

# Screening potential insect vectors in a museum biorepository reveals undiscovered diversity of plant pathogens in natural areas

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## Abstract

Phytoplasmas (*Mollicutes*, *Acholeplasmataceae*), vector-borne obligate bacterial plant-parasites, infect nearly 1,000 plant species and unknown numbers of insects, mainly leafhoppers (Hemiptera, *Deltocephalinae*), which play a key role in transmission and epidemiology. Although the plant-phytoplasma-insect association has been evolving for >300 million years, nearly all known phytoplasmas have been discovered as a result of the damage inflicted by phytoplasma diseases on crops. Few efforts have been made to study phytoplasmas occurring in non-economically important plants in natural habitats. In this study, a sub-sample of leafhopper specimens preserved in a large museum biorepository was analyzed to unveil potential new associations. PCR screening for phytoplasmas performed on 227 phloem-feeding leafhoppers collected worldwide from natural habitats revealed the presence of 6 different previously unknown phytoplasma strains. This indicates that museum collections of herbivorous insects represent a rich and largely untapped resource for discovery of new plant pathogens, that natural areas worldwide harbor a diverse but largely undiscovered diversity of phytoplasmas and potential insect vectors, and that independent epidemiological cycles occur in such habitats, posing a potential threat of disease spillover into agricultural systems. Larger-scale future investigations will contribute to a better understanding of phytoplasma genetic diversity, insect host range, and insect-borne phytoplasma transmission and provide an early warning for the emergence of new phytoplasma diseases across global agroecosystems.

## Introduction

Phytoplasmas (*Mollicutes*, *Acholeplasmatales*, *Acholeplasmataceae*) are a large group of phloem-restricted, cell wall-less, vector-borne bacteria that infect hundreds of plant species and cause serious economic loss worldwide (Rao et al., 2018). In plants, phytoplasma infection may induce a variety of typical symptoms including virescence, phyllody, and witches'-broom, thereby altering plant morphology, growth patterns and architecture (MacLean et al., 2011; 2014; Wei et al., 2013; 2019), although infections may also be asymptomatic (Zwolinska et al., 2019).

Phytoplasmas are transmitted from plant to plant by phloem-feeding hemipteran insect vectors, mainly leafhoppers, in a persistent-propagative manner (Hogenhout et al., 2008; Lee et al., 2000; Weintraub & Beanland, 2006). After acquisition of phytoplasmas from an infected plant by a hemipteran insect, the phytoplasma cells must cross the midgut epithelium, then multiply in the hemolymph in order to invade the salivary glands before being inoculated into another host plant (Hogenhout et al., 2008; Huang et al., 2020).

Attempts to culture phytoplasmas *in vitro* have, thus far, not succeeded. Thus, phytoplasmas are currently assigned to the provisional genus '*Candidatus* (*Ca.*) *Phytoplasma*', and 45 '*Ca.* *Phytoplasma*' species have been described (IRPCM, 2004; Kirdat et al., 2020; Naderali et al., 2017; Rodrigues Jardim et al., 2020; Šafářová et al., 2016; Zhao et al., 2021). Nevertheless, the phytoplasma lineage is a highly diverse

monophyletic group (Gupta et al., 2018; Zhao et al., 2015), having been classified into 36 groups, and more than 150 subgroups based on distinct 16S rRNA gene restriction fragment length polymorphism patterns (Lee et al., 1998; Naderali et al., 2017; Rodrigues Jardim et al., 2020; Seemüller et al., 1998; Wei et al., 2007; Zhao et al., 2009).

The intimate tri-trophic interaction among phytoplasmas, host plants, and insect vectors defines a complex of multiple pathosystems worldwide (Trivellone, 2019). Unfortunately, almost all phytoplasma-host associations have been characterized by testing plants showing symptoms of diseases in agroecosystems. However, because the association between phytoplasmas, plants and insect vectors has been evolving for at least 300 million years (Cao et al., 2020), phytoplasmas and their vectors should also be widespread and diverse in non-managed, native habitats (Trivellone & Dietrich, 2020). Indeed, current theories of infectious disease evolution suggest that most epidemic diseases afflicting humans, livestock and crops emerge as a result of potentially pathogenic organisms “jumping” from a native host to a new host following anthropogenic disturbance of natural habitats (Brooks et al., 2019).

About 100 insect species have been recorded as competent vectors of phytoplasmas; however, for most the of described ‘*Ca.Phytoplasma*’ species and 16S rRNA subgroups the suite of vectors is still unknown (overview in Trivellone, 2019). Because insects are often difficult to identify and individuals infected with phytoplasmas cannot be distinguished from non-infected individuals except through microscopy, molecular screening, or pathogen transmission trials, efforts to identify competent phytoplasma vectors have lagged far behind efforts to characterize phytoplasmas and their host plants. Due to the mobility of insect vectors, spillovers of vector-borne phytoplasmas from adjacent highly diverse natural habitats into agroecosystems were hypothesized to play an important role in emergence of new phytoplasma diseases (see Brooks et al., accepted). However, few attempts have been made to study phytoplasma diversity in natural habitats. Therefore, diversity, plant host range, and insect vector range of phytoplasmas are probably significantly underestimated (Trivellone & Dietrich, 2020).

Due to increased awareness of the importance of wildlife as pathogen reservoirs (Brooks et al., 2020), the use of museum biorepositories to discover and track pathogens is a critical step for anticipating the emergence and re-emergence of zoonotic diseases (DiEuliis et al., 2016; Dunnun et al., 2017). The high levels of biodiversity and geographic coverage represented in such repositories can also help unveil the evolutionary history of pathogens and reveal previously unknown interactions with actual or potential hosts.

In this study, we analyzed specimens of deltocephaline leafhoppers (Hemiptera: Cicadellidae: Deltocephalinae) preserved in the collection of the Illinois Natural History Survey (INHS) (<http://inhsinsectcollection.speciesfile.org/InsectCollection.aspx>). The INHS leafhopper collection is one of the largest in world with over >400,000 specimens stored either pinned or in ethanol at -20°C. In 2018, a subsample of ethanol-preserved leafhoppers collected in natural habitats were tested for presence of phytoplasmas. The results revealed that about 3% of tested insect specimens harbored phytoplasmas. The newly discovered phytoplasmas belong to three distinct taxonomic (16Sr) groups. Phytoplasmas were detected from a total of six leafhopper species including five known and one recently described species, all recorded for the first time as potential phytoplasma vectors. These results indicated that phytoplasma diversity and potential insect host range are indeed underestimated and further large-scale investigation of leafhopper samples collected from natural habitats is needed.

## Materials and Methods

### *Collection and preservation of leafhoppers*

More than 3,000 bulk samples of sap-feeding hemipteran insects were obtained between 1998-2018 through field work by the last author, his students and colleagues during surveys that aimed to document poorly studied insect faunas in various parts of the world and to obtain representatives of all major lineages of Cicadellidae for use in phylogenetic and systematic studies. This material was supplemented by the first author’s collections in Europe between 2001-2018. Specimens were collected using various methods including sweeping and vacuuming of vegetation, night collecting at lights, and in Malaise (flight intercept) traps. Specimens

were collected directly into 95% ethanol in the field, returned to the laboratory and stored in  $-20^{\circ}\text{C}$  freezers at the INHS. Voucher specimens were also pinned for species identification and reference. Some samples included undescribed species from under-investigated areas, and they are waiting to be described in the context of other projects. In 2018, screening was carried out on a subset of 227 samples from independent sampling events in 28 countries (six continents) worldwide (Argentina, Australia, Bolivia, Brazil, Cameroon, Chile, China, Czech Republic, Ecuador, French Guiana, Ghana, Kyrgyzstan, India, Italy, Madagascar, Mongolia, Nicaragua, Papua New Guinea, Peru, Republic of Congo, Serbia, South Africa, Swaziland, Switzerland, Taiwan, Thailand, United States, Zambia). The land cover of the sampling events was analyzed using thematic maps within a geographic information system (QGIS 3.8, 2019) (Figure 1). Although 98% of the collections were intentionally obtained from natural areas or patches of native vegetation within more anthropogenic landscapes, we evaluated the land cover of a larger area including each sampling site using the raster layer Cropland and Pasture area (resolution 10 x 10 Km) (Ramankutty et al., 2008).

In total, the 227 samples encompassed about 1,000 specimens, with each species (or morphospecies) represented by 1 to 20 specimens belonging to the phloem-feeding leafhopper subfamily Deltocephalinae (except 1 sample belonging to the related hemipteran family Membracidae), which includes most of the previously documented vectors of phytoplasmas (Table S1). At least one specimen from each sample was selected randomly (with preference for males when present because species identification usually requires examination of male genitalia) for the molecular analyses.

#### *DNA extraction*

Total DNA was extracted from individual leafhoppers using a non-destructive method to preserve the specimen exoskeletons as vouchers and for subsequent morphological study. For each specimen, the abdomen was dissected, transferred to a 1.5 ml tube containing 400  $\mu\text{l}$  1X TES pH 7.8 buffer (20 mM Tris, 10 mM EDTA, 0.5% SDS) and 4  $\mu\text{l}$  Proteinase K (20 mg/ $\mu\text{l}$ ) and incubated at  $56^{\circ}\text{C}$  overnight. The abdomen was then removed and preserved in ethanol for morphological study. The buffer solution was then blended for 10 minutes using a mixer (MixMate) and the solution was transferred to a new 1.5 ml tube with 400  $\mu\text{l}$  of chloroform, mixed and centrifuged 10 min at  $4^{\circ}\text{C}$  at 11,000 rpm. The supernatant was transferred to a new tube and the chloroform wash was repeated. DNA was then transferred to a new tube and 400  $\mu\text{l}$  of ice-cold isopropanol was added followed by mixing and centrifuging for 15 min at  $4^{\circ}\text{C}$  at 12000 rpm. Supernatant was discarded and the DNA pellet was washed twice using 500  $\mu\text{l}$  of ice-cold 96% ethanol. The DNA pellet was then dried for 20 min and re-suspended in 50  $\mu\text{l}$  of TE buffer (pH 7.8). To each leafhopper sample a molecular code was assigned: e.g., LH078 stands for LeafHopper followed by an ordinal number indicating the collection event.

#### *Leafhopper species identification*

Specimens were sorted to morphospecies and tentatively identified by the last author prior to DNA extraction, with species identifications confirmed following non-destructive DNA extraction through examination of male genitalia. Exoskeletons of extracted specimens were saved as vouchers and deposited in the Illinois Natural History Survey insect collection. After the initial screening all the specimens that tested positive for the presence of phytoplasmas were identified by using published taxonomic keys and related literature (Emeljanov, 1967; Fletcher, 2000; Stiller, 2010; Zahniser, 2008). One of them was a new species for science and was recently described by the last author (Dietrich, in review). The abdomens of voucher specimens (males) were dissected to study the genitalia under an Olympus SZX10 stereoscopic microscope. Habitus photographs of voucher specimens were taken at INHS with a Canon SLR camera and 65 mm macro lens mounted on an automated lift.

#### *DNA amplification and sequencing of phytoplasmas*

TaqMan real time PCR (qPCR) analysis of the 16S ribosomal gene was carried out on DNA extracted from the 227 specimens to identify the presence of phytoplasmas, with the primers and probe described by Christensen et al. (2004). The assays were performed in 96-well plates on a CFX96 thermal cycler (Biorad), according to the protocol of Angelini et al. (2007). The reaction in 10  $\mu\text{l}$  contained 4  $\mu\text{l}$  of

DNA template diluted 1:2, 5  $\mu$ l Platinum Quantitative PCR Supermix-UDG (ThermoFisher scientific), 160 nM for each primer and probe. Because this protocol may yield false positives for other bacteria (e.g., *Bacillus* spp.), samples with Cq value [?] 30.38 (according to Christensen et al., 2004) were tested using nested PCR of the 16S ribosomal RNA gene to confirm the phytoplasma identity. In the 16S rRNA region, nested PCR was performed using universal primer pair P1/P7 (Deng & Hiruki 1991; Smart et al., 1996) followed by F2n/R2 (Gundersen & Lee, 1996). Amplicons were visualized on 1% agarose gel stained with GelRed (Biotium Inc.) under a GelDoc XR UV transilluminator (Biorad). The DNA of ALY (Italian alder yellows) phytoplasma, obtained from experimentally-infected periwinkle (*Catharanthus roseus*), was used as a positive reference strain in all the amplification reactions. Sequencing of the F2n/R2 amplicons was carried out in both directions using automated equipment (BMR Service, Padua, Italy). Forward and reverse reads were assembled using Gap4 and Pregap (Bonfield et al., 1995), followed by manual editing. Nucleotide sequences were deposited in the GenBank database under the accession numbers listed in Table 1. An initial BLAST query (Altschul et al., 1990) was performed in order to evaluate the similarity of newly obtained sequences to phytoplasma reference sequences, the top two similar sequences were included in the dataset for further phylogenetic analyses. The final reference sequence dataset consisted of 21 sequences obtained from the National Center for Biotechnology Information (NCBI) database (Federhen, 2012). The ingroup included 20 phytoplasma strains (11 described as ‘*Ca.*Phytoplasma’ species, including an incidental citation) representing different countries and isolated from distantly related hosts (Table S2) and the outgroup included *Acholeplasma palmae* (*Acholeplasmataceae*). Electropherograms were corrected and aligned using the Muscle algorithm as implemented in MEGA 7.0 (Edgar, 2004; Kumar et al., 2016) with default settings. Phylogenetic trees were constructed with the Maximum Likelihood (ML) and Neighbor Joining (NJ) methods. Branch support was measured using a bootstrap test with 1,000 replicates.

## Results

The 227 specimens analyzed belong to 9 tribes (Athysanini, Chiasmini, Deltocephalini, Macrostelini, Opsiini, Paralimnini, Pendarini, Scaphoideini, Scaphytopiini) which represent most of the groups of deltocephalinae comprising known phytoplasma vectors worldwide. Overall, about 49% of them (111 specimens) were identified to species during earlier sorting and preparation of collected samples, 2% are of uncertain species placement and ~43% represent undescribed species and genera or belong to genera for which comprehensive identification tools are not yet available. Thirteen species (6%) are represented by multiple specimens (Table S1).

GIS analyses with the Cropland and Pasture overlay confirmed that the sampling sites were located mainly in natural areas, with average raster values of  $0.091 \pm 0.13$  (compared to cropland raster value =1).

### *Detection and phylogenetic analysis of phytoplasmas*

Using qPCR on 227 leafhoppers, a positive signal was detected in 111 specimens. Only 14 samples with Cq value [?] 30.38 were selected for further analysis (Table S1). The nested PCR primed by F2n/R2 amplified fragments of the 16S rRNA gene of 1,200 bp from 6 phytoplasma-infected samples (Fig. S1). A lower or different sensitivity of the direct/nested primers compared to the ones used in qPCR may have caused the negative results for the remaining 8 samples. Overall, 6 species tested positive: *Leofa* (*Tortotettix*) *dispar* (Theron) (molecular sample codes: LH078), *Pravistylus exquadratus* (Naude) (LH082), *Macrosteles* (*Macrosteles*) *sordidipennis* (Stal) (LH102), *Mayawa capitata* (Kirkaldy) (LH133), *Mayawa affinisfacialis* Dietrich (LH139), and *Acharis ussuriensis* (Melichar) (LH143) (Table 1, Fig. 2 A-F).

The phylogenetic trees included 27 phytoplasma strains, and the alignment of 16S rRNA consisted of 952 positions (including gaps). The ML phylogenetic tree recovered our new sequences in three main clusters (A, B and C in Fig. 3). The first well-supported cluster (A) includes a monophyletic group of 4 samples from this study (LH078, LH082, LH139, LH143) and 4 strains belonging to 16SrXI phytoplasma group (2 ‘*Ca.* Phytoplasma sacchari’ and 2 Goosegrass white leaf phytoplasma strains) and the *Candidatus* species ‘*Ca.*Phytoplasma oryzae’ (16SrXI) + ‘*Ca.* Phytoplasma cynodontis’ (16SrXIV). Although the internal branches of this clade are very short with low bootstrap support, both samples from South

Africa (LH078, LH082) were recovered in the same subcluster. Samples from China (LH143) and Australia (LH139) are polyphyletic, with LH139 branching more deeply than LH143. A recent comprehensive ML tree for phytoplasmas recovered members of 16SrXI as paraphyletic with respect to 16SrXIV (Cao et al., 2020).

In the second cluster (B), LH133 is sister to ‘*Ca. Phytoplasma brasiliense*’ (16SrXV) and together these two strains are sister to the closely related phytoplasma strains in the 16SrII group. The last cluster (C) includes LH103 and members of the 16SrI phytoplasma group. The NJ analysis yielded the same topology (not shown).

Although the pairwise similarity of the sequences with closely related reference strains is >97.5%, further characterization revealed that all the phytoplasmas detected in this study represent new strains belonging to five new subgroups (Wei et al., in review).

## Discussion

### *Ecological and evolutionary context of the new phytoplasma-host associations*

None of the leafhopper species that tested positive for presence of phytoplasmas in the present study were previously reported as hosts or vectors of phytoplasmas (Trivellone, 2019). These 6 leafhopper specimens were collected from native grassy vegetation in four countries: South Africa, Kyrgyzstan, Australia, and China (Table 1 and Figure 1). *Leofa dispar* (LH078) and *P. exquadratus* (LH082) were collected from native grassland and fynbos vegetation in two different provinces in South Africa (KwaZulu-Natal and Western Cape) in 2004. The distance between these two sampling sites is about 1,120 km. In South Africa only four 16Sr phytoplasma groups were previously recorded (16SrI, 16SrII, 16SrIII, 16SrXII) (for an overview see Trivellone, 2019), and only two species of leafhoppers were recorded as potential vectors and competent vectors of phytoplasmas in the 16SrI group: *Austroagallia* sp. (subfamily Megophthalminae) and *Mgenia fuscovaria* (Stal) (Coelidiinae) (Kruger et al., 2015). Thus, this is the first record of phytoplasma strains in the clade 16SrXI/16SrXIV in South Africa. The leafhopper fauna of Africa is diverse but remains poorly known, with new genera and species continuing to be discovered (e.g., Stiller, 2019; 2020). *Pravistylus exquadratus* and other members of the same genus have never been reported as pests, except for single records of this species on Korog wheat cultivar and on rye grass (Stiller, 2010). The species is mainly associated with native grassland and fynbos vegetation, and it is always macropterous with high potential for dispersal. *Leofa dispar* also occurs in native grassland and has not been reported from crops. Both, *P. exquadratus* and *L. dispar* are restricted to South Africa.

*Macrosteles sordidipennis* (LH102) specimens were collected in a riparian sedge meadow, on a river bank in Kyrgyzstan (Jeti-Oguz District) in 1999. The single tested specimen LH102 was infected by a strain of group 16SrI related to aster yellows phytoplasma strains. Based on a recent review (see Trivellone, 2019), this appears to be the first record of this phytoplasma group in Kyrgyzstan. The only previous phytoplasma record from this country was the potato stolbur disease, associated with the 16SrXII phytoplasma group. Interestingly, other species in the genus *Macrosteles* have been reported as competent vectors of phytoplasmas (Trivellone, 2019). In particular, competent and potential vectors of *Macrosteles* show a strong cophylogenetic signal with the 16SrI phytoplasma group (Trivellone, unpublished data) suggesting that these two lineages have been associated for a long time. Nine additional species of *Macrosteles* have been documented in Kyrgyzstan (Novikov et al., 2000), including four that are competent vectors of 16SrI phytoplasmas in Europe, although 16SrI phytoplasmas have not been previously recorded from this country (Trivellone, 2018). Our discovery of a new association between a *Macrosteles* species not previously recorded as a phytoplasma host and a new 16SrI group strain or host suggest that further surveys and phytoplasma screening in Kyrgyzstan may be important for assessing the potential threat of emerging phytoplasma diseases in this region of Central Asia.

*Mayawa capitata* (LH133) and *M. affinifacialis* (LH139) were collected at lights from two different nature reserves in Australia in 2010 and 2009, respectively. The two sampling sites are about 3,597 km away from each other. *Mayawa capitata* belongs to the grass-specialist leafhopper tribe Paralimnini and reportedly occurs on grasses and *Sida acuta* (Malvaceae) (Fletcher, 2000). *Mayawa affinifacialis* has been recently

described (Dietrich, in review) and little is known about its ecology; however, the species was collected in grassland is likely a grass-feeding species. A specimen of the first species (LH133) was infected with a phytoplasma strain closely related to 16SrXV group and the second one (LH139) with a phytoplasma strain closely related to 16SrXI. Although the last group was never detected in Australia, only 3 competent vectors for phytoplasma strains in group 16SrII are known for this country, two species of *Orosius*, tribe Opsiini (Deltocephalinae), and *Batracomorphus angustatus* (Osborn) in the subfamily Iassinae (for an overview see Trivellone, 2019). A recent review of Australian phytoplasma pathosystems revealed an important gap of knowledge, with several recorded phytoplasma strains not yet assigned to 16Sr groups and subgroups (Liu et al., 2017). Moreover, information on competent vectors is scarce with many species still undescribed, hampering the understanding of epidemiological cycles. Our results expand the spectrum of potential vectors recorded in Australia to include species from the tribe Paralimnini, and reveal new possible epidemiological routes that require further investigation.

*Acharis ussuriensis* (LH143) was collected in China from grasses on a dry hillside at Zhouzhi Nature Reserve (Zhouzhi county, Shaanxi Province). The sampling location is entirely surrounded by forest with the nearest farming settlement about 10 Km away. The specimen testing positive was infected with a strain closely related to strains in the 16SrXI/16SrXIV groups (Figs 4, cluster A). Although both phytoplasma groups were previously detected in China, further investigation on the pattern of transmission and host plants involved in this pristine area will provide useful insights into the characterization of phytoplasma-host relationships in natural areas.

#### *Underestimated phytoplasma diversity*

##### *in natural areas*

Phytoplasmas are a highly diverse group of plant pathogens and new strains continue to be discovered at a steady pace worldwide but most such discoveries still mainly result from screening of plants showing “typical” phytoplasma disease symptoms in human-managed ecosystems.

In our study we unveiled 6 new associations between phytoplasmas and their insect hosts, recording new phytoplasma group records for 3 countries, and we detected new phytoplasma strains that will be further characterized and described separately (Wei et al., in review). Our screening also highlights that potential vectors collected in natural areas worldwide are poorly studied (as suggested by Trivellone & Dietrich, 2020) and potentially harbor phytoplasma species not yet discovered and described. Our discovery of new phytoplasma subgroup strains in natural areas worldwide is not surprising, given the >300-million-year history of co-evolution between phytoplasmas, their plant hosts, and insect vectors and the lack of previous attention to phytoplasmas in non-managed ecosystems (Cao et al., 2020; Trivellone & Dietrich 2020). According to a recent molecular timetree for phytoplasmas (Cao et al., 2020), the earliest divergences of phytoplasmas approximately coincided with those of their vascular plant hosts and some phytoplasma lineages are associated with particular major lineages of plants and hemipteran insects. If many such associations are evolutionarily conservative, then phylogenies may be useful tools for predicting undocumented associations between phytoplasmas, insects and plants. Also, because coevolutionary theory suggests that associations between parasites and their hosts should evolve toward commensalism over time (i.e., virulence should decrease; Alizon et al., 2009; Jansen et al., 2015), plants naturally infected by phytoplasmas in natural areas may not exhibit the classical symptoms of phytoplasma disease found in crop plants. Thus, many naturally occurring plant-phytoplasma associations may be asymptomatic, so screening of potential vectors and/or asymptomatic plants, may be necessary to reveal the true diversity of unknown phytoplasma strains in native ecosystems. Collections of leafhoppers used in our study were obtained in natural habitats. Although no evidence of diseased plant hosts was reported from the investigated sites, the collections were originally made for the purpose of documenting insect biodiversity, rather than within the context of plant pathogen surveys. For this reason, we cannot speculate on the disease epidemiology of phytoplasmas associated with leafhoppers tested for the present study. Further investigations are needed to document the host plants and phenotypic effects of phytoplasma infections for the newly documented strains.

Interestingly, 5 of the 6 leafhopper genera recorded here as phytoplasma hosts have not been previously reported as potential or competent vectors of phytoplasmas but all belong to tribes that include known phytoplasma vectors. Cophylogenetic studies may be used to predict new pathogen-host associations and emerging diseases (reviewed in Brooks et al., 2019). In our study 4 species in the tribe Paralimnini from 3 different countries were found to be associated with new phytoplasma strains related to the 16SrXI/16SrXIV and 16SrII/16SrXV groups. Previous studies have associated species of this tribe with groups 16SrI, 16SrXII, and 16SrIX (Trivellone, 2018). A previous phylogeny of Deltocephalinae recovered Paralimnini as a monophyletic group within a larger clade of leafhoppers that includes notorious groups of competent vectors of phytoplasma groups 16SrII, 16SrXI and 16SrXIV in the tribe Opsiini (Zahniser & Dietrich, 2013; Trivellone, 2018). Preliminary cophylogenetic analysis (Trivellone unpublished) suggests that potential host shifts of phytoplasma strains in the 16SrII/16SrXV, 16SrXI/16SrXIV groups may have occurred among species of deltocephaline leafhoppers. The newly documented associations reported here provide evidence to support this hypothesis. These associations remained undetected in natural areas until now.

Previous research showed that integrating different sources of knowledge is of paramount importance for discovering potentially emergent pathogens. Studies on zoonotic diseases showed that museum biorepositories represent an invaluable but still poorly utilized resource for pathogen discovery, due to the wealth of species represented and prevalent best practices of specimen preservation, identification and collecting-event description (Dunnum et al., 2017). Furthermore, existing databases and traditional ecological knowledge can contribute to discovery of the location and timing of potential spillover of pathogens into human-managed systems worldwide (Brook et al., 2009; Kutz et al., 2009).

Given that most previous research on phytoplasmas has been performed within the relatively narrow context of plant disease epidemiology in agroecosystems, we suggest that the diversity of phytoplasmas is severely underestimated and that natural areas worldwide should harbor a rich undiscovered diversity of phytoplasmas and their actual or potential insect vectors.

Similar phytoplasma infection prevalence in agroecosystems and natural grassland was previously reported in the literature; however, knowledge of the entire range of hosts (plants and insects) and symptoms caused by phytoplasmas in natural habitats remains inadequate (for a review see Trivellone & Dietrich, 2020).

#### *Museum biorepository as source of unknown phytoplasmas*

Plant, fungal and animal specimens deposited in natural history museums and public or private collections are becoming increasingly accessible due to web-based interfaces. Recently it was pointed out that the hundreds of millions of samples preserved in collections are useful for many purposes beyond their traditional uses in comparative morphology, taxonomy and biogeography (Meineke et al., 2019). Because they provide broad taxonomic, spatial and temporal coverage of Earth's biodiversity, such collections provide opportunities to analyze global changes and under-investigated areas. Species interactions documented by collections have mainly been investigated using metadata (e.g., Bartomeus et al., 2019; Meineke & Davies, 2019). The advent of increasingly sensitive molecular methods has recently allowed more cryptic symbiotic associations to be explored directly by the testing preserved tissues of potential hosts for presence of microbes and other symbionts (e.g., Daru et al., 2019). To our knowledge, this is the first time that phytoplasma-insect associations have been documented using museum specimens. Our screening confirmed presence of phytoplasmas in 6 leafhopper specimens (accounting for ~ 3% of the subset of 227 leafhopper analyzed). Most studies conducted until now in agroecosystems reported prevalence of infection on local or regional scales, mainly focusing on epidemiological cycles. Major examples from European crop fields (one or few plots) revealed prevalence of infection in single insect species hosts ranging from 0 to 52% for a single phytoplasma strain (as an example, 33-37.5% Acs et al., 2011; 7-13% Lessio et al., 2016; 0-52% Mitrovic et al., 2012; 0-35% Sforza et al. 1998; 6-50% Trivellone et al., 2005). Such studies employ extensive resources and sampling over a prolonged timespan during the growing season. Also, because these studies usually aim to document ongoing disease outbreaks, they are not directly comparable to our study, in which samples were collected in the absence of any prior evidence of phytoplasma infection at locations where the samples were collected. Because we mostly tested single specimens from collecting events spread over 20 years on multiple continents,

it is not surprising that most of our samples tested negative for presence of phytoplasmas. Our data do not allow us to speculate on local infection rates of the new strains detected. However, considering the spatial, temporal and taxonomic scale of the samples available in museum biorepositories, our results can be taken as a very rough, preliminary estimate of phytoplasma prevalence in natural areas worldwide and suggest that the undiscovered diversity of phytoplasmas in natural areas worldwide is substantial.

The present study provides strong evidence that both phytoplasma diversity and potential insect vector diversity are underestimated in natural habitats. Larger-scale studies of museum biorepositories will likely fill major gaps in our knowledge of this diversity, the evolution of phytoplasma-plant-vector associations and the potential for emergence of new pathogens of agricultural importance.

#### *Potential impact of vector-borne phytoplasma spillovers and large-scale future study*

Centuries of homogenization of agricultural production systems led to decreased genetic and species diversity of crops. Such general biological depletion was previously associated with increased pathogen outbreaks and serious economic losses in agroecosystems (Newton, 2016; King & Lively, 2012). Earlier research recognized the role of wildlife as natural reservoirs where infections are often asymptomatic. The onslaught of emerging infectious diseases in crops often involved alternative sources of inoculum and creation of new ecological interfaces, and global changes (e.g., land use or climate warming) set the stage for new associations to occur. Spillover events from natural habitats in direct contact with cultivated fields have been documented for several plant pathogens (McCann, 2020; Brooks et al., accepted), and the involvement of vectors may facilitate host shifts, accelerating the spread of diseases at the regional level. The phytoplasmas associated with Flavescence doree disease, and related strains, (FDp) represent one of the most well-studied pathosystems (Malembic-Maher et al., 2020), providing a good example of spillover from wild plants to a crop (*Vitis vinifera*) through efficient insect vectors (Trivellone & Dietrich 2020; Brooks et al., accepted). For other phytoplasma pathosystems, epidemiological information and characterization of strains associated with crops have accumulated for over forty years. However, information on genetic diversity, the range of hosts and ecological characteristics of the spreading of phytoplasmas in natural habitats are still broadly missing. This gap of knowledge hinders basic understanding of the evolution of phytoplasmas in association with their hosts, and hampers the implementation of proactive measures to cope with emerging pathogens.

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#### **Authors' contributions**

CHD and VT conceived the rationale, designed this study, and obtained the leafhopper samples. VT and LF performed the initial screening experiments using qPCR, direct and nested PCR, and analyzed the sequences. WW confirmed the phytoplasma identities. VT, CHD and WW drafted the manuscript. All authors contributed to revising and finalizing the manuscript.

#### **Data Availability Statement**

The sequences supporting the conclusions of this article were deposited into the NCBI under the accession numbers MW473669- MW473674.

#### **References**

- Acs, Z., Jović, J., Ember, I., Cvrković, T., Nagy, Z., Talaber, C., ... Koelber, M. (2011). First report of maize redness disease in Hungary. *Bulletin of Insectology*, 64. Retrieved from <https://plantarum.izbis.bg.ac.rs/handle/123456789/161>
- Alizon, S., Hurford, A., Mideo, N., & Van Baalen, M. (2009). Virulence evolution and the trade-off hypothesis: History, current state of affairs and the future. *Journal of Evolutionary Biology*, 22 (2), 245–259. doi:

10.1111/j.1420-9101.2008.01658.x

- Altschul, S. F., Gish, W., Miller, W., Myers, E. W., & Lipman, D. J. (1990). Basic local alignment search tool. *Journal of Molecular Biology* , 215 (3), 403–410. doi: 10.1016/S0022-2836(05)80360-2
- Angelini, E., Bianchi, G. L., Filippin, L., Morassutti, C., & Borgo, M. (2007). A new TaqMan method for the identification of phytoplasmas associated with grapevine yellows by real-time PCR assay. *Journal of Microbiological Methods* , 68 (3), 613–622. doi: 10.1016/j.mimet.2006.11.015
- Bartomeus, I., Stavert, J. R., Ward, D., & Aguado, O. (2019). Historical collections as a tool for assessing the global pollination crisis. *Philosophical Transactions of the Royal Society B: Biological Sciences* , 374 (1763), 20170389. doi: 10.1098/rstb.2017.0389
- Bonfield, J. K., Smith, K. f, & Staden, R. (1995). A new DNA sequence assembly program. *Nucleic Acids Research* , 23 (24), 4992–4999. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC307504/>
- Brooks, D. R., Hoberg, E. P., & Boeger, W. A. (2019). The Stockholm paradigm: Climate change and emerging disease. Chicago: The University of Chicago Press.
- Brooks, D. R., Hoberg, E. P., Boeger, W. A., Gardner, S. L., Araujo, S. B. L., Bajer, K., Botero-Cañola, S., Byrd, B., Földvári, G., Cook, J. A., Dunnun, J. L., Dursahinhan, A. T., Garamszegi, L. Z., Herczeg, D., Jakab, F., Juarrero, A., Kemenesi, G., Kurucz, K., León-Règagnon, V., ... Trivellone, V. (2020). Before the Pandemic Ends: Making Sure This Never Happens Again. *World Complexity Science Academy* ,1 (1), 1–10.
- Brooks D. R., Hoberg E. P., Boeger W. A., & Trivellone V. (accepted). Emerging infectious disease: an underappreciated area of strategic concern for food security. *Transboundary and Emerging Diseases* .
- Cao, Y., Trivellone, V., & Dietrich, C. H. (2020). A timetree for phytoplasmas (Mollicutes) with new insights on patterns of evolution and diversification. *Molecular Phylogenetics and Evolution* ,149 , 106826. doi: 10.1016/j.ympev.2020.106826
- Christensen, N. M., Nicolaisen, M., Hansen, M., & Schulz, A. (2004). Distribution of phytoplasmas in infected plants as revealed by real-time PCR and bioimaging. *Molecular Plant-Microbe Interactions: MPMI* ,17 (11), 1175–1184. doi: 10.1094/MPMI.2004.17.11.1175
- Daru, B. H., Bowman, E. A., Pfister, D. H., & Arnold, A. E. (2019). A novel proof of concept for capturing the diversity of endophytic fungi preserved in herbarium specimens. *Philosophical Transactions of the Royal Society B: Biological Sciences* , 374 (1763), 20170395. doi: 10.1098/rstb.2017.0395
- Deng, S., & Hiruki, C. (1991). Amplification of 16S rRNA genes from culturable and nonculturable Mollicutes. *Journal of Microbiological Methods* , 14 (1), 53–61. doi: 10.1016/0167-7012(91)90007-D
- Dietrich, C. H. (in review). New species of *Mayawa* Fletcher and description of a related new Australian leafhopper genus (Hemiptera: Cicadellidae: Deltocephalinae: Paralimnini). *Zootaxa*
- DiEuliis, D., Johnson, K. R., Morse, S. S., & Schindel, D. E. (2016). Opinion: Specimen collections should have a much bigger role in infectious disease research and response. *Proceedings of the National Academy of Sciences* , 113 (1), 4–7. doi: 10.1073/pnas.1522680112
- Dunnun, J. L., Yanagihara, R., Johnson, K. M., Armien, B., Batsaikhan, N., Morgan, L., & Cook, J. A. (2017). Biospecimen Repositories and Integrated Databases as Critical Infrastructure for Pathogen Discovery and Pathobiology Research. *PLOS Neglected Tropical Diseases* ,11 (1), e0005133. doi: 10.1371/journal.pntd.0005133
- Edgar, R. C. (2004). MUSCLE: Multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Research* ,32 (5), 1792–1797. doi: 10.1093/nar/gkh340
- Emeljanov, A. F. (1967). Suborder Cicadinea (Auchenorrhyncha). In GY. Bei-Bienko (Ed.), Keys to the insects of the European USSR. I. Apterygota. Palaeoptera. Hemimetabola. (pp. 421–551). Moscow-Leningrad:

Nauka.

- Federhen, S., 2012. The NCBI Taxonomy database. *Nucleic Acids Res.* 2012; 40 (Database issue): D136–43. <http://www.ncbi.nlm.nih.gov/taxonomy> (accessed 28 December 2021).
- Fletcher, M. J. (2000). A new genus, *Mayawa*, for the reception of *Limotettix capitatus* Kirkaldy (Hemiptera: Cicadellidae: Deltocephalinae) and descriptions of five new species of *Mayawa*. *Australian Journal of Entomology*, 39 (3), 103–110. doi: 10.1046/j.1440-6055.2000.00158.x
- Gundersen, D. E., & Lee, I.-M. (1996). Ultrasensitive detection of phytoplasmas by nested-PCR assays using two universal primer pairs. *Phytopathologia Mediterranea*, 35 (3), 144–151. JSTOR. Retrieved from <https://www.jstor.org/stable/42685262>
- Gupta, R. S., Sawnani, S., Adeolu, M., Alnajjar, S., & Oren, A. (2018). Phylogenetic framework for the phylum Tenericutes based on genome sequence data: Proposal for the creation of a new order Mycoplasmoidales ord. nov., containing two new families Mycoplasmoidaceae fam. nov. and Metamycoplasmataceae fam. nov. harbouring Eperythrozoon, Ureaplasma and five novel genera. *Antonie van Leeuwenhoek*, 111 (9), 1583–1630. doi: 10.1007/s10482-018-1047-3
- Hogenhout, S. A., Oshima, K., Ammar, E. D., Kakizawa, S., Kingdom, H. N., & Namba, S. (2008). Phytoplasmas: bacteria that manipulate plants and insects. *Molecular Plant Pathology*, 9 (4), 403–423. <https://doi.org/10.1111/j.1364-3703.2008.00472.x>
- Huang, W., Reyes-Caldas, P., Mann, M., Seifbarghi, S., Kahn, A., Almeida, R. P. P., Béven, L., ... Coaker, G. (2020). Bacterial Vector-Borne Plant Diseases: Unanswered Questions and Future Directions. *Molecular Plant*, 13 (10), 1379–1393. doi: 10.1016/j.molp.2020.08.010
- IRPCM Phytoplasma/Spiroplasma Working Team–Phytoplasma Taxonomy Group (2004). ‘Candidatus Phytoplasma’, a taxon for the wall-less, non-helical prokaryotes that colonize plant phloem and insects. *International Journal of Systematic and Evolutionary Microbiology*, 54 (4), 1243–1255. doi: 10.1099/ijms.0.02854-0
- Jansen, G., Crummenerl, L. L., Gilbert, F., Mohr, T., Pfefferkorn, R., Thänert, R., ... Schulenburg, H. (2015). Evolutionary Transition from Pathogenicity to Commensalism: Global Regulator Mutations Mediate Fitness Gains through Virulence Attenuation. *Molecular Biology and Evolution*, 32 (11), 2883–2896. doi: 10.1093/molbev/msv160
- King, K. C., & Lively, C. M. (2012). Does genetic diversity limit disease spread in natural host populations? *Heredity*, 109 (4), 199–203. doi: 10.1038/hdy.2012.33
- Kirdat, K., Tiwarekar, B., Thorat, V., Sathe, S., Shouche, Y., & Yadav, A. (2020). ‘Candidatus Phytoplasma sacchari’, a novel taxon-associated with Sugarcane Grassy Shoot (SCGS) disease. *International Journal of Systematic and Evolutionary Microbiology*. doi: 10.1099/ijsem.0.004591
- Kruger, K., Pietersen, G., Smit, N., Carstens, R. 2015. Epidemiology of aster yellows phytoplasma: alternate host plants and the vector *Mgenia fuscovaria* (Hemiptera: Cicadellidae) in South Africa. In *Proceedings of the 18th Meeting of the International Council for the study of Virus-like diseases of the Grapevine (ICVG)*, September 7-11, Ankara, Turkey, pp. 126-127.
- Kumar, S., Stecher, G., & Tamura, K. (2016). MEGA7: Molecular Evolutionary Genetics Analysis Version 7.0 for Bigger Datasets. *Molecular Biology and Evolution*, 33 (7), 1870–1874. doi: 10.1093/molbev/msw054
- Kutz, S. J., Jenkins, E. J., Veitch, A. M., Ducrocq, J., Polley, L., Elkin, B., & Lair, S. (2009). The Arctic as a model for anticipating, preventing, and mitigating climate change impacts on host–parasite interactions. *Veterinary Parasitology*, 163 (3), 217–228. doi: 10.1016/j.vetpar.2009.06.008
- Lee, I. M., Gundersen-Rindal, D. E., Davis, R. E., & Bartoszyk, I. M. (1998). Revised classification scheme of phytoplasmas based on RFLP analyses of 16S rRNA and ribosomal protein gene sequences. *International Journal of Systematic and Evolutionary Microbiology*, 48 (4), 1153–1169. doi: 10.1099/00207713-48-4-1153

- Lee, I. M., Davis, R. E., & Gundersen-Rindal, D. E. (2000). Phytoplasma: phytopathogenic mollicutes. *Annual Reviews in Microbiology* , 54 (1), 221-255. doi: 10.1146/annurev.micro.54.1.221
- Lessio, F., Picciau, L., Gonella, E., Mandrioli, M., Tota, F., & Alma, A. (2016). The mosaic leafhopper *Orientalus ishidae*: Host plants, spatial distribution, infectivity, and transmission of 16SrV phytoplasmas to vines. *Bulletin of Insectology* , 69 (2), 277–289.
- Liu, J., Gopurenko, D., Fletcher, M. J., Johnson, A. C., & Gurr, G. M. (2017). Phytoplasmas—the “crouching tiger” threat of Australian plant pathology. *Frontiers in Plant Science* , 8 , 599. doi: 10.3389/fpls.2017.00599
- McCann, H. C. (2020). Skirmish or war: The emergence of agricultural plant pathogens. *Current Opinion in Plant Biology* , 56 , 147–152. doi: 10.1016/j.pbi.2020.06.003
- MacLean, A. M., Sugio, A., Makarova, O. V., Findlay, K. C., Grieve, V. M., Tóth, R., ... Hogenhout, S. A. (2011). Phytoplasma effector SAP54 induces indeterminate leaf-like flower development in Arabidopsis plants. *Plant Physiology* , 157 (2), 831-841.
- MacLean, A. M., Orlovskis, Z., Kowitwanich, K., Zdziarska, A. M., Angenent, G. C., Immink, R. G., & Hogenhout, S. A., 2014. Phytoplasma effector SAP54 hijacks plant reproduction by degrading MADS-box proteins and promotes insect colonization in a RAD23-dependent manner. *PLoS Biology* , 12 (4), e1001835. doi: 10.1371/journal.pbio.1001835
- Malembic-Maher, S., Desqué, D., Khalil, D., Salar, P., Bergey, B., Danet, J.-L., ... Foissac, X. (2020). When a Palearctic bacterium meets a Nearctic insect vector: Genetic and ecological insights into the emergence of the grapevine Flavescence dorée epidemics in Europe. *PLOS Pathogens* , 16 (3), e1007967. doi: 10.1371/journal.ppat.1007967
- Meineke, E. K., & Davies, T. J. (2019). Museum specimens provide novel insights into changing plant–herbivore interactions. *Philosophical Transactions of the Royal Society B: Biological Sciences* , 374 (1763), 20170393. doi: 10.1098/rstb.2017.0393
- Meineke, E. K., Davies, T. J., Daru, B. H., & Davis, C. C. (2019). Biological collections for understanding biodiversity in the Anthropocene. *Philosophical Transactions of the Royal Society B: Biological Sciences* , 374 (1763). doi: 10.1098/rstb.2017.0386
- Mitrović, M., Jović, J., Cvrković, T., Krstić, O., Trkulja, N., & Toševski, I. (2012). Characterisation of a 16SrII phytoplasma strain associated with bushy stunt of hawkweed ox tongue (*Picris hieracioides*) in south-eastern Serbia and the role of the leafhopper *Neolaliturus fenestratus* (Deltocephalinae) as a natural vector. *European Journal of Plant Pathology* , 134 (3), 647–660. doi: 10.1007/s10658-012-0044-z
- Naderali, N., Nejat, N., Vadamalai, G., Davis, R.E., Wei, W., Harrison, ... Zhao, Y. (2017). ‘*Candidatus* Phytoplasma wodyetiae’, a new taxon associated with yellow decline disease of foxtail palm (*Wodyetia bifurcata*) in Malaysia. *International Journal of Systematic and Evolutionary Microbiology* , 67 (10), pp.3765-3772. doi: 10.1099/ijsem.0.002187
- Newton, A. C. (2016). Exploitation of Diversity within Crops—The Key to Disease Tolerance? *Frontiers in Plant Science* , 7 . doi: 10.3389/fpls.2016.00665
- Novikov, D. V., Novikova, N. V., Anufriev, G. A., & Dietrich, C. H. (2006). Auchenorrhyncha (Hemiptera) of Kyrgyz grasslands. *Russian Entomological Journal* , 15 (3), 303–310.
- QGIS Development Team. (2019). QGIS (Version 3.10-Bonn). OpeSource Geospatial Foundation Project. <https://qgis.org/en/site/>.
- Ramankutty, N., Evan, A. T., Monfreda, C., & Foley, J. A. (2008). Farming the planet: 1. Geographic distribution of global agricultural lands in the year 2000. *Global Biogeochemical Cycles* , 22 (1). doi:10.1029/2007GB002952.

- Rao, G. P., Bertaccini, A., Fiore, N., & Liefting, L. W. (2018). *Phytoplasmas: Plant Pathogenic Bacteria - I: Characterisation and Epidemiology of Phytoplasma - Associated Diseases*. Singapore: Springer. <https://doi.org/10.1007/978-981-13-0119-3>
- Rodrigues Jardim, B., Kinoti, W. M., Tran-Nguyen, L. T., Gambley, C., Rodoni, B., & Constable, F.E. (2020). ‘*Candidatus* Phytoplasma stylosanthis’, a novel taxon with a diverse host range in Australia, characterised using multilocus sequence analysis of 16S rRNA, secA, tuf, and rp genes. *International Journal of Systematic and Evolutionary Microbiology*, doi: 10.1099/ijsem.0.004589
- Šafářová, D., Zemánek, T., Válová, P., & Navrátil, M. (2016.) ‘*Candidatus* Phytoplasma cirsií’, a novel taxon from creeping thistle [*Cirsium arvense* (L.) Scop.]. *International journal of systematic and evolutionary microbiology*, 66 (4), pp.1745-1753. doi: 10.1099/ijsem.0.000937
- Seemüller, E., Marcone, C., Lauer, U., Ragozzino, A., & Göschl, M. (1998). Current status of molecular classification of the phytoplasmas. *Journal of Plant Pathology*, 3–26.
- Sforza, R., Clair, D., Daire, X., Larrue, J., & Boudon-Padiou, E. (1998). The Role of *Hyalesthes obsoletus* (Hemiptera: Cixiidae) in the Occurrence of Bois noir of Grapevines in France. *Journal of Phytopathology*, 146 (11–12), 549–556. doi: <https://doi.org/10.1111/j.1439-0434.1998.tb04753.x>
- Smart, C. D., Schneider, B., Blomquist, C. L., Guerra, L. J., Harrison, N. A., Ahrens, U., ... Kirkpatrick, B. C. (1996). Phytoplasma-specific PCR primers based on sequences of the 16S-23S rRNA spacer region. *Applied and Environmental Microbiology*, 62 (8), 2988–2993. Retrieved from <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC168085/>
- Stiller, M. (2010). Revision of the Southern African leafhopper genus *Pravistylus* (Hemiptera, Cicadellidae, Deltocephalinae). *Zootaxa*, 2468, 1–81.
- Stiller, M. (2019). A new leafhopper genus *Discolopeus* and nine new species (Hemiptera, Cicadellidae, Deltocephalinae) associated with shrubs, trees and poisonous plants in South Africa. *Zootaxa*, 4559 (2), 201-244.
- Stiller, M. (2020). A new leafhopper genus *Geelus* and 12 new species (Hemiptera, Cicadellidae, Deltocephalinae) from Southern Africa. *Zootaxa*, 4786 (3), 301-344.
- Trivellone, V., Pinzauti, F., & Bagnoli, B. (2005). *Reptalus quinquecostatus* (Dufour) (Auchenorrhyncha Cixiidae) as a possible vector of Stolbur-phytoplasma in a vineyard in Tuscany. *Redia*, 88, 103–108.
- Trivellone, V. (2018). Hemiptera–Phytoplasma–Plant database links. Retrieved from <http://trivellone.speciesfile.org/>
- Trivellone, V. (2019). An online global database of Hemiptera-Phytoplasma-Plant biological interactions. *Biodiversity data journal*, (7). doi: 10.3897/BDJ.7.e32910.2
- Trivellone V., & Dietrich C.H. 2020. Evolutionary Diversification in Insect Vector–Phytoplasma–Plant Associations. *Annals of the Entomological Society of America*, doi: 10.1093/aesa/saaa048
- Wei, W., Davis, R. E., Lee, I. M., & Zhao, Y. (2007). Computer simulated RFLP analysis of 16S rRNA genes: identification of ten new phytoplasma groups. *International Journal of Systematic and Evolutionary Microbiology*, 57 (8), 1855–1867. doi: 10.1099/ijms.0.65000-0
- Wei, W., Davis, R. E., Nuss, D. L., & Zhao, Y. (2013). Phytoplasmal infection derails genetically preprogrammed meristem fate and alters plant architecture. *Proceedings of the National Academy of Sciences of the United States of America*, 110 (47), 19149-54.
- Wei, W., Davis, R. E., Bauchan, G.R., & Zhao, Y. (2019) New symptoms identified in phytoplasma-infected plants reveal extra stages of pathogen-induced meristem fate-derailment. *Molecular Plant-Microbe Interactions: MPMI*, 32 (10), 1314–1323. <https://doi.org/10.1094/MPMI-01-19-0035-R>

Wei, W., Trivellone, V., Dietrich, C. H., Zhao, Y., Bottner, K., & Ivanauskas, A. (in review). Identification of phytoplasmas representing four distinct taxonomic 16Sr groups from phloem-feeding leafhoppers highlight the diversity of phytoplasmas and their potential vectors. *Pathogens* .

Weintraub, P. G., & Beanland, L. (2006). Insect vectors of phytoplasmas. *Annual Review of Entomology* , 51, 91–111. <https://doi.org/10.1146/annurev.ento.51.110104.151039>

Zahniser, J. N. (2008). Seven new species and new distributions of Old World Chiasmini (Hemiptera: Cicadellidae: Deltocephalinae), with a redescription, key to genera, and species checklist for the tribe. *Zootaxa* , 1808 , 1–32.

Zahniser, J. N., & Dietrich, C. (2013). A review of the tribes of Deltocephalinae (Hemiptera: Auchenorrhyncha: Cicadellidae). *European Journal of Taxonomy* , (45). doi: 10.5852/ejt.2013.45

Zhao, Y., Wei, W., Lee, I. M., Shao, J., Suo, X., & Davis, R. E. (2009). Construction of an interactive online phytoplasma classification tool, iPhyClassifier, and its application in analysis of the peach X-disease phytoplasma group (16SrIII). *International Journal of Systematic and Evolutionary Microbiology* , 59 (10), 2582–2593. doi: 10.1099/ijs.0.010249-0

Zhao, Y., Davis, R. E., Wei, W., & Lee, I.-M. (2015). Should “Candidatus Phytoplasma” be retained within the order Acholeplasmatales? *International Journal of Systematic and Evolutionary Microbiology* , 65 (3), 1075–1082. doi: 10.1099/ijs.0.000050

Zhao, Y., Wei, W., Davis, R. E., Lee, I. M., & Bottner-Parker, K. D. (2021). The agent associated with blue dwarf disease in wheat represents a new phytoplasma taxon, ‘Candidatus Phytoplasma tritici’. *International Journal of Systematic and Evolutionary Microbiology* , p.ijsem004604

Zwolińska, A., Krawczyk, K., Borodynko-Filas, N., & Pospieszny, H. (2019). Non-crop sources of Rapeseed Phyllody phytoplasma (‘Candidatus Phytoplasma asteris’: 16SrI-B and 16SrI-(B/L)L), and closely related strains. *Crop Protection* , 119 , 59–68. <https://doi.org/10.1016/j.cropro.2018.11.015>

Table 1. List of species collected in natural areas that tested positive for the presence of phytoplasmas. Description of locations of the new associations between phytoplasmas and insect hosts detected in this study. The last column reports the 16Sr phytoplasma group detected in the leafhoppers collected from natural habitats and the GenBank accession numbers of the deposited sequences.

Code	Tribe	Species	Country	Coordinate	Habitat, altitude (m a.s.l.)	Date Collection method Collector	Phytoplasma (Accession number)
LH78	Chiasmini	<i>Leofa dispar</i>	South Africa	28°53’59”S 29°26’05”E	Grassland, 1,583	27 Dec 2004 Sweep net J.N. Zahniser	16Sr XI/XIV (MW473669)
LH82	Paralimnini	<i>Pravistylus exquadratus</i>	South Africa	33°51’01”S 19°03’16”E	Fynbos, 201	15 Dec 2004 Vacuum J.N. Zahniser	16Sr XI/XIV (MW473673)
LH102	Macrostelini	<i>Macrosteles sordidipennis</i>	Kyrgyzstan	41°47’52”N 78°39’44”E	Sedge meadow 2,950	04 Jul 1999 Vacuum D. Novikonv & C.H. Dietrich	16Sr I (MW473674)

Code	Tribe	Species	Country	Coordinate	Habitat, altitude (m a.s.l.)	Date Collection method Collector	Phytoplasma (Accession number)
LH133	Paralimnini	<i>Mayawa capitata</i>	Australia	32°57'06"S 115°54'49"E	Yarloop Nature Reserve, 76	10 Jan 2010 Sweep net K. Hill, et. al.	16Sr II/XV (MW473671)
LH139	Paralimnini	<i>Mayawa affinisfacialis</i>	Australia	27°56'03"S 153°04'42"E	Flagstone Creek Reserve Park, on grassland, 50	04 Jan 2009 Sweep net K. Hill, et. al.	16Sr XIV (MW473672)
LH143	Paralimnini	<i>Acharis ussuriensis</i>	China	33°58'53"N 108°09'50"E	Forest Natural Reserve, 660	12 July 2012 Vacuum Wei Cong	16Sr XI/XIV (MW473670)



Figure 1. Map of the sampling sites of the 227 leafhopper samples screened in the present study. Symbols indicating the Cq values results of qPCR (small empty circle: negative; big empty circle: Cq >41; big black circle: 35.54 < Cq < 40; empty triangle: 31 < Cq < 35.52; black triangle Cq < 30). Map created QGIS 3.8 and was modified with Adobe Photoshop CC 2019. This map is licensed under an X/MIT style Open-Source License by the Open Source Geospatial Foundation.



Figure 2. Dorsal views of the 6 species of leafhoppers that tested positive for phytoplasmas. A, *Acharis ussuriensis* (Melichar) (LH143); B, *Leofa (Tortotettix) dispar* (Theron) (molecular code, LH078); C, *Macrosteles (Macrosteles) sordidipennis* (Stål) (LH102); D, *Mayawa capitata* (Kirkaldy) (LH133); E, *Mayawa affini-facialis* Dietrich (LH139); F, *Pravistylus exquadratus* (Naudé) (LH082). Scale bar 1.0 mm.

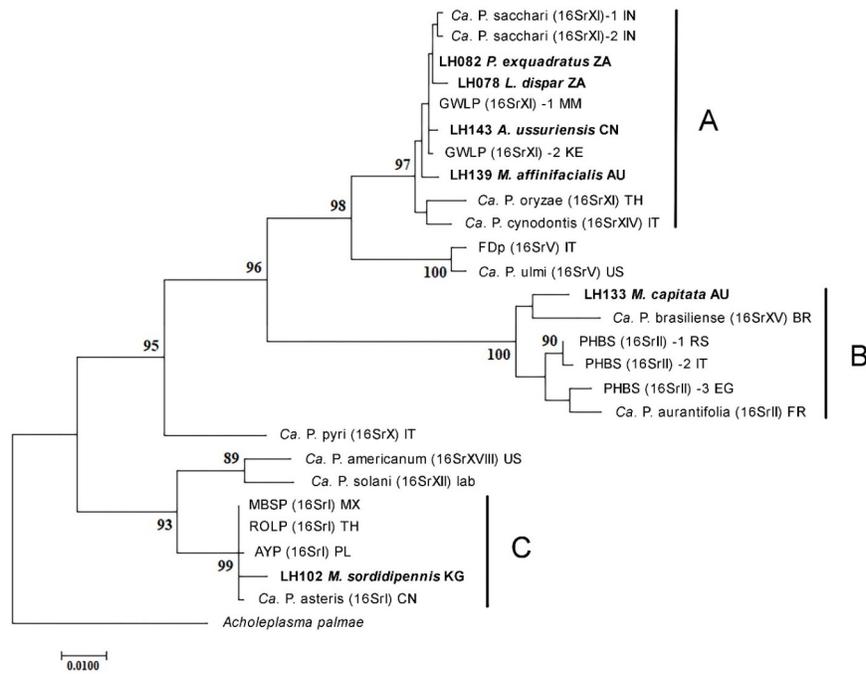


Figure 3. Maximum likelihood tree based on 952 positions of the F2n/R2 fragment of the *16S rRNA* gene obtained from 6 samples of the present study (in bold), 20 phytoplasma strains from GenBank (used as references) and *Acholeplasma palmae* (outgroup). Bootstrap values (> 63%) are shown above or below the branches. Branch lengths are proportional based on the scale indicated. GenBank accession numbers and details of the reference phytoplasma strains are listed in Supplementary Table S2. The names at the tip of the tree include the following: the phytoplasma strain (acronym or *Candidatus* species name), the 16Sr phytoplasma group in parenthesis or the name of the insect species host, and the Country Code where the strain was detected (AU, Australia; BR, Brazil; CN, China; EG, Egypt; FR, France; IT, Italy; IN, India; KE, Kenya; KG, Kyrgyzstan; MM, Myanmar; MX, Mexico; PL, Poland; RS, Serbia; TH, Thailand; US, United States; ZA, South Africa). lab: laboratory-maintained strain. A, B and C indicate the clusters that include the samples from this study.

## Supplementary material

Figure S1. Agarose gel (1%) showing F2n/R2 amplicons obtained in nested PCR for 14 leafhopper samples of study. B1: blanks, negative control; ALY, alder yellow phytoplasma; M: marker of molecular weight, 1 kb DNA ladder (RBC Bioscience). Numbers refer to the leafhopper IDs in Table S1.

Table S1. List of 227 samples of leafhoppers analyzed in 2018. All taxa belong to subfamily Deltocephalinae (Hemiptera, Cicadellidae) except the species 216 (ID) belonging to Hemiptera in family Membracidae. Each record (ID) represents a sample of 1 or more specimens of one species (columns E-F) collected in an independent collecting event (columns I-O). Results of the qPCR and nested PCR are reported in columns B-C.

Table S2. List of 16S rRNA gene sequences downloaded from GenBank and used as reference strains in this study.



