Response Manipulation vs Diurnal Observation: A Direct Comparison of Two Methods for Quantifying Stomatal Behavior in Tropical Forests

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Abstract

A primary source of uncertainty in terrestrial biosphere model (TBM) projection of carbon uptake and water cycling from ecosystems is the relationship between CO_2 assimilation (A) and water loss via stomatal conductance (g_s). A common mathematical framework for modeling this relationship is the "Unified Stomatal model", which relates A to g_s over environmental conditions and is governed by two terms, the stomatal slope (g_1) and intercept (g_0). Given their importance in determining the relationship between forest productivity and climate, an accurate and mechanistic understanding of the g_1 and g_0 parameters is crucial, particularly in wet tropical broadleaf forests where changes in water cycling could impact global weather patterns. These stomatal parameters are estimated using leaf-level gas exchange by two alternative methods: (1) a response curve where the environmental conditions are modified for a single leaf, or (2) a survey approach, where repeated measurements are made on multiple leaves over a diurnal range of environmental conditions. We compare the curve and survey approaches by conducting a comprehensive measurement campaign in which we paired diurnal gas exchange surveys with leaf level response curves for the estimation of g_1 and g_0 on six tropical species across a full range of leaf phenological stages. We examine how these different estimates impact model projection of g_s, and how the consideration of a diurnal effect on g_1 and g_0 can improve predictions relative to a model using parameter estimates which are fixed over the photoperiod. Our results showed that age is an important factor to consider in estimates of g_0, however there was no effect of leaf age on estimates of g_1. The survey approach identified a diurnal trend associated with g_1 and g_0, which when accounted for improved model projections of diurnal trends in g_s. We found that while both approaches yield equally statistically valid estimates of g_1 and g_0 at a fixed point in time, they are not directly comparable across diurnal timescales, where shifting water supply and carbon demand lead to dynamic canopy scale water use efficiency (WUE). These results suggest that to improve the accuracy of modelled g_s in tropical forests, TBMs should recognize and implement diurnal variation in stomatal parameters which are associated with diurnal shifts in WUE.

Response Manipulation vs Diurnal Observation: A Direct Comparison of Two Methods for Quantifying Stomatal Behavior in Tropical Forests

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BACKGROUND

Terrestrial Land Sensor

One of the largest uncertainties in current model predictions of plant water uptake and water cycling is how stomata in the terrestrial biosphere will respond to increasing atmospheric CO2 concentrations (Ca) (Lin et al., 2015). Ca is the driving force behind the Ball-Berry stomatal conductance model (Ball et al., 1987), which is used in many land surface models (e.g. CLM, ED2, SiB3, SiB4) (Lin et al., 2015). Ball-Berry model predicts stomatal conductance (gs) as a function of Ca, relative humidity (h), and the slope of the relationship between stomatal conductance and photosynthesis (m):

$$g_s = m \frac{A_n h}{C_a} + b$$

In this model, a doubling atmospheric [CO2] will cause a 50% decrease in the slope (m) of gs and a 50% increase in the slope (m) of the relationship between gs and An (Lin et al., 2015). Although this model is widely used, it has been shown to overestimate the sensitivity of gs to increasing Ca (Lin et al., 2015). Therefore, it is important to understand how stomata will respond to increasing Ca in order to improve the accuracy of land surface models.

Prevailing view on stomatal effects on transpiration

Numerous studies have shown a strong response to elevated [CO2] (Ca) in which stomatal closure by plants leads to a reduction in transpiration (E) (Ainsworth & Rogers, 2007; Lin et al., 2015; Medlyn et al., 2001). Reduced E can increase soil moisture, leaf temperature, and water use efficiency (WUE) (Ainsworth & Rogers, 2007; Lin et al., 2015; Medlyn et al., 2001).

Stomatal sensing mechanisms?

Conflicting data from stomatal conditions from short-term stomatal responses, i.e. in which stomata do not acclimate, i.e. Free-Air CO2 Enrichment (FACE) experiments, suggest that stomata may respond to changes in the rate of change of Ca, rather than absolute Ca (Eamus et al., 2013; Lin et al., 2015). Other studies suggest that stomata may respond to changes in the internal CO2 concentration (Ci) (Lin et al., 2015). Ci is the CO2 concentration in the leaf airspace. Ci is the CO2 concentration at the site of carboxylation, specifically the region of the chloroplast where photosynthesis occurs. Ci strongly affects stomatal conductance.

Objective

The objective of this research was to compare the response of gs, using the Ball-Berry (BB) Free Stomata Model (FSM), Baseline, and Free Stomata Model Optimization (FSMO) under three different scenarios:

1. Ambient Ca (aCa), Ca+ 200 ppm (eCa), and Ca+ 400 ppm (e2Ca)
2. Ambient Ca (aCa), Ca+ 200 ppm (eCa), and Ca+ 400 ppm (e2Ca) with a 2 hour step change
3. Ambient Ca (aCa), Ca+ 200 ppm (eCa), and Ca+ 400 ppm (e2Ca) with a 2 hour step change and a 2 hour recovery period

METHODS

Study Site. Data was collected at Brookhaven National Laboratory's Forest Research Area (BNL) (40.87° N, 72.87° W), using Free-Air Carbon Dioxide Enrichment (FACE) technology (Brookhaven National Laboratory, n.d.). Established in 1988, the FACE facility is located in the Pine Barrens region of Long Island, New York. The forest is dominated by oak species (*Quercus* spp.) and pitch pine (*Pinus rigida*) (Brookhaven National Laboratory, n.d.).

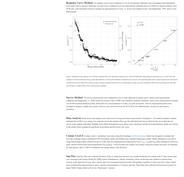

Figure 1. Location of BNL in relation to the surrounding region (Brookhaven National Laboratory, n.d.).

Canopy Access. Data for this project was collected from the Canopy Access Array (CAA) (Serbin et al., 2019). The CAA is a 30m tall tower that is surrounded by a 3-dimensional scaffolding system. The scaffolding system allows researchers to access the canopy at different heights (Serbin et al., 2019).

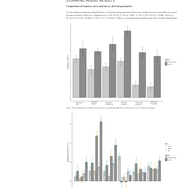

Figure 2. Canopy Access Array (CAA) at Brookhaven National Laboratory (Serbin et al., 2019).

Study Species. Six species were selected for study and data was collected in each species over the course of the growing season. For each species, data was collected from 3 individuals in each of the 3 CO2 treatments (aCa, eCa, e2Ca).

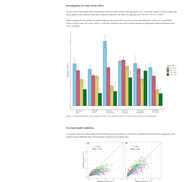

Figure 3. Species selected for study. From left to right: Black Oak (*Quercus velutina*), Scarlet Oak (*Quercus coccinea*), White Oak (*Quercus alba*), Pitch Pine (*Pinus rigida*), Sassafras (*Sassafras albidum*), Black Cherry (*Prunus serotina*).

Empirical Data Collection. All species were collected during the growing season. Measurements were collected with LI-6800 Portable Photosynthesis System (LI-COR Biosciences, Lincoln, NE, USA). A 6 cm2 chamber was used for all measurements.

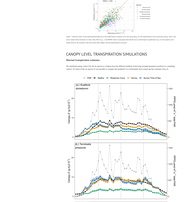

Figure 4. Diagram of the LI-6800 Portable Photosynthesis System (LI-COR Biosciences, n.d.). The LI-6800 Portable Photosynthesis System was used to collect data for this project.

Leaf Enclosure. Leaf temperature was controlled using the LI-6800's built-in LED-RGB photosynthetic system (LI-COR Biosciences, n.d.). Flow was set to 600 µmol/s and relative humidity to 60%.

Empirical Data Processing. All datasets were cross-validated to ensure accurate datasets. Records that contained leaf temperature, photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained negative values for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained values of 0 for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed.

Figure 5. Example of data collected during the 2 hour step change period. The step change period occurred between 10:00 and 12:00. Data was collected every 10 seconds. The step change period was removed from the dataset.

Step Change. Each species was subjected to a step change in Ca (ΔCa) (Eamus et al., 2013; Lin et al., 2015). The step change period occurred between 10:00 and 12:00. Data was collected every 10 seconds. The step change period was removed from the dataset.

Recovery Period. Each species was subjected to a recovery period in which Ca was returned to ambient. The recovery period occurred between 12:00 and 14:00. Data was collected every 10 seconds. The recovery period was removed from the dataset.

Free Stomata Model. Data was collected over the course of the growing season. Measurements were collected at 10:00, 12:00, and 14:00. Data was collected every 10 seconds. Records that contained negative values for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained values of 0 for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed.

Free Flux. Free flux data was collected over the course of the growing season. Measurements were collected at 10:00, 12:00, and 14:00. Data was collected every 10 seconds. Records that contained negative values for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained values of 0 for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. The data was collected and processed as part of the Free Flux project (Brookhaven National Laboratory, n.d.).

STOMATAL MODEL RESULTS

Comparison of Leaf gas exchange and Ball-Berry Model parameters

Figure 6. Leaf gas exchange and Ball-Berry Model parameters for Black Oak (*Quercus velutina*). Data was collected during the growing season. Measurements were collected at 10:00, 12:00, and 14:00. Data was collected every 10 seconds. Records that contained negative values for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained values of 0 for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed.

Figure 7. Leaf gas exchange and Ball-Berry Model parameters for Scarlet Oak (*Quercus coccinea*). Data was collected during the growing season. Measurements were collected at 10:00, 12:00, and 14:00. Data was collected every 10 seconds. Records that contained negative values for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained values of 0 for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed.

Investigation of a low m2 by effect

To investigate a low m2 value, data was collected for Scarlet Oak (*Quercus coccinea*). Data was collected during the growing season. Measurements were collected at 10:00, 12:00, and 14:00. Data was collected every 10 seconds. Records that contained negative values for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained values of 0 for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed.

Figure 8. Leaf gas exchange and Ball-Berry Model parameters for Scarlet Oak (*Quercus coccinea*). Data was collected during the growing season. Measurements were collected at 10:00, 12:00, and 14:00. Data was collected every 10 seconds. Records that contained negative values for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained values of 0 for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed.

Stomatal model validation

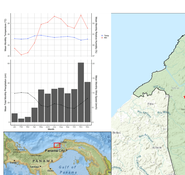

Figure 9. Stomatal model validation for Black Oak (*Quercus velutina*). Data was collected during the growing season. Measurements were collected at 10:00, 12:00, and 14:00. Data was collected every 10 seconds. Records that contained negative values for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained values of 0 for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed.

CANOPY LEVEL TRANSPIRATION SIMULATIONS

Stomatal model effect on transpiration

Figure 10. Stomatal model effect on transpiration. Data was collected during the growing season. Measurements were collected at 10:00, 12:00, and 14:00. Data was collected every 10 seconds. Records that contained negative values for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained values of 0 for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed.

Figure 11. Stomatal model effect on transpiration. Data was collected during the growing season. Measurements were collected at 10:00, 12:00, and 14:00. Data was collected every 10 seconds. Records that contained negative values for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained values of 0 for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed.

Forward model validation

Figure 12. Forward model validation for Black Oak (*Quercus velutina*). Data was collected during the growing season. Measurements were collected at 10:00, 12:00, and 14:00. Data was collected every 10 seconds. Records that contained negative values for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained values of 0 for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed.

CONCLUSIONS AND DISCUSSION

Summary of Key results

1. Measuring the step change response of stomata may be a more accurate way to predict stomatal response to increasing atmospheric CO2 concentrations.
2. Stomatal conductance (gs) is more sensitive to changes in Ca than previously thought.
3. Stomata may respond to changes in the rate of change of Ca, rather than absolute Ca (Eamus et al., 2013; Lin et al., 2015).
4. Stomata may respond to changes in the internal CO2 concentration (Ci) (Lin et al., 2015).
5. Changes in stomatal conductance can have significant impacts on transpiration, leaf temperature, and water use efficiency.
6. Some land models may overestimate the sensitivity of gs to increasing Ca (Lin et al., 2015).

Conclusions

1. Stomatal conductance is sensitive to short-term changes in Ca.
2. Land models may overestimate the sensitivity of gs to increasing Ca.
3. The mechanism used to sense Ca in stomata is currently unknown, several mechanisms have been proposed.

Figure 13. Summary of conclusions and impacts of this research.

Lack of significant p_i correlation

Data was collected during the growing season. Measurements were collected at 10:00, 12:00, and 14:00. Data was collected every 10 seconds. Records that contained negative values for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained values of 0 for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed.

Differences may between methods

Data was collected during the growing season. Measurements were collected at 10:00, 12:00, and 14:00. Data was collected every 10 seconds. Records that contained negative values for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained values of 0 for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed.

Sensitivities may by by-leaf effect

Data was collected during the growing season. Measurements were collected at 10:00, 12:00, and 14:00. Data was collected every 10 seconds. Records that contained negative values for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained values of 0 for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed.

Limits to methods due to application for canopy scale models

Data was collected during the growing season. Measurements were collected at 10:00, 12:00, and 14:00. Data was collected every 10 seconds. Records that contained negative values for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained values of 0 for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed.

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DISCLOSURES

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ABSTRACT

Traditional views of stomatal control of transpiration model CO2 sensing as a function of stomatal apertures during steady-state conditions. However, recent studies suggest that stomata may respond to changes in the rate of change of CO2 concentrations (ΔCa) rather than absolute Ca. Other studies suggest that stomata may respond to changes in the internal CO2 concentration (Ci). Ci is the CO2 concentration in the leaf airspace. Ci is the CO2 concentration at the site of carboxylation, specifically the region of the chloroplast where photosynthesis occurs. Ci strongly affects stomatal conductance.

We investigated the use of step changes in Ca to determine if stomatal conductance changes in response to Ca or Ci. Leaf gas exchange measurements were collected on six species in the Canopy Access Array (CAA) at Brookhaven National Laboratory (BNL) in Upton, New York. Each species was subjected to a step change in Ca (ΔCa) and a recovery period. Data was collected every 10 seconds. Data was collected during the growing season. Measurements were collected at 10:00, 12:00, and 14:00. Records that contained negative values for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed. Records that contained values of 0 for photosynthetic rate, stomatal conductance, and intercellular CO2 concentration were removed.

Preliminary results suggest that stomatal conductance is more sensitive to changes in Ca than previously thought. Stomatal conductance may respond to changes in the rate of change of Ca, rather than absolute Ca. Stomatal conductance may respond to changes in the internal CO2 concentration (Ci). Changes in stomatal conductance can have significant impacts on transpiration, leaf temperature, and water use efficiency. Some land models may overestimate the sensitivity of gs to increasing Ca.

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