by right heart catheterization, the presence/absence of esophagopathy evaluated at esophageal scintigraphy or esophageal manometry or chest HR-CT scan, the presence of digital pitting-scars and/or digital ulcers (past or active), the nailfold capillaroscopic pattern "Early", "Active" and "Late" (14), the disease activity according to the ESSG (European Scleroderma Study Group) disease activity index (15), were recorded. Drugs investigated were: peripheral vasodilators, iloprost, immunosuppressive drugs, glucocorticoids, antihypertensive and lipid-lowering agents, targeted therapy for the treatment of PAH (bosentan, sildenafil, tadalafil, ambrisentan), oxygen therapy. The BMI, waist circumference (Waist C) and Waist-to-Hip Ratio (WHR), cardiovascular risk index, acute phase proteins, cholesterol and triglycerides serum levels, previous smoking status, comorbidity (such as arterial hypertension, diabetes mellitus and hyperlipidemia) were also assessed. The BMI was calculated as weight in kilograms divided by the height in square meters and in accordance with the WHO BMI category classification, patients were considered underweight (BMI <18.5), normal-weight (BMI= 18.5-24.99), overweight (BMI= 25-29.99) and obese ([?]30). The cardiovascular (CV) risk was estimated as the ratio of total cholesterol/high-density-lipoprotein (HDL), or applying the Framingham Risk Score (FRS).

### Statistical Analysis

Data were analyzed using IBM-SPSS Statistics 20 software. The comparison between two groups were done by Mann-Whitney U-test or t-Student, or chi-square test where appropriate. Comparisons among several groups were made using Kruskal-Wallis test or ANOVA, as appropriate. Spearman's or Pearson's test were used for correlations between cytokine/adipokines and clinical characteristics. Multiple linear regressions were performed for the analysis of predictors. A *p-value* [?] 0.05 was considered statistically significant.

### Results

#### Patient characteristics

Clinical characteristics of SSc patients are shown in Table 1. Most patients had limited cutaneous SSc (83%), and 27% had active disease ([?] 3 ESSG score). Of note, only 5 (6%) patients were obese (BMI [?] 30), while 9 (10%) were underweight (BMI [?] 18.5). At study entry, drugs taken were: calcium channel blockers (CCB) (67.4%), iloprost (48.3%) glucocorticoids (34.8%), endothelin receptor antagonists (ERA) (22.3%), methotrexate (19.5%), mycophenolate mofetil (13.5%), phosphodiesterase-5 inhibitors (PDE5i) (tadalafil, sildefanil) (8.9%). No patient was taking non-steroidal anti-inflammatory drugs. In Table 2, the comparison of demographics, anthropometric characteristics, and serum levels of cytokines and adipokines between SSc and HD are shown. No significant differences in anthropometric characteristics were detected, but a trend toward a lower BMI was seen in SSc patients.

#### Adipokines and Cytokines

Table 2 shows the comparison of cytokines and adipokines between HD and the whole SSc cohort. Leptin and resistin were significantly higher in SSc than in HD. Visfatin was also increased in SSc, but the difference from HD did not reach the statistical significance. Adiponectin was measured only in the SSc group (see below). Remarkably, SSc patients showed significantly higher levels of TNF $\alpha$  and IL-2 than HD. Because of a possible bias by drugs, serum levels of cytokines and adipokines were correlated to the different treatments by multiple linear regression models. To this end, patients were grouped into SSc-glucocorticoids, SSc-ERA, SSc-immunosuppressants, SSc-CCB, SSc-iloprost, SSc-PDE5i. Only PDE5 inhibitors were independently associated with higher levels of leptin (OR 3.27, 95% CI 12892-52716,  $p=0.002$ ) and visfatin (OR 2.66, 95% CI 4340-29950,  $p=0.009$ ). Therefore, SSc patients were stratified into those taking PDE5i and those who did not, and compared by cytokines and adipokines levels (Table 2). Although only 8 SSc patients were on treatment with PDE5i, their levels of leptin and visfatin were significantly higher than in SSc patients without PDE5i (Table 2), while no difference was detected of the remaining cytokines.

No statistically significant difference in cytokines/adipokines levels between early SSc (<2 years of disease duration) and late disease (>2 years of disease duration) was observed. We also stratified SSc patients as high and low CV risk according to the Framingham score and other indices (cholesterol /HDL ratio >4.5, waist

circumference >88 cm, WHR >0.8). As only 3 patients had high/medium CV FRS, statistical comparison with those showing low risk CV FRS could not be made (data not shown). Furthermore, no significant differences were found when patients were stratified according to DLCO <75% or FVC <70% values, or the disease activity index ESSG < 3. Likewise, no peculiar pattern of cytokines/adipokines was found in lcSSc or dcSSc subsets.

### BMI categories

When SSc patients were subdivided by BMI categories, some meaningful differences emerged (Table 3). As expected, leptin (Figure 1) and visfatin levels were significantly higher in obese than in other BMI classes ( $p=0.0001$  and  $p=0.002$ , respectively), while no significant changes were observed for resistin. Obese SSc patients also had the highest serum levels of IL-17A ( $p=0.01$ ), IL-2 ( $p=0.001$ ) and IL-10 ( $p=0.01$ ). Adiponectin levels did not significantly change among the BMI subgroups, but the leptin/adiponectin ratio, a functional biomarker of adipose tissue inflammation, was significantly higher in obese patients ( $p=0.0001$ ). A striking finding was the increased TNF $\alpha$  levels observed in underweight SSc patients. The latter had significantly higher TNF $\alpha$  levels ( $p=0.01$ ) than the other BMI classes (Table 3, Figure 2). IL-17A, leptin, and visfatin levels were found to be positively correlated, and TNF $\alpha$  negatively, with increasing BMI values (table 4). BMI was also correlated with ESR, CRP, triglycerides and cholesterol levels (Table 4).

### Discussion

In this study, we investigated the serum levels of different adipokines (adiponectin, leptin, resistin, visfatin) and cytokines (TNF $\alpha$ , INF, IL-12, IL-10, IL-17A) in SSc patients and searched for possible correlations with BMI and specific clinical manifestations of the disease. TNF $\alpha$ , IL-2, leptin and resistin were higher in SSc patients than in HD. These findings are globally consistent with the literature reporting an increase in cytokines/adipokines in SSc to different extents (4,8,9,16-24). However, attempts to correlate each cytokine/adipokine to the disease activity of SSc and to BMI have yielded conflicting results. All the studies but one (25) showed increased serum levels of leptin in SSc, sometimes correlating with BMI. We found significantly higher leptin levels in SSc than HD and a positive correlation of leptin with BMI, but no correlations with PHA or other clinical manifestations were detected. Instead, a previous study had demonstrated that leptin serum levels were higher in idiopathic PHA and SSc-PAH patients than controls and that dysfunctional endothelial cells from SSc-PAH lung produced leptin “*in vitro*”, although a link with BMI was not investigated (26). Furthermore, we detected significantly higher resistin in SSc patients than in HD, but it was not regulated by BMI, as already reported (6). On the contrary, we found that visfatin levels rose with BMI increases in SSc patients but they were still statistically comparable to HD. Masui et al (22) had detected similar levels of visfatin in SSc patients and controls, but noticed higher visfatin levels in dcSSc patients with late disease, without exploring BMI status. This general inconsistency may be also influenced by treatments, as we found that SSc patients taking PDE5i (tadalafil or sildenafil) but not bosentan, had significantly higher leptin and visfatin levels than patients without PDE5i. Furthermore, SSc-PDE5i patients had 2-3 folds the odds to have high leptin and visfatin levels. It is conceivable that this might be a specific PDE5i effect, rather than related to PAH, as adipokines secretion by white adipocytes is regulated by cAMP and increases upon PDE inhibition “*in vitro*” (27).

Adiponectin can generally be accounted as leptin antagonist with anti-inflammatory properties and decreases in obesity (28). Adiponectin has also been suggested to have also anti-fibrotic activities and seems to be regulated in SSc, depending on the skin fibrosis extension and disease duration. Some studies demonstrated that adiponectin is low in dcSSc patients both in serum and in lesional skin, but increases in dcSSc patients with a disease duration longer than 5 years, when skin thickness reduces (9,20,21,29). We found low, although not statistically significantly low levels of adiponectin in obese SSc, but we could not confirm previous data as we studied only 15 patients with dcSSc. Interestingly, the ratio leptin/adiponectin was 10-fold higher in obese SSc patients suggesting that the reciprocal leptin/adiponectin regulation is functionally unbalanced in SSc. At this point, a critical question to be addressed is “why is leptin increased in SSc patients as their BMI was lower than in HD”? Indeed, no study has ever demonstrated an increased frequency of obesity among SSc patients. In our cohort, only 6% had a BMI >30 compared to 12.3% of the general population

in Apulia (*ISTAT, report Osservasalute 2016* ) implying that leptin overexpression in SSc might be due to some adipocyte dysfunction rather than to an increase production by visceral fat.

Among the investigated cytokines, we found significantly higher levels of TNF $\alpha$  and IL-2 in SSc as compared to HD, presumably linked to the biologic activity of the disease, despite no correlation with the clinical manifestations nor with the global disease activity was found. Within the SSc cohort, obese patients had significantly higher levels of IL-17A and IL-10. A correlation between IL-17A and obesity was expected as high IL-17 mRNA expression has been found in visceral fat of morbidly obese women (30). On the other hand, the significantly higher IL-10 levels in SSc obese patients were unexpected, as in obese subjects IL-10 tends to be low and increases with exercise and weight loss (31). Maybe the most intriguing finding in our analysis was the strikingly high levels of TNF $\alpha$  in underweight SSc patients, roughly 10-folds higher than in normal-weight patients. Increased levels of TNF $\alpha$  in SSc had already been reported (6,18,19,32) although a link with a particular phenotype was not shown. Only one study had demonstrated a correlation of TNF $\alpha$  levels with lung fibrosis and impairment of pulmonary vital capacity (32). Of note, TNF $\alpha$  blocking agents have been successfully used in SSc patients with arthritis (33) and further investigations should focus on this possible pathogenic association. In our study, 10% of SSc patients were underweight, and loss of body mass has been associated mainly with malabsorption (34). Besides, an overexpression of TNF may also be considered as a further mechanism involved in the cachexia-like status of SSc. During the 1980s, that TNF $\alpha$  and cachectin were demonstrated to be the two faces of the same coin (35). In an experimental model, TNF $\alpha$  induced weight loss directly proportional to the decreased food and water intake (36). Moreover, it is known that anti-TNF $\alpha$  drugs may increase body weight and it has been reported that etanercept treatment promoted weight gain and reduced cachexia in patients with rheumatoid arthritis (37).

In conclusion, despite some limitations, such as the cross-sectional design, drug interference, mainly PDE5i, the relatively small sample size of our SSc cohort, this study suggests that an abnormal twist between cytokines, adipokines and BMI takes place in SSc, and these changes in adipokines maybe related to a disfunction of adipocytes (or of other different sources) rather than to the BMI. Further investigation is warranted to establish whether these findings may represent the pathogenetic background of specific clinical manifestations of SSc.

### Authors' contributions

FI and EP conceived the study, were the major participants in its design, coordination, interpretation of results and statistical analysis, they also prepared draft manuscript. DN, RB and NL carried out biological assays, RC, MF and FC collected clinical data and participated in study design coordination. All authors were involved in draft manuscript modifications and approved the final version of the manuscript.

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Table 1. Clinical findings of Systemic Sclerosis (SSc) cohort (n. 89 patients).

Disease duration (years), mean±SD	8.1 ±6.6
Underweight (BMI [?]18.5), n. (%)	9 (10 %)
Normal weight (BMI >8.5 <25), n. (%)	49 (55 %)
Overweight (BMI [?]25 [?]30), n. (%)	26 (29 %)
Obese (BMI >30), n. (%)	5 (6 %)
Diabetes Mellitus, n. (%)	1 (9 %)
Dyslipidemia, n. (%)	11 (12 %)
Tot-Cholesterol/HDL, mean ±SD	3.3 ±1.0
Framingham risk score, mean ±SD	3.7 ±2.9
Previous smokers, n (%)	5 (6 %)
Arterial Hypertension, n (%)	8 (7 %)
Active patients with disease activity index [?]3 ESSG, n. (%)	24 (27 %)
Erythrocyte Sedimentation Rate (mm/h), mean±SD	19 ± 14
C-Reactive Protein (mg/l), mean±SD	4.3 ± 5
Limited cutaneous SSc, n. (%)	74 (83 %)
Interstitial lung disease, n. (%)	51 (57 %)
Esophageal Involvement, n. (%)	79 (89 %)
Bowel incontinence, n. (%)	2 (3 %)
Active digital ulcers, n. (%)	18 (20 %)
Digital pitting scars, n. (%)	55 (62 %)
Calcinosis, n. (%)	79 (89 %)
Renal Involvement, n. (%)	5 (6 %)
Pulmonary Arterial Hypertension, n. (%)	7 (8 %)
Capillaroscopy scleroderma pattern, n. (%)	Early 6 (7 %) Active 49 (55 %) Late 34 (38 %)

Table 2. Demographics and cytokines/adipokines serum levels in Systemic Sclerosis (SSc) patients as whole cohort and

subdivided according to phosphodiesterase-5 (PDE5) inhibitors (tadalafil/sildenafil) intake, and healthy donors (HD).

Data are shown as mean ± standard deviation.

	Comparison SSc-all vs healthy donors (HD)	Comparison SSc-all vs healthy donors (HD)	Comparison SSc-all vs healthy donors (HD)	Comparison SSc tadalafil/sildenafil vs SSc other drugs (OD)	Comparison SSc tadalafil/sildenafil vs SSc other drugs (OD)	Comparison SSc tadalafil/sildenafil vs SSc other drugs (OD)
	SSc all (n. 89)	HD (n. 26)	p	SSc PDE5 inhibitors (n. 8)	SSc OD (81)	p
Age (years)	52.1 ± 14	49.4 ± 11	0.07	64.5 ± 14	50.3 ± 13	0.0001
Female n (%)	84 (94 %)	23 (88%)	0.37	7 (87%)	76 (93%)	0.37
Waist cir- cumference (cm)	80.7 ± 9.5	88.4 ± 12	0.49	90 ± 5.6	79.6 ± 9.4	0.06

	Comparison SSc-all vs healthy donors (HD)	Comparison SSc-all vs healthy donors (HD)	Comparison SSc-all vs healthy donors (HD)	Comparison SSc tadalafil/sildenafil vs SSc other drugs (OD)	Comparison SSc tadalafil/sildenafil vs SSc other drugs (OD)	Comparison SSc tadalafil/sildenafil vs SSc other drugs (OD)
Hip circum- ference (cm)	97.5 ± 10	102.5 ± 9.9	0.15	110 ± 4.2	95.9 ± 10	0.06
WHR (Waist/Hip ratio)	0.83 ± 0.7	0.86 ± 0.08	0.26	0.81 ± 0.01	0.83 ± 0.07	0.53
Body Mass Index (Kg/m <sup>2</sup> )	23.6 ± 4.2	25.13 ± 4.4	0.12	27.1 ± 5.5	23.2 ± 3.9	0.01
IL-17A	2.4 ± 6	1.7 ± 3.1	0.96	1.3 ± 2	1.8 ± 7	0.94
TNF-α	57.2 ± 62	1.0 ± 1.1	0.02	31.4 ± 60	60.4 ± 280	0.50
INF-γ	88.9 ± 172	29.6 ± 23	0.09	110 ± 175	166.1 ± 492	0.25
IL-2	42 ± 162	3.57 ± 1.3	0.01	104.5 ± 285	38 ± 149.6	0.17
IL-10	40 ± 175	4.3 ± 9.8	0.29	127.6 ± 342	33.4 ± 153.2	0.12
Leptin	15266 ± 26679	2822 ± 2045	0.00003	28353 ± 33778	11768 ± 11488	0.0001
Resistin	6146 ± 2344	1705 ± 279	0.00001	6019 ± 3211	6322 ± 2147	0.85
Visfatin	4459 ± 15177	2555 ± 12083	0.55	17732 ± 46746	3245 ± 6650	0.01
Adiponectin		/		5848562 ± 6204076	6833220 ± 5694341	0.64

Table 3. Cytokines and adipokines in Systemic Sclerosis patients by BMI categories (mean ± SD).

	Underweight	Normal-weight	Overweight	Obese	<i>p</i>
IL-17A	0.05 ± 0.1	2.24 ± 9.1	0.8 ± 1.5	6.9 ± 6.7 <i>vs:</i> <i>UW, NW OW</i>	0.01
TNFα	144.2 ± 271 <i>vs:</i> <i>NW OW, Ob</i>	12.1 ± 43	31.1 ± 121	42 ± 71	0.03
INFγ	76.4 ± 125	83.7 ± 251	105.0 ± 303	91.5 ± 249	0.97
IL-2	23.4 ± 85	11.4 ± 45	35.9 ± 129	233 ± 350 <i>vs:</i> <i>UW, NW OW</i>	0.001
IL-10	12.0 ± 31	12.8 ± 35	30.1 ± 112	204 ± 431 <i>vs:</i> <i>UW, NW OW</i>	0.01
Leptin	9033 ± 7181	9623 ± 14097	17465 ± 10335	64970 ± 89346 <i>vs: UW, NW OW</i>	0.0001
Visfatin	127 ± 234	2154 ± 4199	4340 ± 7900	28551 ± 58672 <i>vs: UW, NW OW</i>	0.002
Resistin	6997 ± 956	6109 ± 2519	6637 ± 1727	6964 ± 1800	0.57
Adiponectin	9829075 ± 66275	6582923 ± 53445	7459125 ± 65585	3248520 ± 10425	0.22

	Underweight	Normal-weight	Overweight	Obese	<i>p</i>
Leptin/Adiponec	0.002 ± 0.002	0.002 ± 0.004	0.003 ± 0.0006	0.02 ± 0.04 <i>vs:</i> <i>UW, NW OW</i>	0.0001

UW= Underweight, NW= Normal-weight, OW= Overweight, Ob= Obese.

Table 4. Correlation of cytokines/adipokines with anthropometric and clinical findings. Data are shown as mean ± SD. Only the significant correlations are reported.

	IL-17A	Leptin	Visfatin	TNF $\alpha$	Trigl	Chol LDL	ESR	CRP
BMI	<i>r</i> 0.25 <i>p</i> 0.02	<i>r</i> 0.41 <i>p</i> 0.0001	<i>r</i> 0.25 <i>p</i> 0.02	<i>r</i> -0.27 <i>p</i> 0.05	<i>r</i> 0.28 <i>p</i> 0.02	<i>r</i> 0.46 <i>p</i> 0.002	<i>r</i> 0.30 <i>p</i> 0.007	<i>r</i> 0.32 <i>p</i> 0.004

BMI= Body, Mass, Index, Trigl= Triglycerides, Chol= Cholesterol, ESR= Erythrocyte Sedimentation Rate, CRP= C-reactive protein, *r*= coefficient correlation, *p*= significance level

### Legends

**Figure 1** . Serum leptin levels in healthy donors (HD) and Systemic Sclerosis (SSc) patients with different categories of body mass index. (U-W: under-weight, N-W normal-weight, O-W over-weight). Mean (95% CI)

**Figure 2** . Serum TNF $\alpha$  levels in healthy donors (HD) and Systemic Sclerosis (SSc) patients with different categories of body mass index. (U-W: under-weight, N-W normal-weight, O-W over-weight). Mean (95% CI)



